AM-0595

CHEMICAL AND BIOLOGICAL CHARACTERIZATION OF A LOCALLY GROWN GANODERMA SPECIES

by

HOW KAM CHIONG

A DISSERTATION SUBMITTED TO THE INSTITUTE OF POSTGRADUATE STUDIES AND RESEARCH, UNIVERSITY OF MALAYA, KUALA LUMPUR

IN PARTIAL FULFILMENT OF THE REQUIREMENTS FOR THE DEGREE OF MASTER OF BIOTECHNOLOGY

NOVEMBER 1999





ACKNOWLEDGEMENTS

I thank both of my supervisors Dr. S. Vikineswary and Assoc. Prof. Dr. Chuah Cheng Hock for their support and guidance during this course of study. Without their consistent help and corrections, this study will not possible to be finished within six months.

I would like to thank Mr. Chong and Mr. Ong for teaching and helping me with some of the chemistry techniques during my stay in Chemistry Department. I would also like to acknowledge Cik Norzalida, Mr. Siew, Mr Chong and Puan Vijaya for their respective help on the NMR, MS, WDS and SEM units.

Many thanks to Tan, Ara, Getha, Faizah, Renu, Avenish, May Heng and the entire mycology Lab for helping me in their own ways during my study in IPSP.

Lastly, I would like to share my greatest pleasure in completing this thesis with my parents and Chen. Thanks for having faith in me.

ABSTRACT

Two locally grown *Ganoderma* species obtained from a mushroom farm in Semenyih, Malaysia were tentatively identified as *G. tsugae* and *G. lucidum* based on the morphology of basidiospores and cultural characteristics. Basidiospores of *G. tsugae* were 'rough' walled and had broad inter-wall pillars. Isolates of *G. tsugae* do not produce chlamydospores or primordia in culture and had an average growth rate of 6.8 mm/day at the optimal temperature range of 33-37°C.

In contrast, the basidiospores of G lucidum appeared 'smooth' walled, characterized by narrow, numerous inter-wall pillars. Isolates of G. lucidum produce chlamydospores and primordia in culture and had a higher average growth rate of 11.8 mm/day at a lower optimal temperature range between 28-33°C as compared to G. tsugae.

The fruit body of G. ssugae was investigated for its phytochemical content. Extraction followed by repeat chromatography of the hexane and methanol crude extracts resulted in the isolation of four compounds. The elucidation of structures were carried out by various spectroscopic methods.

Terpenoids content were found to be present in the highest levels, followed by alkaloids, saponins and flavonoids. Chemically guided fractionations based on TLC patterns of terpenoids allowed four compounds (GM 1.1, GM 2.2, GM 3.6.5 and

GM4.3) to be isolated from the hexane crude extract. These compounds were identified as stellasterol (GM 3.6.5), ergosterol (GM 4.3), di-(2-ethylhexyl) phthalate (DEHP) and a new brominated ergosta-type sterol (GM 1.1)

All the fractions collected from chromatography of both hexane and methanol crude, together with the four isolated compounds were used to test for antibacterial and antifungal activity. Using 6mm-paper disc diffusion method, all fractions showed weak (7.0 to 8.9 mm) to moderate (9.0 to 10.9 mm) activity against one or more of the test microorganisms. Candida albicans was the most susceptible organism, with 35% of the fractions showing moderate activity against this opportunistic pathogen.

DEHP showed a moderate broad-spectrum inhibition against the Grampositive bacteria, Bacillus cereus and Bacillus subtilis and two fungi, Candida
albicans and Schizosaccharomyces pombe. The other three ergosta-type sterols only
exhibited mild activity against one or more of the test microorganism. Based on the
results obtained, the occurrences of the isolated compounds were discussed.

ABSTRAK

Dua spesies Ganoderma tempatan yang diperolehi daripada sebuah ladang cendawan di Semenyih, Malaysia telah dikenalpasti secara tentatif sebagai Ganoderma tsugae and Ganoderma lucidum berdasarkan morfologi basidiospora dan ciri-ciri kultur.

Dinding basidiospora *G. tsugae* adalah 'kasar' dan mempunyai 'inter-wall pillar' yang lebar. Pencilan *G. tsugae* juga didapati tidak menghasilkan klamidospora atau 'primordia' dalam kultur dan mempunyai purata kadar pertumbuhan 6.8 mm/hari pada suhu optimum 33-37°C.

Sebaliknya, dinding basidiospora G. lucidum adalah 'licin' dan mempunyai 'inter-wall pillar' yang sempit. Kultur pencilan G. lucidum juga menghasilkan klamidospora dan 'primordia', dan mempunyai purata kadar pertumbuhan yang lebih tinggi, iaitu 11.8 mm/hari pada suhu optimum yang lebih rendah, iaitu 28-33°C berbanding dengan G. tsugae.

Jasad buah G. tsugae digunakan untuk pengajian kandungan fitokimia. Ekstraksi dengan heksana dan metanol, diikuti oleh kromatografi berulang, berjaya menyaringkan empat sebatian. Struktur keempat-empat sebatian ini dikenalpasti dengan kaedah-kaedah spektroskopi.

.

Kandungan 'terpenoids' adalah tertinggi diikuti 'alkaloids', 'saponins' dan 'flavonoids'. Empat sebatian yang disaringkan daripada ekstrak heksana dikenalpastikan sebagai 'sellasterol' (GM 3.6.5), 'ergosterol' (GM 4.3), 'di-(2-ethylhexyl) phthlate' (DEHP) dan satu sebatian baru yang merupakan 'brominated sterol'

Semua fraksi-fraksi dan kempat-empat sebatian yang disaringkan digunakan untuk menguji activiti antibakteria dan antikulat. Dengan menggunakan kaedah difusi kertas 6 mm, semua fraksi menunjukkan aktiviti lemah (7.0 – 8.9mm) sehingga sederhana (9.0 – 10.9 mm) terhadap satu atau lebih mikroorganisma yang diuji. Candida albicans merupakan organisma yang paling sensitif, dengan 35% daripada fraksi-fraksi menunjukkan activiti yang serdahana terhadap patogen oportunis ini.

DEHP pula menunjukkan aktiviti spektrum luas terhadap perencatan bacteria Gram-positif, Bacillus cereus dan Bacillus subtilis, dan dua kulat, Candida albicans dan Schizosaccharomycetes pombe. Tiga sterol yang lain pula hanya menunjukkan activiti yang lemah terhadap satu atau lebih mikroorganisma yang diujikan. Berdasarkan keputusan yang didapati, sebatian-sebatian yang wujud dibincangkan.

CONTENT

			PAGE
Abst Abst List List	rak of Figur of Table of Plates	res rs	
Char	oter One	2	
Intro	duction		1
Char	oter Two	2	
Liter	ature R	eview	
2.1	Origin	and Taxonomy of Ganoderma spp.	10
2.2	Pharm	nacologically Active Components in Ganoderma spp.	14
	2.2.1	Polysaccharides	16
	2.2.2	Bitter terpenoids	17
	2.2.3	Steroids	22
	2.2.4	Organic germanium	22
	2.2.5	Nucleotides	23
	2.2.6	Others	24
2.3	Triter	penoids Isolated from Ganoderma spp.	24
2.4	Analy	tical Techniques	34
Chap	ter Thr	<u>ee</u>	
Char	acteriza	tion of Two Locally Grown Ganoderma spp.	
3.1	Introdu	action	43
3.2	Materi	als and Methods	45

	3.2.1	Fruiting body collections and descriptions	45
	3.2.2	Microscopic studies of the basidiospore	45
	3.2.3	Cultural characteristics studies Isolation of pure culture Microscopic cultural studies Enzyme activity during growth Effect of temperature on growth rate	46 46 47 47
3.3	Resu	lts and Discussion	48
	3.3.1	Macromorphology of fruit body	48
	3.3.2	Microscopic studies of the basidiospore	51
	3.3.3	Macroscopic cultural studies	57
	3.3.4	Microscopic cultural studies	59
	3.3.5	Enzyme activity during growth	61
	3.3.6	Temperature studies	64
<u>Cha</u> j	oter Fou	<u>ır</u>	
Extr	action a	nd Isolation of Terpenoids	
4.1	Introd	luction	69
4.2	Mater	ials and Methods	71
	4.2.1	Fruit body	71
	4.2.2	Extraction of terpenoids	71
	4.2.3	Phytochemical tests Alkaloid test Terpenoid test Saponin test Flavonoid test	71 71 73 73 73
	4.2.4	Chromatographic separation of the crude extracts Column chromatography of hexane crude extract Column chromatography of methanol crude extract Thin layer chromatography (TLC)	73 = 76 76 77

	4.2.5	Instrumentation Mass spectrometry (MS) Nuclear magnetic resonance (NMR)	78 78 78
4.3	Resul	It and Discussion	79
	421	District and a second	
	4.3.1	Phytochemical tests	79
	4.3.2		80
		Isolation of Compound 1 (GM 1.1)	85
		Isolation of Compound 2 (GM 2.2) Isolation of Compound 3 (GM 3.6.5)	86 86
		Isolation of Compound 4 (GM 4.3)	87
	4.3.3	Characterization of isolation compounds	87
		Compound 2 (GM 2.2)	87
		Compound 4 (GM 4.3)	99
		Compound 3 (GM 3.6.5) Compound 1 (GM 1.1)	112 120
	ter Fiv		
Scree	ning fo	r Antibacterial and Antifungal Activity by Agar Diffus	ion Method
5.1	Introd	uction	132
5.2	Mater	ials and Methods	133
	5.2.1	Test bacteria	133
	5.2.2	Test fungal	134
	5.2.3	Screening for antibacterial and antifungal activity	134
5.3	Result	s and Discussion	135
	5.3.1	Activity of overall fractions	135
	5.3.2	Activity of isolated compounds	145
Chapt	ter Six		
Gener	al Disc	ussion & Conclusion	1473
Futur	e Work		153

Conclusion	155
References	156
Appendix	
Appendix I: Media and Reagents	167
Appendix II: Experimental and Statistical Data of Ganoderma spp.	168
Appendix III: ¹ H NMR of All the Fractions Collected from Column Chromatography of Hexane and Methanol Crude Extracts	174

LIST OF TABLES

Tables		Page
1.1	Pharmaceuticals developed from mushrooms in Japan	3
1.2	Pharmaceutical components of mushroom species	4
2.2	Pharmacologically active compounds and their medicinal benefits from the mushroom ${\it Ganoderma}$	15
2.3	The main classes of terpenoids	18
2.4	Triterpenoids isolated from Ganoderma spp.	36
3.1	Biometrics of basidiospores of two locally grown <i>Ganoderma</i> species.	51
3.2	Summary results of comparative studies on sample A and B	68
4.1	Solvent system and approximate volume used for elution of hexane crude	76
4.2	Solvent system and approximate volume used for elution of methanol crude	77
4.3	Phytochemical screening of the hexane and methanol crude extract of $G.\ tsugae.$	80
4.4	$R_{\rm f} values$ and colours obtained from TLC plates of Figure 4.4	81
4.5	$R_{\rm f} values$ and colours obtained from TLC plates of Figure 4.5	82
4.6	$R_{\rm f}$ values and colours obtained from TLC plates of Figure 4.6	83
4.7	$R_{\rm f} values$ and colours obtained from TLC plates of Figure 4.7	84
4.8	$R_{\rm f}$ values and colours of the isolated pure compounds.	85
4.9	$^{l}\text{H NMR}$ (400 MHz) spectral data for DEHP (GM 2.2) in CDCl $_{3}$	93
4.10	¹³ C NMR (400 MHz) and DEPT spectral data for DEHP (GM2.2) in CDCl ₃	93

4.11	¹ H NMR (400 MHz) spectral data for ergosterol (GM 4.3) in CDCl ₃ .	104
4.12	$^{13}\text{C NMR}$ (400 MHz) and DEPT spectral data for ergosterol (GM 4.3) in CDCI $_3$.	107
4.13	Partial ^{1}H chemical shifts of stellasterol (GM 3.6.5) in CDCl ₃	119
4.14	$^{13}\mathrm{C}$ NMR (400 MHz) and DEPT spectral data for stellasterol (GM 3.6.5) in CDCl ₃ .	119
5.1	Significance of the test microorganisms	133
5.2	Antibacterial activity of the fractions from G. tsugae	136
5.3	Antifungal activity of the fractions from G. tsugae	138
5.4	Fractions that show moderate inhibition (i.e. inhibition zones of 9 mm and above) to one or more of the test organism.	141
5.5	Grouping of the fractions that showed moderate inhibition	141

LIST OF FIGURES

Figures		Page
2.1	A generalized scheme of the life cycle of Basidiomycetes	11
2.2	Lanostane triterpenoid skeleton	18
2.3	Ganoderal A	20
2.4	Ganodermic acids R and S	21
2.5	C ₂₇ and C ₃₀ triterpenoids	25
2.6	Fruit body type (type I) and mycelium type (type III) of ganoderic acids.	26
2.7	Isolation of Ganoderic acids A and B by Kubota and Asaka (1982)	27
2.8	Isolation procedures adopted by Hirotani et al. (1985) and Hirotani and Furuya (1986).	29
2.9	Isolation procedures adopted of by Kikuchi et al. (1985 a, b; 1986a, b).	30
2.10	Isolation procedures adopted by Nishitoba et al. (1984; 1985a,b,c).	32
2.11	Lucidenic acid D	31
3.1	The growth rate of G . $tsugae$ (Gt) and G . $lucidum$ (Gl) at different temperatures.	64
4.1	General scheme for the extraction of terpenoids from <i>G. tsugae</i> .	72
4.2	Detailed flow diagram of the isolation of terpenoids from hexane crude extract.	74
4.3	Detailed flow diagram of the isolation of terpenoids from methanol crude extract.	75
4.4	TLC patterns obtained from chromatographic separation of GM1 (Fr 3-6) and GM 2 (Fr 10-11)	81

4.5	TLC patterns obtained from chromatographic separation of GM 3 (Fr 12-13).	82
4.6	TLC patterns obtained from chromatographic separation of GM 4 (Fr 14-15) and GM 5 (Fr16)	83
4.7	TLC patterns obtained from chromatographic separation of methanol crude extract	84
4.8	EIMS spectrum of DEHP (GM 2.2)	88
4.9	¹ H NMR spectrum of DEHP (GM 2.2)	90
4.10	¹³ C NMR spectrum of DEHP (GM 2.2)	91
4.11	DEPT spectrum of DEHP (GM 2.2)	92
4.12	COSY spectrum of DEHP (GM 2.2)	95
4.13	HETCHOR spectrum of DEHP (GM 2.2)	96
4.14	EIMS spectrum of ergosterol (GM 4.3)	100
4.15a	¹ H NMR spectrum of ergosterol (GM 4.3)	102
4.15b	Expanded section of ¹ H NMR spectrum of ergosterol (GM 4.3)	103
4.16	¹³ C NMR spectrum of ergosterol (GM 4.3)	105
4.17	DEPT spectrum of ergosterol (GM 4.3)	106
4.18a	HETCHOR spectrum of ergosterol (GM 4.3)	109
4.18b	HETCHOR spectrum of ergosterol (GM 4.3)	110
4.19	COSY spectrum of ergosterol (GM 4.3)	111
4.20a	EIMS spectrum of stellasterol (GM 3.6.5)	114
4.20b	EIMS spectrum of stellasterol (GM3.6.5) (expanded)	115
4.21	¹ H NMR spectrum of stellasterol (GM 3.6.5)	116
4.22	¹³ C NMR spectrum of stellasterol (GM 3.6.5)	117
4.23	DEPT spectrum of stellasterol (GM 3.6.5)	118

4.25a	EIMS spectrum of compound 2 (GM 1.1)	121
4.25b	EIMS spectrum of compound $\mathfrak{Z}(GM \ 1.1)$ (expanded)	122
4.25c	Confirmation of EIMS spectrum of compound 1 (GM 1.1) (repeat run)	123
4.26	^{1}H NMR spectrum of compound 2 (GM 1.1)	125
4.27	¹³ C NMR spectrum of compound 1 (GM 1.1)	126
4.28a	DEPT spectrum of compound \mathfrak{Z} (GM 1.1)	127
4.28b	DEPT spectrum of compound $\mathfrak{L}(GM 1.1)$	128
4.28c	DEPT spectrum of $compound \underbrace{1}_{c}(GM 1.1)$ (expanded)	129
4.29	Proposed structure of compound $\widetilde{\underline{1}}$	130
4.30	Indication of bromine in fresh substrate by wave dispersion spectrum.	131
6.1	Synthesis of ergosterol in fungi	151

LIST OF PLATES

Plates	3	Page
1	a: Rubber wood sawdust (substrate) b: Mixing of sawdust with rice bran and calcium carbonate	6
	c: Substrate is packed into plastic bags d: The bags are sterilized and e: inoculated with quality mushroom mycelium culture (the koji) f: Bags are kept in a dark shed with average temperature of 29-32 °C for spawn running	7
	g: Spawn running (mycelium growth in solid substrate) h: Appearance of young fruit body (primordia) i: Mature fruit body	8
2	Two Ganoderma species collected from a mushroom farm in Semenyih. i. Sample A was claimed to be G. tsugae ii. Sample B was claimed as G. hucidum. (a) Front view and (b) back view.	49
3	The context colour and pore size of the <i>Ganoderma</i> species. (a) Sample A appeared white and smooth with no visible pore and (b) sample B appeared grayish brown with visible round to angular pore.	50
1	Light microscopy of basidiospores of the <i>Ganoderma</i> species. Basidiospores were brown and ovate. (a) Inter-wall pillars (arrow) of sample A are well defined. (b) Inter-wall pillars (arrow) of Sample B are not as apparent as basidiospores of sample A.	52
5	Scanning electron microscopy of the basidiospores. (a) The attachment of the spore to the germ tubes in sample A. (b) Mature spores with characteristic germ tubes observed in sample B.	53
5	Scanning electron microscopy of the Ganoderma species (a), (b), (c): Basidiospores of sample A (tentatively identified as G. Isugae). Outer wall surface with prominent depressions (arrow) and the fractured basidiospore exposing large, inter-wall pillars (arrow). (d), (e), (f): Basidiospores of sample B (tentatively identified as G. Iucidum). Outer wall surface with shallow depressions (arrow) and the fractured basidiospore exposing small, numerous inter-wall pillars (arrow).	56

7 Appearance of the mycelium mat of Ganoderma spp. after five weeks 58 of incubation at room temperature. (a) i. No production of primordia in G. tsugae, while ii. primordia was observed in G. lucidum. (b) and (c) primordia of G. lucidum (close-up). 8 Microscopic characters of cultures of Ganoderma species observed 60 under light microscope. (a) Thin-walled hyphae, with consistently nodose-septate (400x). (b) i. Fiber hyphae and ii. cuticular cells (1000x). (c) clamp connections via anastomosis (1000x). Chlamydospores observed only in G. lucidum under light microscope. 9 62 (a) Intact chlamydospores (b) Detached chlamydospore. 10 Production of extracellular enzymes by the Ganoderma species. 63 Production of polyphenol oxidase by G. tsugae, ii. Production of polyphenol oxidase by G. lucidum 11 Linear growth of the Ganoderma spp. after 7 days at a range of 65

temperature on MEA plates. (a) Linear growth of *G. tsugae* at the following temperatures (upper row, left to right) 17, 22, 25, 28, (bottom row, left to right) 30, 33, 37, 40°C. (b) Linear growth of *G. lucidum* at the following temperatures (upper row, left to right) 17, 22, 25, 28, (bottom row, left to right) 30, 33, 37, 40°C.

LIST OF ABBREVIATIONS

% percentage aqueous

aq

CDCL₃ deuterated chloroform

CHCl₁ chloroform conc concentrated

Correlation Spectroscopy (homonuclear ¹H-¹H correlation spectroscopy) COSY

d doublet

dd double doublet

Distortionless Enhancement by Polarisation Transfer DEPT EIMS electron impact mass spectrum

HETCHOR Heteronuclear Correlation

Hex Hexane

m multiplet

MS Mass Spectroscopy mass/ charge m/z

MeOH methanol milligram mg MHz megahertz

NMR Nuclear Magnetic Resonance NOE Nuclear Overhauser Effect

q quartet singlet

s TLC Thin Layer Chromatography

trimethylsilane TMS vol volume

weight/ weight w/w