

## APPENDIX

### BUFFERS AND SOLUTION FOR DNA EXTRACTION FROM FROZEN CERVICAL TISSUE, PAP SMEARS AND CELL CULTURE

#### *Phosphate buffered saline (PBS), pH 7.2*

Sodium phosphate (anhydrous)	1.52 g
Potassium dihydrogen phosphate	0.58 g
NaCl	8.50 g
dH <sub>2</sub> O	made up to 1 liter

#### *20X SSC, pH 7.0*

NaCl	175.3 g
Sodium Citrate trihydrate	88.2 g
dH <sub>2</sub> O	made up to 1 liter

#### *1M Tris - HCL, pH 8.0*

Trizma base	121.1g
dH <sub>2</sub> O	made up to 1 liter

The pH was adjusted with concentrated HCL

#### *SSC-buffered phenol*

Phenol was melted at 68°C. An antioxidant, 8-hydroxyquinoline was added to a final concentration of 0.1%. The melted phenol was then extracted with with equal volume of 1 X SSC, pH 8 by stirring at 4°C, overnight, until the pH of the aqueous phase is more than 7.6. The SSC buffered phenol was stored in a dark bottle at 4°C for up to 1 month.

### CELL CULTURE

All media were filter sterilized and stored at 4°C.

#### *RPMI 1640 base medium*

RPMI 1640 base media	1 sachet
Sodium bicarbonate	2.0 g
Hydroxyethylpiperazine ethanesulfonic Acid (HEPES)	1.0 g
dH <sub>2</sub> O	made up to 1 liter

### **Reviving media**

RPMI base media	77 ml	
Fetal calf serum (FCS)		20 ml
Penicillin	2 ml	
Kanamycin	1 ml	
L - glutamine	0.012 g	

Media was filtered sterilized with 0.22  $\mu$ m milipore filter

### **Culture Media**

Preparation of culture media was the same as reviving media with the addition of 10% FCS.

### **Freezing solution**

FCS	10 ml
Dimethyl sulfoxide (DMSD)	4 ml
RPMI 1640 base medium	6 ml

The solution was prepared fresh and sterilized with 0.22 $\mu$ m millipore filter.

### **GEL ELECTROPHORESIS**

#### **Tris-borate EDTA (TBE) 5X**

Trizma base	54g
Boric acid	27.5 g
EDTA 0.05M pH 8.0	20 g
dH <sub>2</sub> O	made up to 1 liter

#### **Loading Buffer 6 X**

Bromophenol blue	0.25%
Sucrose	40% (w/r)
dH <sub>2</sub> O	made up to 100 ml

20x SSC pH 7.0

### **SOUTHERN BLOT HYBRIDIZATION**

Solutions are prepared as described by Boehringer Mannheim (1989). Reagents used were from the Non-radioactive DNA labelling and Detection kit.

### **Hybridization solution**

5X SSC	22.5 ml
N-lauroylsarcosine, Na-salt (Sigma)	0.1% (w/v)
SDS	0.02% (w/v)
Blocking reagent (vial 11)	1% (w/v)

### **ISOLATION OF PLASMID**

#### **LB broth**

Bacto-Tryptone	10 g
Bacto-yeast extract	5 g
NaCl	10 g
ddH <sub>2</sub> O	up to 1 litre

### **IN SITU HYBRIDIZATION**

#### **Denhardt's solution (50x)**

Ficoll 400	5 g
Polyvinylpyrrolidone-360	5 g
BSA	5 g
dH <sub>2</sub> O	added to 500 ml

The solution was sterilized using the disposable 0.22  $\mu$ m Nalgene filter, dispensed into 25 ml aliquots and stored at -20°C.

#### **4% paraformaldehyde**

Paraformaldehyde	4 g
PBS	100 g

Paraformaldehyde was dissolved by heating the solution 60°C-70°C and by adding drops of 2 N NaOH while adjusting the pH to 7.2.

#### **Phosphate Buffered Saline (PBS) pH 7.6**

Sodium chloride	7.75 g
Potassium phosphate dibasic	1.5 g
Potassium phosphate, monobasic	0.2 g
dH <sub>2</sub> O	to 1 litre of the solution

The pH of the solution was adjusted to 7.6 with 3 M NaOH or 8.5% phosphoric acid.

### **Prehybridization mixture**

50x Denhardt's solution	0.2 ml
50% dextran sulphate	1.0 ml
Sonicated salmon sperm DNA (10 mg/ml)	0.2 ml
20X SSC	2.0 ml
Formamide	5.0 ml
Sterile dH <sub>2</sub> O	added to 10 ml

### **Hybridization solution (probe-cocktail)**

50x Denhardt's solution	0.2 ml
50% dextran sulphate	1.0 ml
Sonicated salmon sperm DNA (10mg/ml)	0.2 ml
20X SSC	2.0 ml
100% deionized formamide	5.0 ml
Digoxigenin labelled probe	500 ng/slide
Sterile dH <sub>2</sub> O	to 10 ml

### **100% deionized formamide**

Formamide	100 ml
Resin	10 g

The mixture was stirred vigorously for 30 minutes, filtered twice through Whatman filter paper and stored at -20°C.

## **IMMUNOSTAINING**

The LSAB kit (Dako) contained the following reagents :-

1. Biotinylated anti-mouse and anti-rabbit immunoglobulin in 0.05 M Tris-HCL buffer, pH 7.6.
2. Streptavidin-HRP conjugate in 0.05 M Tris-HCL buffer pH 7.6.

AEC substrate system (K697,Dako) contains the following reagents:-

1. 0.1 M Acetate buffer, pH 5.2
2. 3% Hydrogen peroxide in water
3. 3% 3-amino-9-ethylcarbazote (AEC) in N,N dimethylformamide.

## **ENZYME-LINKED IMMUNOSORBENT ASSAY (ELISA)**

### **0.1 M Carbonate buffer, pH 9.6**

Na <sub>2</sub> CO <sub>3</sub>	1.65 g
NaHCO <sub>3</sub>	3.14 g
dH <sub>2</sub> O	up to 500 ml

### **Phosphate buffered Saline (PBS), pH 7.3**

Sodium chloride	8.0
Potassium phosphate monobasic	0.2
Sodium phosphate dibasic	1.15
Potassium chloride	0.2
dH <sub>2</sub> O	up to 1 litre

PBS-0.05% Tween 20 (PBS-T) was prepared by adding 0.05% of Tween 20 to the above solution.

### **Lamb serum**

Lamb serum (Gibco) was removed from the freezer and thawed at 37°C after which it was heat inactivated at 56°C for 30 minutes. The serum was then filtered through Whatman filter paper and 45 µm membrane filter. It was dispensed into aliquots and stored at -20°C.

### **ABTS (50x) (20mg/ml)**

(2,2-azinodi-(3-ethylbenz thiazolin sulfate(Boeringer)	0.2 g
dH <sub>2</sub> O	10 ml

### **0.1 M Citrate Buffer, pH 4.0**

Citric acid monohydrate	6.62 g
Sodium citrate dihydrate	5.44 g
dH <sub>2</sub> O	to 1 litre

### **Peroxidase substrate**

50X ABTS	150 µl
30% Hydrogen peroxidase	1.8 µl
0.1M Citrate buffer	up to 6 ml

The solution was prepared fresh.

### **Diethanolamine buffer - pH 9.8**

Diethanolamine	97 ml
Magnesium chloride	0.1 g
dH <sub>2</sub> O	up to 1 litre

The solution was filtered through Whatman paper and stored in a dark bottle at room temperature.

### **Phosphatase substrate**

Phosphatase substrate 104 (Sigma)	0.1 g
Diethanolamine buffer	100 ml

**20% glucose**

Glucose	10 g
Sterile dH <sub>2</sub> O	to 50 ml

The solution was transferred to a 50 ml tube and rotated end-over-end overnight at room temperature and stored at 4°C.

**0.2 M NaH<sub>2</sub>PO<sub>4</sub>**

NaH <sub>2</sub> PO <sub>4</sub>	27.8 g
dH <sub>2</sub> O	to 1 litre

**0.2 M Na<sub>2</sub>HPO<sub>4</sub>**

Na <sub>2</sub> HPO <sub>4</sub>	53.65 g
dH <sub>2</sub> O	to 1 litre

**0.1 M Phosphate buffer, pH 6**

0.2 M NaH <sub>2</sub> PO <sub>4</sub>	87.7
0.2 M Na <sub>2</sub> HPO <sub>4</sub>	12.3
dH <sub>2</sub> O	to 100 ml