#### 5.0 RESULTS AND DISCUSSIONS

### 5.1 DNA extraction

#### 5.1.1 Quality comparisons

DNA from Sargassum oligocystum and S. binderi collected from the intertidal coral reef flat bed at Cape Rachado and Teluk Kemang, Port Dickson, Negeri Sembilan was isolated by using two protocols, both involving the use of CTAB. Quality of DNA isolated from protocol 1 and protocol 2 is shown in Table 6. Protocol 2 was chosen for extraction of intact DNA as this protocol is simple to handle, safe, inexpensive and less time consuming as compared to protocol 1. A higher purity of DNA and no pigmentation was obtained using protocol 2. The A<sub>260/280</sub> ratios of the DNA extracted using protocol 2 are shown in Table 7. The value of a pure DNA is shown by the A<sub>260/280</sub> ratio in the range 1.800 to 2.000. However, when working with the DNA of plants, the presence of the secondary compounds such as phenolic compounds and polysaccharides will somehow give a lower value of A<sub>260/280</sub> ratios. As in this study, the value of A<sub>260/280</sub> ratios is in between 1.642 and 1.956. Although the A<sub>260/280</sub> ratios is slightly lower than expected, the amplifications of PCR using random primers were still giving a positive result.

A total of 60 samples were extracted by using protocol 2. Clear, non-pigmented intact DNA was spooled out by using a glass rod after precipitation by isopropanol. The A260/280 ratio of the first extraction ranged from 1.114 to 1.625 (Table 7). In order to obtain a successful amplification of RAPD-PCR, the extraction was carried out once again to get purer DNA with a higher A260/280 ratio (Table 8). The genomic DNA bands isolated from protocol 1 and 2 (first and second extraction) is shown in Figure 2. When tested with RAPD-PCR, 95% of the samples gave positive results compared with 55% for the DNA without re-extraction.

S. oligocystum and S. binderi need extra attention while cleaning as they are normally dirty. The plant may be full of sand, salt and epiphytised with diatoms, fungi and bacteria. The stem of the plants was too difficult to grind into powder form due to the stiffness of the cell wall thickening and polysaccharide deposition. The receptacles of the plants were always full of epiphytes and the probality to get them clean is very low and the structure of the receptacles themselves were very small and the surface were not even. Therefore, only leaves and young shoots were used as these parts are easy to clean. More DNA can be obtained from young shoots as these parts are actively involved in mitosis.

Thoroughly clean tissues were ground using liquid nitrogen to break the tough cell walls in order to release the cellular constituents. The cell membrane was then disrupted by using a detergent, usually PVP (polyvinylpropaline), SDS (sodium dodecyl sulfate) or CTAB so that the DNA is released into the extraction buffer. The DNA was protected from endogenous nucleases by adding EDTA as a chelating agent that binds magnesium ions, generally considered a necessary cofactor for most nucleases. To denature and separate the proteins from the DNA, emulsified homogenate of chloroform: isoamylalcohol (24:1) was added (Rogers and Bendich, 1994).

In protocol 1, according to Doyle and Doyle (1990), 1% (w/v) of PVP-40 added into DNA isolation buffer to isolate DNA from plants was found to be effective as an absorbent for tannin and other secondary plant compounds, including high concentration of phenolic compounds. However, this was not good enough to get rid of the polysaccharides and phenolic compounds in S. oligocystum and S. binderi. A "jelly-like" and insoluble DNA preparation was obtained and this is most probably due to the carbohydrates as only 1% CTAB was used initially, which was insufficient. 10 % of CTAB used after the first chloroform-isoamyalcohol extraction generated significant amounts of hazardous waste that will endanger the environment as the disposal of hazardous waste will add to the actual extraction costs.

The described procedure in protocol 2 is reliable, relatively quick and simple to perform. This method works very well with small quantities of fresh algal tissues as compared to protocol 1 which required a large amount of dried tissues. Also, in protocol 1 DNA is lost at each step, thus reducing the overall efficiency of the

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isolation as too many steps were involved. Thus, a rather simple and quick procedure was developed in protocol 2. By incubating tissue powder with 4% CTAB in the extraction buffer, nucleic acids are precipitated selectively despite the presence of phenolic compounds, chlorophyll and cell proteins. Insoluble polysaccharides are precipitated by hot CTAB (Murray and Thompson, 1980 and Moller, 1992). As a replacement for 1% PVP, 0.2% (w/v) of 2-β-mercaptoethanol was used in order to absorb secondary plant compounds including the phenolic compounds. The polysaccharides can be removed from isolated DNA by adjusting the salt concentration from 0.7 M to 1.4 M (Michaels et al., 1994).

0.830-1.125	10 0.830-1.
1.114-1.625	60 1.

Table 6: Comparison of DNA extraction methods

DNA	A <sub>260/280</sub>	Amount of DNA	DNA	A <sub>260/280</sub>	Amount of DNA
samples	ratio	(µg/ml)	samples	ratio	(µg/ml)
SO 01	1.556	28.0	SB 01	1.262	26.5
SO 02	1.192	28.0	SB 02	1.268	26.0
SO 03	1.225	24.5	SB 03	1.281	32.0
SO 04	1.625	47.0	SB 04	1.176	40.0
SO 05	1.197	42.5	SB 05	1.229	43.0
SO 06	1.217	36.5	SB 06	1.233	45.0
SO 07	1.128	48.5	SB 07	1.282	32.0
SO 08	1.154	45.0	SB 08	1.254	44.5
SO 09	1.158	44.0	SB 09	1.241	54.0
SO 10	1.151	46.0	SB 10	1.265	43.0
SO 11	1.158	44.0	SB 11	1.329	54.5
SO 12	1.152	45.5	SB 12	1.267	38.0
SO 13	1.156	44.5	SB 13	1.243	43.5
SO 14	1.114	49.0	SB 14	1.214	62.5
SO 15	1.161	44.0	SB 15	1.202	59.5
SO 16	1.188	67.0	SB 16	1.296	35.0
SO 17	1,356	30.5	SB 17	1.308	51.0
SO 18	1.464	50.5	SB 18	1.277	60.0
SO 19	1.476	46.5	SB 19	1.297	41.5
SO 20	1.288	71.5	SB 20	1.247	48.0
SO 21	1.409	58.5	SB 21	1.264	57.5
SO 22	1.444	65.0	SB 22	1.327	36.5
SO 23	1.401	38.5	SB 23	1.204	59.0
SO 24	1.517	69.5	SB 24	1.227	54.0
SO 25	1.367	30.0	SB 25	1.231	91.0
SO 26	1.261	84.5	SB 26	1.187	42.5
SO 27	1.337	69.5	SB 27	1.269	87.0
SO 28	1.261	68.0	SB 28	1.344	40.5
SO 29	1.258	63.5	SB 29	1.303	76.0
SO 30	1.185	65.0	SB 30	1.512	65.0

SO: Sargassum oligocystum

SB: Sargassum binderi

Table 7: DNA quality of protocol 2 (first extraction)

DNA	A <sub>260/280</sub>	Amount of DNA	DNA	A <sub>260/280</sub>	Amount of DNA
samples	ratio	(µg/ml)	samples	ratio	(μg/ml)
SO 01	1.722	8.0	SB 01	1.940	8.0
SO 02	1.956	7.5	SB 02	1.866	8.0
SO 03	1.642	6.0	SB 03	1.778	10.0
SO 04	1.855	15.0	SB 04	1.684	12.0
SO 05	1.773	10.0	SB 05	1.750	15.0
SO 06	1.655	9.0	SB 06	1.850	15.0
SO 07	1.922	12.0	SB 07	1.865	12.0
SO 08	1.930	12.0	SB 08	1.920	13.5
SO 09	1.885	12.0	SB 09	1.690	18.0
SO 10	1.722	15.0	SB 10	1.725	10.5
SO 11	1.763	15.0	SB 11	1.750	14.0
SO 12	1.695	15.0	SB 12	1.859	12.5
SO 13	1.880	12.0	SB 13	1.880	15.0
SO 14	1.900	18.0	SB 14	1.900	18.0
SO 15	1.778	12.0	SB 15	1.925	15.0
SO 16	1.748	25.0	SB 16	1.910	10.0
SO 17	1.802	12.0	SB 17	1,880	17.5
SO 18	1.667	20.0	SB 18	1.875	20.0
SO 19	1.890	15.0	SB 19	1.790	15.0
SO 20	1.920	28.0	SB 20	1.800	15.0
SO 21	1.960	25.0	SB 21	1.800	25.0
SO 22	1.918	28.0	SB 22	1.836	12.0
SO 23	1.899	12.0	SB 23	1.720	25.0
SO 24	1.793	25.0	SB 24	1.900	25.0
SO 25	1.699	8.5	SB 25	1.930	33.0
SO 26	1.800	30.0	SB 26	1.800	15.0
SO 27	1.825	25.0	SB 27	1.850	30.0
SO 28	1.927	20.0	SB 28	1.750	15.0
SO 29	1.880	20.0	SB 29	1.875	28.0
SO 30	1.900	20.0	SB 30	1.790	25.0

SO: Sargassum oligocystum

SB: Sargassum binderi

Table 8: DNA quality of protocol 2 (re-extraction)

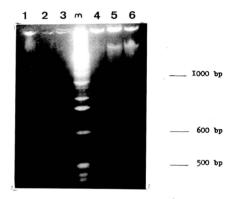


Figure 2. Example of Sargassum oligocystum genomic DNA on 0.8% agarose gel.
Lanes 1 and 2, Protocol 1;
Lanes 3 and 4, Protocol 2 (single extraction)
Lanes 5 and 6, Protocol 2 (double extraction).
The lane marked with m is a 100-bp DNA ladder (from 1500 to 100 bp, respectively).

## 5.2 Optimisation of PCR

### 5.2.1 PCR parameters

To optimise the PCR conditions with various RAPD primers is a laborious task. Each PCR parameter has to be considered in order to obtain stable and reliable band patterns. The parameters of RAPD-PCR amplification include Mg<sup>2+</sup> concentration, amount of dNTP, amount of primer, DNA concentration and annealing temperature, number of cycles, and most importantly, the amount of *Taq* polymerase, which plays a very important role in successful amplification.

## 5.2.2 Optimised conditions

From the study, five primers (OPA13, OPK7 and 9, OPN6 and 16) were examined with the two *Sargassum* species: *S. oligocystum* and *S. binderi*. For the optimisation of the RAPD-PCR, seven parameters were optimised. The DNA with the concentration of 25 ng, 50 ng and 100 ng were tested. The primers with a different concentrations of 5 pmol and 10 pmol were tested. 100 µM and 200 µM of the dNTP concentrations were also tested. For the *Taq* DNA polymerase, one and two units were tested. The concentrations of 1.5 mM, 2.0 mM and 2.5 mM of MgCl<sub>2</sub> were tested. The number of cycles for the amplification of 30, 35, 40, 45, 50, 55 and 60 cycles were tested. The annealing temperature of the reactions were tested at 30°C,

33°C, 36°C, 40°C and 43°C. A combination of the optimised conditions for each primer with the PCR parameters are shown in Table 9. With the optimisation of the reaction conditions and parameters, the variability of the replicon was reduced.

For the optimisation of the number of cycles, fewer than 30 thermal cycles provided insufficient amplification, while more than 55 cycles did not significantly increase yield. Magnesium concentrations below 1.5 mM, primer concentrations below 5 pmole, and individual dNTP concentrations below 100 µM failed to yield sufficient amplification.

Among the parameters tested, Mg<sup>2+</sup> concentration was found to be very important for reproducibility of the method. At lower concentrations (1.0-1.5 mM), only lower molecular weight bands were amplified (Figure 3, a and b, lane 1-5). As the concentration was increased to 2.0 -2.5 mM, the banding patterns were consistent with both high and low molecular weight bands (Figure 3, a and b, lane 6-10). At a concentration of 2.5 mM, the higher molecular weight bands were more intense and sharp but the bands of lower molecular weight were very faint (Figure 3, a and b, lane 6-10). Therefore, 2.5 mM of Mg<sup>2+</sup> concentration was chosen for the rest of the RAPD-PCR amplification.

In this study, not only the Mg<sup>2+</sup> concentration (Figure 3) but annealing temperature (Figure 4) should be optimised for each species and primer combinations separately according to the suggestion of Wolff et al. (1993). Differences in results were noted

with various amounts of *Taq* polymerase (Figure 5), dNTP used (Figure 6), primer (Figure 7) and DNA (Figure 8) used. Therefore, optimisation of each parameter was carried out to obtain the most suitable conditions to gain the consistent and sharpest bands in the amplification.

Amplification of target DNA loci occurred at all tested *Taq* polymerase concentrations. The yield and complexity of amplified products were relatively low at 0.5 and 1 unit of polymerase per 25 µl reaction. As the quantity of enzyme was increased to 2 units per reaction, many additional bands, both prominent and faint, became evident (Figure 5). This result suggests a competition between loci for a limiting amount of enzyme. Presumably, some loci are bound by one or two sites that are less perfectly complementary to the primer that are bound to other sites. Loci bound by these former sites are at a disadvantage when polymerase is limiting and are efficiently amplified only with an excess of enzyme.

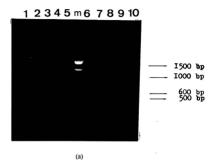
If secondary bands are produced under a condition of excess enzyme, then the addition of sufficient DNA should overcome this condition. When 2.0 unit of polymerase and varying quantities of DNA were used in standard amplifications (Figure 8), the secondary band formation observed with 25 ng of DNA was suppressed with 50 ng. Primer concentrations of 10 pmole enhanced the production of some bands (Figure 7) but did not improve the discrimination of major bands from the background signal.

Significant changes in the DNA fingerprint generated are not related to the annealing temperature as in the study carried out by Welsh and McClleland, 1990. The above statement did not apply to this study where the annealing temperature affected the amplified RAPD-PCR products (Figure 4), that is, changes in annealing temperature affected the number of bands produced.

From this study, it is clear that the RAPD technique requires careful attention to detail in execution and cautious interpretation only when sufficient replication has been performed. The plant material should be clean and as free as possible of contaminating organisms. Extreme care must be taken during the preparation of reagents and amplification reaction mixtures to prevent contamination or DNA carryover from one vessel to another so as to reduce the possibility of false positives or false negatives (Kwok, 1989). Variation in efficiency of amplification among heating block wells in the temperature cycler can lead to variable results (Linz, 1990). Therefore, it is necessary to determine the temperature of each well, both in preliminary experiment and from experiment to experiment.

Primer Optimised PCR parameter	OPA 13	OPK 7	OPK 9	OPN 6	OPN 16
Mg <sup>2+</sup>	2.5 mM	2.5 mM	2.5 mM	2.5 mM	2.5 mM
Taq polymerase	1 unit	2 unit	2 unit	2 unit	2 unit
dNTP	100 μΜ	200 μΜ	200 μΜ	200 μΜ	200 μΜ
primer	5 pmol	5 pmol	10 ρmol	10 pmol	10 pmol
annealing temperature	40°C	30°C	30°C	36°C	33°C
DNA concentration	25 ng	50 ng	50 ng	50 ng	50 ng

Table 9: Optimised conditions for various randomised primers



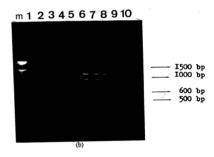


Figure 3. The effect of  $Mg^{2*}$  concentration on RAPD amplification of Sargassum oligocystum SO17 by OPN16 . a. Lane 1-5, 1.0 mM  $Mg^{2*}$ ; Lane 6-10, 2.0 mM  $Mg^{2*}$ ; b. Lane 1-5, 1.5 mM  $Mg^{2*}$ ; Lane 6-10, 2.5 mM  $Mg^{2*}$ . The lane marked with m is a 100 bp DNA ladder (from 1500 bp to 100 bp, respectively).

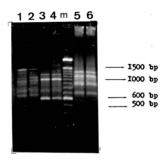


Figure 4. The effect of annealing temperature on RAPD amplification of Sargassum oligocystum SO25 by OPA13. Lane 1 and 2, 30°C Lane 3 and 4, 36°C Lane 5 and 6, 40°C Lane 5 and 6, 40°C tane 5 and 6, 40°C Lane 5 one depend on the same marked with m is a 100 bp DNA ladder (from 1500 bp to 100 bp, respectively).

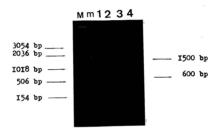


Figure 5. The effect of Taq polymerase on RAPD amplification of Sargassum oligocystum SO11 by OPK7. Lane 1 and 3, 1 unit of Taq polymerase; Lane 2 and 4, 2 unit of Taq polymerase. The lanes marked with m is a 100 bp DNA ladder (from 1500 bp to 100 bp) and M is a 1 kb DNA ladder (from 5095 bp to 154 bp), respectively.



Figure 6. The effect of dNTP concentration on RAPD amplification of Sargassum of Socyossum SO9 by OPK7. Lane 1-3, 100 µmole of dNTP.
Lane 4-6, 200 µmole of dNTP.
The lanes marked with m is a 100 bp DNA ladder (from 1500 bp to 100 bp) and M is a 1 kb DNA ladder (from 5095 bp to 154 bp), respectively.

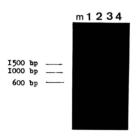


Figure 7. The effect of primer concentration on RAPD amplification of Sargassum aligocystum SO20 by OPK7. Lane 1 and 2, 5 pumble of primer; Lane 3 and 4, 10 pmole of primer. The lane marked with m is a 100 bp DNA ladder (from 1500 bp to 100 bp), respectively.

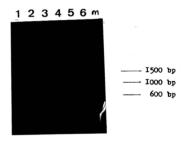


Figure 8. The effect of DNA concentration on RAPD amplification of Surgassum oligocystum SO12 by OPK7. Lane 1-2, 25 ng of genomic DNA; Lane 3-4, 50 ng of genomic DNA; Lane 5-6, 100 ng of genomic DNA. The lane marked with m is a 100 bp DNA ladder (from 1500 bp to 100 bp), respectively.

## 5.3 RAPD analysis

RAPD-PCR has been used in recognition of animal species (Fong et al., 1995; Pfenninger et al., 1995). In higher plants, this technique has been used for the identification, that is, Apium (Yang and Quiros, 1993), Solamum (Singsit and Ozias-Akins, 1993), Stylosanthes (Kazan et al., 1993), Theobroma (Wilde et al., 1992) and Brassica (Hu and Quiros, 1991). Application in algae such as heterosis study in Gelidium vagum (Rhodophyta) (Patwary et al., 1994), gene mapping study in Chlamydomonas eugametos (Chlorophyta) (Haring et al., 1996), assessment study of populations of Gracilaria chilensis (Rhodophyta) (Meneses, 1996), and comparison of polymorphisms in Gelidium vagum (Rhodophyta) (Patwary et al., 1993). In chracterisation of three species of Porphyra (Bangiales, Rhodophyta), a 20-mer M 13 primer yields amplification products which can be used to fingerprint specific genotypes (Dutcher and Kapraun, 1994). In Alberto and Santos study (1997), RAPD analysis is appropriate to characterise the genetic variability of three natural populations of Gelidium sesquipedale (Rhodophyta). As for Sargassum species, Ho et al., 1995 has reported that this technique is suitable to use in characterisation of Sargassum polycystum and S. siliquosum (Phaeophyta). In 1996, van Oppen et al. reported that RAPD data are robust in identifying large-scale biogeographic populations study of Lophocladia trichoclados (Ceramiales, Rhodomelaceae).

# 5.3.1 RAPD size

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Primer	Selected Region
OPA13	400 - 4000 bp
OPK7	300 - 4000bp
OPK9	300 - 4000 bp
OPN6	300 - 2072 bp
OPN16	400 - 3000 bp

Table 10: The sizes of amplified DNA fragments generated by RAPD primers which are selected for the analysis.

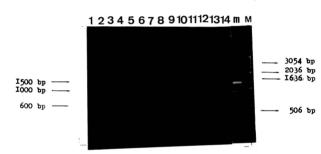


Figure 9. RAPD banding patterns of Sorgassum binderi and S. oligocystum. Results obtained with primer OPA13. Lane 1, SB2; Lane 2, SO15; Lane 3, SB12; Lane 4, SB5; Lane 5, SO27; Lane 6, SO8; Lane 7, SO14; Lane 8, SB11; Lane 9, SO18; Lane 10, SB29; Lane 11, SB17; Lane 12, SO22; Lane 13, SB6; Lane 14, SO19. The lanes marked with m is a 100-bp DNA ladder (from 1500 bp to 100 bp) and M is a 1 kb DNA ladder (from 5095 bp and 154 bp), respectively.

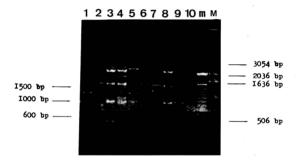


Figure 10. RAPD banding patterns of Sargassum oligocystum and S. binderi. Results obtained with primer OPK7. Lanel, SB3. Lane 2, SB6. Lane 3, SO8. Lane 4, SO11, Lane 5, SO14, Lane 6, SB12, Lane 7, SO18, Lane 8, SO 21, Lane 9, SO30. Lane 10, SB28. The lanes marked with m is a 100 bp DNA ladder (from 1500 bp to 100 bp) and M is a 1 kb DNA ladder (from 5095 bp to 154 bp), respectively.

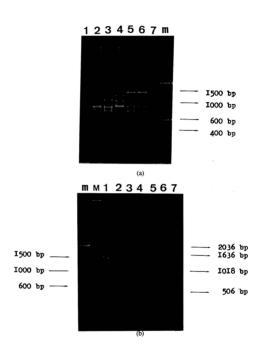


Figure 11. RAPD banding patterns of Sargassum oligocystum and S. binderi. Results obtained with primer OPNI6. a. Lane1, SO1; Lane 2, SO4; Lane 3, SO9; Lane 4, SO14; Lane 5, SO19; Lane 6, SO 25; Lane 7, SO28. b. Lane 1, SB2; Lane 2, SB6; Lane 3, SB10; Lane 4, SB16; Lane 5, SB20; Lane 6, SO23; Lane 7, SO27. The lanes marked with m is a 100-bp DNA ladder (ffrom 1500 to 100 bp) and M is a 1 kb DNA ladder (from 5095 bp to 154 bp), respectively.

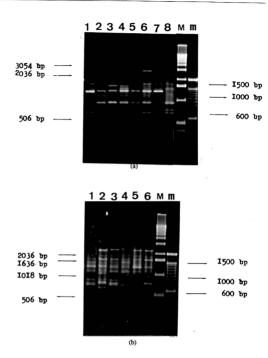
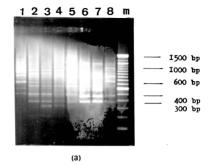


Figure 12. RAPD banding patterns of Sargassum oligocystum and S. binderi. Results obtained with primer OPK9. a. Lane 1, SO1; Lane 2, SO5; Lane 3, SO10; Lane 4, SO14; Lane 5, SO19; Lane 6, SO20; Lane 7, SO25; Lane 8 SO28. b. Lane 1, SB3; Lane 2, SB9; Lane 3, SB 12; Lane 4, SB17; Lane 5, SB25; Lane 6, SO28. The lanes marked with m is a 100-bp DNA ladder (from 1500 to 100 bp) and M is a 1 kb DNA ladder (from 5095 bp to 154 bp), respectively.



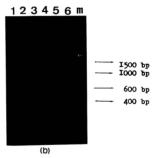


Figure 13. RAPD banding patterns of Sargassum oligocystum and S. binderi. Results obtained with primer OPN6. a. Lane1, SO1; Lane 2, SO7; Lane 3, SO11; Lane 4, SO18; Lane 5, SO23; Lane 6, SO25; Lane 7, SO29; Lane 8, SO30. b. Lane 1, SB2; Lane 2, SB8; Lane 3, SB 13; Lane 4, SB19; Lane 4, SB19; Lane 5, SB24; Lane 6, SB29. The lanes marked with m is a 100-bp DNA ladder (from 1500 to 100 bp) and M is a 1 kb DNA ladder (from 5095 bp to 154 bp), respectively.

### 5.3.3 RAPD profiles

There are bands that are shared between the individual samples of *S. binderi* and *S. oligocystum* (intraspecifically) by the five primers tested. In addition to that, there are bands which are shared by samples between *S. binderi* and *S. oligocystum* (interspecifically).

For OPA13, four bands were common to *S. binderi* in 27 out of 30 samples tested (Figure 9). They were bands at 500 bp, 700 bp, 900 bp and 1100 bp. For *S. oligocystum*, 28 out of 30 samples tested, there were three bands which were common, that is, a 500 bp band, a 700 bp band and a 1300 bp band (Figure 9). The bands that were common between the two *Sargassum* species were 500 bp and 700 bp. Therefore, 900 bp and 1100 bp were only found in *S. binderi* and band 1300 bp was only found in *S. oligocystum*. These bands may be species specific bands for *S. binderi* and *S. oligocystum*, respectively, when amplifying using OPA13.

For OPK7, two common bands appeared in 26 out of 30 samples of *S. binderi* tested, that is, a 1500 bp band and 1636 bp band (Figure 10). Three common bands sharing with 28 out of 30 samples of *S. oligocystum* were 900 bp, 1500 bp and 2072 bp (Figure 10). The single band that shared between the two *Sargassum* species was a 1500 bp band. Therefore, a 1636 band is a species specific band for *S. binderi* and

bands 900 bp and 2072 bp are species specific for S. oligocystum when OPK7 is the primer for the amplification.

For OPK9, three common bands were found in 27 out of 30 samples of *S. binderi* tested (Figure 12a). They were 1100 bp, 1300 bp and 1636 bp. For *S. oligocystum*, three common bands were also found in 27 out of 30 samples tested (Figure 12b). They were 800 bp, 1000 bp and 1400 bp. This is a very specific primer to the two *Sargassum* species tested as there was not even one common band found between the two *Sargassum* species.

For OPN6, five common bands were found in 28 out of 30 samples of *S. binderi* tested, that is, 400 bp, 500 bp, 600 bp, 750 bp and 1100 bp (Figure 13a). In 28 out of 30 samples of *S. oligocystum* tested, three common bands were found, that is, 500 bp, 600 bp and 900 bp (Figure 13b). There were two bands which/shared between *S. binderi* and *S. oligocystum*, that is, 500 bp and 600 bp. There were three species specific bands found in *S. binderi*, that is, band 400 bp, 750 bp and 1100 bp. One species specific band, 900 bp, was found in *S. oligocystum* samples. This is not a very good primer to use for *S. binderi* as there are too many bands in common.

For OPN16, six bands were found to be common in 26 out of 30 samples of *S. binderi* tested (Figure 11a). They were 400 bp, 550 bp, 800 bp, 900 bp, 1100 bp and 1500 bp. For *S. oligocystum*, four common bands appeared within 27 samples out of 30

samples tested (Figure 11b). They were 700 bp, 900bp, 1100 bp and 1500 bp. There were three common bands found between the samples of *S. binderi* and *S. oligocystum*, that is, 900 bp, 1100 bp and 1500 bp. This primer is not a good primer to use for DNA polymorphisms as there were too many sharing bands between intra-and inter-speices.

A 500 bp band was found in both Sargassum species tested while using OPA13 and OPN6 as the primers. When testing with OPK7 and OPN16, a 1500 bp band was found in both Sargassum species. There is a possibality that the two pair of primers of OPA13 and OPN6, OPK7 and OPN16 are amplifying the same region of the genomic DNA of the two Sargassum species.

### 5.3.4 Reproducibility

When appropriate precautions were taken in conducting the RAPD procedure and with the optimised conditions for PCR by a particular primer was used, the major banding pattern of a particular RAPD-PCR reaction was remarkably reproducible for independent DNA extractions of any given sample. This was true for replicates both in an experiment and between experiments (Figure 14 a, b, c, d, and e). The patterns were stable regardless of whether total nucleic acid or RNase treated DNA was used as a substrate. This showed that the method was well-standardised since only minor variations among duplicates were detected.

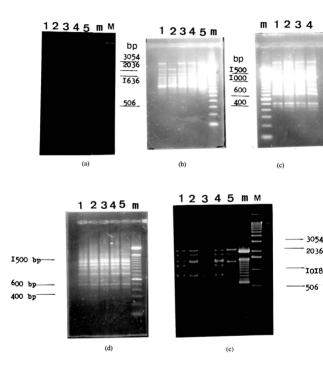


Figure 14. Reproducibility of RAPD profiles for independent DNA extractions of Sargassum oligocystum SO25. a. RAPD patterns generated by OPA 13. b. RAPD patterns generated by OPK 7. c. RAPD patterns generated by OPK 9. d. RAPD patterns generated by OPN 6. e. RAPD patterns generated by OPN 6. e. RAPD patterns generated by OPN 10. The lanes marked with m is a 100-bp DNA ladder (from 1500 to 100 bp) and M is a 1 kD DNA ladder (from 5095 bp to 154 bp), respectively.

## 5.4 Data analysis

### 5.4.1 Tables of F values

The F values for the five primers used to compare individuals of Sargassum binderi and S. oligocystum and between S. binderi and S. oligocystum are shown in Tables 13-27.

### 5.4.2 Comparison of F values

The application of similarity coefficient calculations formulated by Nei and Li (1979) gave results showing a high degree of similarity between individuals of *S. binderi* and also between individuals of *S. oligocystum*. This applied to all the primers tested (OPA13, OPK7, OPK9, OPN6 and OPN16) being used to compare for polymorphisms. The summary of the coefficient of similarity values for the 5 primers used with *S. binderi* and *S. oligocystum* are given in Table 11.

For primer OPA13, samples that showed F value > 0.75 for S. binderi is 94.67 % and 92.67 % for S. oligocystum. 95 % (S. binderi) and 74 % (S. oligocystum) of the samples gave a F value > 0.75 for primer OPK7. As for OPK9, 88.89% and 85.11% of samples from S. binderi and S. oligocystum presented a F value > 0.75, respectively. The F value > 0.75 amplified by OPN6 for S. binderi was 88.67% and

84.22% for S. oligocystum of all the samples examined. For OPN16, it was a 100% and 91% of samples from S. binderi and S. oligocystum that gave a F value > 0.75. Therefore, all the five primers used in this experiment showed high level (F value > 0.75) of intraspecific similarity for the two Sargassum species.

The two samples (species) exhibited different degrees of similarity when tested with different primers (Table 11). This is due to the fact that the primers amplify different regions of the DNA randomly. A primer that amplifies a more conserved region will give a higher degree of similarity whereas a higher dissimilarity will be obtained when a primer amplifies a more variable region of DNA.

The results of a comparison between *S. oligocystum* and *S. binderi*, is shown in Table 12. The results showed that *S. oligocystum* and *S. binderi* are not closely related as the range of the F values varies from 0.222 to 0.625 for OPA13; 0.190 to 0.667 for OPK7; 0.261 to 0.667 for OPK9; 0.250 to 0.720 for OPN6 and 0.281 to 0.632 for OPN16. The majority of the pairwise comparisons of the samples show that the two species of *Sargassum* are dissimilar (F value < 0.50) while less than 25% of the samples showed F value > 0.50. F value > 0.50 was shown in 13.89%, 16.56%, 21.33% and 24.33% using OPA13, OPK7, OPK9, OPN6, and OPN16, respectively.

Nei and Li's (1979) coefficient considers only the common presence of bands and errors may occur when considering the common presence of bands as the sole

evidence for identity. Similarly sized RAPD bands that co-migrate can have different sequences that represent different polymorphism. The problem of co-migration of non homologous bands may be a concern, particularly when testing for similarities between members of higher taxonomical levels (Hardy et al., 1992). However, the possible co-migration of the RAPD-generated bands was not considered in this study.

Primer Sample	OPA13	OPK7	OPK9	OPN6	OPN16
Sargassum binderi	0.588 - 1.000	0.588 - 1.000 0.737 - 1.000 0.609 - 1.000 0.615 - 1.000 0.783 - 1.000	0.609 - 1.000	0.615 - 1.000	0.783 - 1.000
Sargassum oligocystum	0.667 - 1.000	0.667 - 1.000   0.636 - 1.000   0.632 - 1.000   0.545 - 1.000   0.667 - 1.000	0.632 - 1.000	0.545 - 1.000	0.667 - 1.000

Table 11: Pairwise similarity coefficient (F value) of S. binderi and S. oligocystum (Intraspecies relationship studies)

Primer	F value of Sargassum binderi and S. oligocystum
OPA13	0.222 - 0.625
OPK7	0.190 - 0.667
ОРК9	0.261 - 0.667
OPN6	0.250 - 0.720
OPN16	0.281 - 0.632

Table 12: Pairwise similarity coefficient (F value) between Sargassum binderi and S. oligocystum (Interspecies relationship studies)

18 20 21 22 23 24 23	200 200 200 200 200 200 200 200 200 200
14 15 16 17 18	0.05 0.05
12 13	1860 C841 C841 C841 C841 C841 C841 C841 C841
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		_	_				_	_	_	С	_		_	_	_	_	_	0.353	0.375	0.353	0.400	0.400	0.375	0.375	0.375	0.353	0.267		3250	3,376
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	. «	_						_	_							_	_	7	0.353	0.333	0 500	9	0.471	1740	0.353	0 222	0.250	-	3363	588
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	_	_					_	_	_		_		-	_	_	_	_	0.333	0.353	0.333	0.375	0.375	0.353	0.353	0.363	0.333	0.375		1471	353
	_	_	_	_	_		_	_	_	-			_	_	_	_	_	0.353	0000	0.471	0.533	0.533	0090	0000	0.50	0.471	0.533		999	3.375
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	_	_	_	_	-		_	_	_	Ξ	_		_	_	_	_	_	0.353	0.375	0.353	0.400	0.40	0.375	0.375	0.375	0.353	0.400		3.376	88
	_	_		_	-			_	_		_		_	_	-	_	_	0.353	0.375	0.353	0.400	0.400	0.375	0.375	0.375	0.353	0.400		3.375	3.375
	_	_	_	_	Ξ	۰	_	-	_	Ξ	_	_	_	_	_	_	_	0.353	0,375	0.353	9,400	940	0.375	0.375	0.500	0.353	0.400		0250	3.376
	_	_		_	_	٠	_	_	_	Ξ	-	٠	_	_	_	_	_	0.471	0.500	0.471	0.533	0.633	0.500	0.500	0.500	0.471	0.533		3.376	8
1	_	_		_	_	_	_	_	_	_	_	_	_	_		_	_	0.353	0.500	0.471	0.400	0.533	0.500	0.375	0.500	0.471	0.400		3,375	8
	_	_	_	_	_	_	_	_	_	_	_	٠	_	_	_	_	_	0.444	0.471	0.444	0.500	0.600	0.471	0.471	0.471	147.0	0.375		0.471	353
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086 086 080 080 081 087 080 088 083 080 080 015 87 081 083 080 083 083 083 083 083 083 083 083	_	-		_	-	_	_	-	_	-	_	٠	_	_	_	_	_	4	0.353	0.333	0.500	0.375	0.471	0.588	0.353	0.222	0.250		747	383
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le 15: F value of Sargassum binderi and S. oligocystum by OPA:

90 80 80	666666666666666666666666666666666666666
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•	24 1000 24 1000 25 1000 26 1000 26 1000 27
9	0.0547
=	0.000 0.000
2	0.0947 0.0800 0.0800 0.0800 0.0800 0.0737 0.0737 0.0800 0.
2	0.0857 0.0778 0.0778 0.0800 0.
*	0.0547 0.0500 0.
2	0.0888 0.0778 0.0778 0.0778 0.0778 0.0779 0.0779 0.0779 0.0779 0.0779 0.0779
9	0.778 0.089 0.080 0.080 0.080 0.080 0.080 0.080 0.080 0.080 0.080 0.080 0.080
-	0.941 0.0842 0.0842 0.0842 0.0842 0.0842 0.0402 0.0402
10	0.0589 0.0737 0.0737 0.0737 0.0500 0.0500 0.0500 0.0500
9	0.089 0.029 0.034 0.041 0.053 0.053 0.054 0.054
R	0.842 0.0882 0.0889 0.0374 0.0374 0.0378 0.0389 0.0389
5	0.0224 0.
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Table 16: F value of Sargassum binderi by OPK7 intra

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6	0.857	0.857		_	_	-	_	908																					
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7	0.762	0.762		_	_	-	_	-	_	_	727																		
	0.737	0.737		_	_	-	_	_	_	_	_	8																	
_	0.762	0.762		_	_	_	_	_	_	_		_	8																
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9	0.818	6080	_	_	_		_	_	_	_	_	٠	_	_	_	_	_		_	_		_	_						
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22	0.857	0.857	_	_	_	_	_	_	_	_	_	٠	_	-	_	_	_	-	_	٠		_	_	_		25			
90	0.857	0.857	_	_	_	_	_	_	_	_	_	٠	_	_	_	_	_	-	_	_		_	_	-		_	9		
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Table 17: Fivalue of Samassum olicocystum by OPK7 intrasc

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8	0.632	908	2		3	88	0.421	90.836	9	9	9	0 200	0.400	0.444	0300	0.421	0.526	9.60	0,400	0.40	0.316	0300	0.526	0.626	0.421	0.381	0.400	0.00	040	0.571	0.476
21	0.286	0 28	9 6		3	0.456	0.476	0.286	8	0.364	0000	0.364	0.364	0.400	0.273	0.476	0.381	0.273	0273	0.455	0.476	0.364	0.476	0.381	0.571	0.435	0.455	0.455	0.455	0.348	0.348
8	0.40	8	8	3	9,479	0.476	940	8	642	0.381	0.528	0.381	0.478	0.528	0.476	90,400	0300	0.381	0.286	0.476	9090	0.381	9.40	999	0.400	0.456	0.476	0.381	0.478	0.455	0.364
8	0.400	9	8	3	0.470	0.476	0300	970	0.424	0.381	0.421	0.381	0.381	0.316	0.381	030	0.300	0.286	0.381	0.571	0.500	0.476	0.600	0.300	0.400	0.364	0.381	9740	0.571	0.545	0.455
75	0330	040	9	3	9070	0.381	0300	9	0.316	0.381	0.316	0.381	0.476	0.526	0.286	0300	0.600	0.381	0.476	0.476	0.400	0.381	0.500	0.500	0.400	0.455	0.381	0.286	0.381	0.364	0.545
ន	0.424	0318	0.40		3	0.400	0.421	1040	9550	900	9990	0090	0.500	0.444	0.400	0.526	0.528	0.400	0.300	0.300	0.421	0.400	0.528	0.421	0.632	0.476	0.500	0300	0.400	0.381	0.478
8	1040	0.318	0.424		3	8	0.421	1040	9520	9090	9990	0300	0.400	0.444	0.400	0.421	0.526	0.500	0.400	0.300	0.316	0.400	0.316	0.316	0.421	0.381	0.500	0.400	0.500	0.476	0.476
2	0.400	0.400	0.400	200	0.470	0.381	0.400	970	0.421	0.476	0.421	0.476	0.476	0.526	0.381	0.300	0.400	0.286	0.476	0.476	0.400	0.381	0.500	0.400	0.400	0.273	0.571	0.571	0.381	0.455	0.455
R	0.400	0.400	040	9000	007.0	0.381	0.300	0300	9316	0.381	0.316	0.571	0.381	0.421	0.286	0.500	0.400	0.381	0.286	0.381	0300	0.381	0.500	0.500	0.400	0.545	0.571	0.478	1/20	0.364	0.456
9	0.526	1421	0 632	200	3	0.400	0.421	0.421	9590	0050	0.444	0.400	0300	0.333	900	0.421	0.421	0.600	0.400	0.500	0.526	0.500	0.316	0.632	0.526	0.476	009	0300	940	0.381	1381
9	9250	0.333	9990	346	2	0.421	9990	0 444	0.471	0.316	0353	0.316	0.421	0.353	0.421	0.556	0.444	0.421	0.316	0.421	0.556	0.526	9220	*	#	9	3.316	3316	3.526	0.400	0000
=	١.									0.316																					300
9	0.421	0.316	9316	900		88	0.421	0.316	0.333	0.400	0.444	0300	0.400	0.444	0.300	0.421	0.316	0.500	0.400	0.400	0.421	0.300	0.421	0.421	3.316	1381	97.	909	009	3.476	381
2	0.316	0.316	0.316	900	3	900	0.421	0.316	0.333	97	0.444	0.500	0.500	99970	0000	0.421	0.421	0090	0.400	0.400	0.421	0300	9729	7.421	3,316	286	88	909	009	0.381	3.476
•	l.,						_		_		_			_												7		Ξ	_	1522 (	-1
2	97.0	300	300	326		8	89	300	316	381	0.421	1381	381	975	3286	8	900	9.476	1381	286	8	586	8	8	8	364	1476	1381	1381	273	455
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. S. olipocyatum

able 18: Fivalue of Sargassum bindert and S. olgocystum by OPKT Interspecifically

8	
8	666.0
83	0.870
52	0.909 0.870 0.942
8	0.952 0.902 1.000
8	0.818 0.757 0.833
75	 0.870 0.809 0.809 0.809 0.809
8	0870 0800 0833 0833
R	0.397 0.308 0.308 0.808 0.808 0.807 0.807
7	0.957 1.000 0.917 1.000 0.917 0.833 0.833
R	1,000 1,000
6	0.957 0.957 0.957 0.950 0.950 0.950 0.850
6	0.857 1.000
4	0.909 0.909 0.909 0.909 0.909 0.909 0.909 0.909 0.909 0.909 0.909 0.909 0.909 0.909
16	1,000 0,993 0,993 0,993 0,993 0,993 1,000 0,993 1,000
5	0,909 0,909 1,000
7	0.857 0.9852 0.9852 0.9852 0.9857 0.9857 0.9857 0.9858 0.9
5	0.952 0.953
12	0.999 0.999
=	0.000 0.000
5	(1000) (1
œ	0.952 0.952 0.957 0.957 0.957 0.957 0.959 0.957 0.959 0.957 0.957 0.959
40	1,000 0,995
_	0.952 0.952
φ	0.959 0.959
so.	0.000 0.000
	2562 2562 2562 2562 2562 2562 2562 2562
e	200 200 200 200 200 200 200 200 200 200
5	1,000 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0
_	2582 2587 2587 2587 2587 2587 2587 2587
Sample SB SB	
S S	жикиминателетеления

83	
83	888
22	0.0824
83	0.7.0 0.7.0 4.28.0
23	0.700 0.700 0.778
24	
8	0.0889 0.778 0.778 0.778
n	0.889 0.778 0.982 0.0424
21	0.0889 0.0889 0.0778 0.0706 0.0778
8	1,000 0 0,000 0 0,000 0,000 0,000 0,000 0,000 0,000 0,000 0,000 0,000 0,000 0,000 0,000 0,000 0,000 0,000 0,000 0,000 0,000 0,000 0 0,000 0 0 0
5	1,000 1,000
92	1,000 1,000
17	0.889 0.889 0.889 0.089 0.089 0.0776 0.089
16	0.889 0.889 0.889 0.889 0.889 0.089 0.089
12	0.889 0.778 0.889 0.889 0.889 0.889 0.889 0.824 0.827 0.824
7	0.889 0.889 0.889 0.889 0.889 0.889 0.889 0.889 0.889
13	0.858 0.0858 0.0858 0.0858 0.0858 0.0858 0.0858 0.0858 0.0858
12	1,000 1,000
=	0.824 0.824 0.824 0.944 0.944 0.944 0.944 0.944 0.944 0.944
5	0,000 0 0,000 0,000 0,000 0,000 0,000 0,000 0,000 0,000 0,000 0 0,000 0
•	2000 2000 2000 2000 2000 2000 2000 200
	0.0667 0.0708 0.0889 0.0889 0.0889 0.0889 0.0788 0.0778 0.0788 0.
-	0.0778 0.0788 0.0788 0.0778 0.0778 0.0778 0.0778 0.0778 0.0778 0.0778 0.0778 0.0778 0.0778 0.0778
۰	0.778 0.0289 0.0289 0.07788 0.07788 0.077
e0	0.842 0.7347 0.842
-	0.042 0.0778
~	0.778 0.778 0.778 0.778 0.089
2	0.000 0.000
1	00×00×0×0×0×0×0×0×0×0×0×0×0×0×0×0×0×0×
Semple SO 1	0.889 0.0299

able 20: F value of Sargassum oligocystum by OPN16 intras

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S																														8		
83																														0.42		
83																														8		
22	0.476	0.476	22.0		8	9	0.476	0.476	0.476	0.476	0.671	0 400	0.476	0.673	0.476	9	9 4 4 9	9	200	900	0.470	000	3	9 5	200	9	8	0.00	9	0.571	0.38	0.476
83	0090	8	3	3	3	0.476	8	8	8	940	0090	0.421	8	8	8	3 8	38	3 8	3	3	8	36	3	3 8	38	300	0.526	900	0.528	990	9	8
23	0.500	8	3 8	3	3	93	8	800	900	0300	8	9080	8	8	3 5	3	38	3	3	8	8	800	3	8	3 8	8	0.476	0.421	988	999	80	0,40
72	1070	9090	2000	0.000	0.421	8	0.421	0.526	9000	1040	0.424	9880	0.476			3	0.421	000	0.421	0.526	0.526	0.421	0.526	0.526	0.421	0.421	0.444	0.556	8	9.50	0.42	0632
B	0.334			010	0.476	0.456	0.571	0.381	0.476	8740	0.671	000	300	300	2	200	0.476	9/4/0	0.476	0.476	0.476	0.476	0.476	0.571	0.476	0.478	0.400	0.50	0.400	0.476	0.381	0.381
2	9000	3 8	3	3	880	0.381	0.50	989	0090	8	8	3 4 5	200	3 6	36	3	900	8	880	990	0.500	900	8	8	8	800	920	0.526	0.421	0.600	0.500	0.50
77	1000	300	1000	33	0.286	0.364	0.476	0.476	0.476	9476	200	200	3	000	9	200	980	0.926	0.526	0.526	0.526	0.526	0.526	0.381	0.476	0.476	900	0.50	000	0.500	0.476	0.571
8	92.5		1/00	1981	0.381	0.545	0.476	0.571	0090	2000	97.0	200	3 5	0.470	0.470	0.476	0.476	0.476	0.476	0.476	0.476	0.476	0.476	0.381	0.476	0.571	0.50	0090	0300	0.571	0.478	0.286
92	830	3	3	8	8	0.476	0.400	0020	980	200	3 5	3 8	3 3	3	3	8	900	8	89	999	020	8	0.50	0.500	900	9	0.421	0.526	0.421	0090	0.500	0090
90	9740		2	200	0.671	0.455	0.476	0.476	0.476	0.476	0.476	2	3 5	200	0.470	700	0.381	0.472	0.476	0.381	0.381	0,381	0.381	0.286	0.476	0.571	0.40	899	0.400	0.476	0.381	0.476
-	100		9700	77.	0.526	0.400	0.421	0.526	0.626	1070		0 6 6 6 0	3 3	970.0	0.00	0700	0.526	0.526	8	0.566	0.556	0.556	0,556	0.526	0.421	0.632	0.444	929	0.44	0.526	0.526	0.632
16	100		700	9750	0.632	999	0.421	0.526	0.632	100		7	0.470	3	0.4/0	8	0.50	88	0.526	0.421	0.421	0.421	0.421	0.421	0.500	0.528	0.556	0.556	0.333	0.526	0.316	0.632
22	1630	200	0.07	0.476	0.476	0.455	0.381	0.571	0.478	0.00	0.50	0.470	3	9/4/0	0.4/6	0.421	0.476	0.571	0.381	0.281	0.281	0.281	0.281	0.381	0.571	9.58	0.50	0.500	0000	0.571	0.476	0.571
7	200	3	8	8	920	0.571	0.400	040	9	3 3	3 6	3	700	8	9	8	0.400	0.400	0.400	0.300	0.300	0300	0300	0.400	0.400	0.500	0.421	0.526	0.421	0.600	0.400	0.400
5	0000	3	0.476	0.472	88	0.500	0.526	7770	9	3	200	0.421	000	0.526	0.526	965	0.526	0.526	0.526	0.526	0.526	0.526	0.526	0.421	0.526	999	0.444	0.476	0 444	0.526	0000	0.500
52	0	0.4/0	0.476	0.472	0.571	0.455	0.476	0.478	0.874		9	3	3	0.57	0.571	0.476	0.476	0.476	0.476	0.381	0.381	0.381	0.381	0.476	0.476	0.476	0.400	0.500	0.500	0.476	0.381	0.476
=	200	9700	800	1	88	0,500	0.421	9650	9		,	750	3	720	8	0.42	0.526	8	0.632	0.421	0.421	0.421	0.421	0.421	0.526	0.632	0.44	9999	0.333	0.632	0.526	0.632
9	1		8	0.526	0.421	0.500	0.526	0.400	0.632		75.0	75.0	0.00	8	8	0.526	0.556	0.526	0,600	0.421	0.421	0.421	0.421	0.421	0.526	0.526	0.44	0.444	0.333	0.526	0.421	9250
00	200	3	8	8	9	0.381	0.500	0400	000	3 3	3 5	3	0.520	8	890	8	0.60	800	900	0.500	0.500	0.500	0.500	0.400	0.400	0.500	0.421	0.421	0.421	0.500	0.500	0090
	0000	3	8	8	800	0.478	0.500	0.50	97.50	200	3	3	8	999	889	8	0.500	88	0000	0.400	0.400	0.400	0.400	0.500	0.500	0.500	0.421	0.528	0.421	0 800	0 500	0.600
-		045	0.526	0.526	0.632	0 200	0.526	9636		3	0.42	0.421	920	8	0.632	88	0.526	0.526	0.526	0.526	0.526	0.526	0.526	0.421	0.421	0.526	0 444	7770	7770	1270	0.42	0.632
6		0.476	0.571	0.476	0.571	0.455	0.476	977		200	0.36	0.381	0.476	99	0.571	0.476	0.476	0.571	0.476	0.476	0.476	0.476	0.476	0.381	0.381	0.381	0.400	030	9	0.785	0.786	0.286
0	1	0.476	8	0.526	0.456	0.400	7770	900	2000	9	0.421	0.421	9990	0.476	900	440	0.632	9000	0.526	0.421	0.421	0.421	0.421	0.421	0.526	0.832	9990	7770	1	9636	200	0.526
-		89	889	999	909 0	97.40	8		3	3	8	0.400	0.381	0.600	0.500	0000	0090	0000	0000	000	0090	0090	0050	0.500	0.50	0.600	9080	200		200	38	300
6		0.476	0.476	0.571	0.674	9990			9	0.470	0.381	0.381	990	0.455	0.571	0.400	0.671	0.476	0.478	8740	977	0.476	9770	0.476	0.476	0.571	8	3 8	3	3	0.00	0.381
2		0.421	0.526	0.421	0.832	180	35	7	0.00	0.421	1	0.421	#	0.632	0000	0.526	0.632	0.444	20.00	10	1040	0.42	1070	1070	0.526	0.632	986	3		333	250	0.421
-		0.400	0000	0070	8	100		3	300	800	9.40	0300	0.526	999	9000	0090	0 200	0.500	940	8	8	9	8	9	0050	000		200	770	0.310	3 8	8 8
BS 88		ĺ	_	_							_									_	_	_					_				_	
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8	
	606 0
23	2,952 2,857 2,870
8	7777 2777 2765 00
27	2,783 2,870 2,977 2,977
98	2720 2720 2720 2833 2890 2890 2890
123	
24	
23	2923 2923 2941 0.9 2750 0.8 2750 0.8
z	7720 7720 7720 7720 7730 7730 7730 7730
21	00000000
8	77 0.788 0.750 0.788 0.750 0.768 0.750 0.768 0.750 0.7
18	00 0.057 00 0.773 00 0.773 00 0.773 00 0.052 00 0.052 00 0.052 00 0.052 00 0.052 00 0.052
18	77 0800 77 0800 77 0.802 78 0.802 78 0.763 70 0.763 70 0.818 71 0.818 71 0.818 72 0.818 73 0.805 74 0.805
1,	3 0.857 7 0.0257 7 0.0257 3 0.0258 0 0.0258 0 0.0258 0 0.0258 0 0.0258 0 0.0258 0 0.0258 0 0.0258
9	3 0.783 8 0.783 8 0.727 9 0.838 0 0.830 0 0.830 0 0.800 0 0.800 0 0.800 0 0.800 0 0.800 0 0.800 0 0.800 0 0.800 0 0.800 0 0.800
\$	2 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0
=	0.0870 0.0762 0.0762 0.0762 0.0762 0.0857 0.0857 0.0858 0.0858 0.0858 0.0858 0.0858 0.0858 0.0858 0.0858 0.0858
5	0.8370 0.723 0.723 0.723 0.0370 0.037
12	0.917 0.917 0.017
=	0.833 0.633 0.633 0.633 0.635 0.630 0.730
9	0.800 0.800 0.800 0.800 0.800 0.800 0.800 0.700
00	0.917 0.666 0.818 0.870 0.870 0.870 0.870 0.870 0.727 0.818
0	0.762 0.763 0.6138 0.6138 0.6138 0.6139 0.61
-	0.000 0.000
	0.657 0.7
o	0.783 0.816 0.816 0.850 0.880 0.880 0.880 0.817 0.818
-	0.000 0.000
m	0.923 0.930 0.733 0.733 0.733 0.930
2	0.0960 0.0970 0.0970 0.0970 0.0970 0.0970 0.0970 0.0970 0.0970 0.0970 0.0980 0.
-	0.960 0.0633 0.0
Sample S8 S8	

8	
,	ବେହ
	0.000 1.000
	1000
.	0.842 0.842 0.842
,	0.889 0.947 0.947
	0.706 0.706 0.778 0.778
,	2.889 0.842 0.737 0.736 0.708 0.708 0.708
	0.889 0.875 0.778 0.778 0.778
.	0.889 1.000 0.889 0.885 0.887 0.857 0.800 0.900 0.900 0.900
,	2337 2337 2337 2337 2337 2332 2382 2382
.	0.941 0.750 0.657 0.657 0.657 0.657 0.657 0.657 0.657 0.657 0.657
.	0.875 0.941 0.778 0.778 0.776 0.000
.	0.875 0.0278 0.0378 0.0778 0.0374 0.0374 0.0374 0.0374 0.0375 0.0375 0.0375
	0.0842 0.0842 0.0847 0.0857 0.0857 0.0842 0.0842 0.0842 0.0752 0.0752 0.0752
2	0.750 0.778 0.778 0.737
	0.737 0.850 0.706 0.706 0.706 0.706 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.842 0.844
2	0.778 0.700
,	0.778 0.778 0.000
=	0.000 0.000
	0.941 0.706 0.706 0.800 0.817 0.817 0.817 0.817 0.817 0.817 0.817 0.817 0.817 0.817 0.817 0.817 0.817 0.817 0.817
.	0.0341 0.058 0.0778 0.0750 0.0750 0.0274 0.0273 0.0824 0.0824 0.0824 0.0824 0.0824 0.0824 0.0824 0.0824 0.0826 0.0827 0.0826 0.0827 0.0826 0.0827 0.0826 0.0827 0.0826 0.0827 0.0826 0.0827 0.0827 0.0826 0.0827 0.0826 0.0827 0.0826 0.0827 0.0826 0.0827 0.0827 0.0826 0.0827 0.0
	1,000 0,941 0,078 0,078 0,023
	1,000 1,000 1,000 0,0178 0,0178 0,0178 0,0178 0,0178 0,0178 0,0178 0,0178 0,0178 0,0178
	0.941 0.0941 0.0941 0.00000 0.0000 0.0000 0.0000 0.0000 0.0000 0.0000 0.0000 0.0000 0.00000 0.00000 0.00000 0.00000 0.00000 0.00000 0.00000 0.00000 0.00000 0.00000 0.00000 0.00000 0.00000 0.00000 0.00000 0.00000 0.000000
	0.024 0.0778 0.0778 0.0778 0.0778 0.0778 0.080 0.080 0.080 0.090 0.0000 0.000
	0.0224 0.706 0.706 0.706 0.706 0.706 0.824 0.824 0.837 0.837 0.837 0.738
,	0.750 0.750 0.750 0.750 0.750 0.750 0.778 0.778 0.778 0.778 0.779 0.779 0.770
N	0.706 0.828 0.828 0.828 0.778 0.778 0.778 0.778 0.778 0.778 0.779 0.770
_	0.778 0.024

able 23. F value of Sargassum oliocoverum by OPKG in

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۳	22	۳	۲	۲	۳	۳	1-	0.476	0.476	ľ	0.500	0.381	0.476	0.400	0.421	0.526	0.50	0.381	0.364	0.364	0.500	0.465	0.381	9 9	9 5	0.526
-		_	_	٦	۰	_	-	_	_	_	_	0	120	8		~	_	0.57	_	3	3	8	9	3	3	
		-		•	٠	•	-	-	_	_	_	0	9	0.421		~	_	8	_	0.476	9790	0.38	8	0.316	0.316	1
			•	•	•				-	-	-		0.500	0.526		~	_	88	_	0.571	0.526	0.476	8	0.42	929	3
-		-	•	•	•							, ,	0.478	000		-	-	0.381	_	0.545	0.500	950	0.571	990	8	0.526
_		_	_	_	•	_						•		3			•	8		0.381	0.42	0.38	98	0.421	0.421	9990
_		_	_	_	•	_			_	_	_	•	3	0.000			•			7000	8	95.0	974	040	0300	0.316
_		_	_	_	•	_			_	_	_	•	0.470	3			•				1			8	000	9630
_		_	~	٠	٠	_			_	_	_	۰	0.476	8		-	_	200			3				2	2
7		-	`	-	٦	_			_	_	-	_	0.476	8		-	_	0.381	-	3	3	3	975	3	3	
3	6		•	•	•				-	-	-	•	0.500	0.421		-	_	9	-	0.476	0.526	0.381	8	0.526	0.421	0.476
2	3 6	-			•								0.478	0.40		_	_	0.476	-	97	940	0.364	0.381	9.40	940	0.526
ş	3	-	-	-	•	-						•	0.470	8			-	0.781	-	0.455	0.400	0.364	0.381	0300	8	0.42
\$	ò	_	_	_	٠.	_						•		3 8				O Tat		977	9	0.465	0.476	0000	0.400	0.526
36	ò	_	_	_	_	-			7	-	-	•	200	3 8				200		0.456	0	9770	10.00	0400	909	0.526
Z	ò	_	_	_	•	-			-	-	-	٠.	9 6	3				9		200	9778	250	3	0.476	0.476	0.400
522	0	-	_	_	-	_			-	-		٠.	3					3 2		2417	0354	0333	0.622	0.455	0.455	0.381
g	ò		_	-	_	_			-			•	200	200				18		9776	9630	0.671	8	0.526	0.381	1
57	ò	-	_	-	_	_							3	0.50				3 8		927	200	0.476	8	3	9630	7770
0.476	o	0.476 0.	0,400	0.421 0.558	Se 0.556	6 0.526	6 0.381			9		٠.	950	0.000			-	3 8		0 284	0.528	0.476	9	0.526	0.526	*
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OC. Carpasanti Carcopanti

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S. обросувыт

27: Fivalue of Seroassum bindert and S. oliocoystum by OPN6 Interspecifically

# 5.4.3 Phylogenetic analysis

The phylogenetic analysis has been widely used in studying the relationships of algae. Saunders et al. (1995) reported that this technique is useful to study the relationships of species of uncertain taxonomic postition within the Achrochaetales-Palmariales complex (Rhodophyta) using 18S rDNA sequence. In that particular study, the phylogenetic trees were inferred by neighbour-joining (Saitou and Nei, 1987) using the DNADIST program of the PHYLIP package version 3.5 (Felsenstein, 1985). The cryptic diversity study of Porphyra (Rhodophyta) by mapping the restriction sites of several restriction enzymes to the small sub-unit (SSU) rRNA genes and the phylogenetic analysis was done by using the mix parsimony and maximum likelihood programs in the PHYLIP computer package (version 3.41) (Felsenstein, 1989) by Stiller and Waaland (1993). In 1996, Lee and King had reported the usefulness of the phylogenetic analysis to characterise five genera of Dictyotaceae (Phaeophyta) based on the DNA sequences of nuclear rDNA internal transcribed spacer (ITS) and 5.8S. The tree distance analyses were conducted using DNADIST in the PHYLIP 3.57 package (Felsenstein, 1993). In the close evolutionary relationships study among the Sporochnales, Desmarestiales and Laminariales (Phaeophyceae) (Tan and Druehl, 1996), parsimony analysis of the 18S rDNA sequences was done with the PAUP computer package (PAUP 3.1.1, Swofford, 1993) and the phylogenetic trees were conducted by neighbour-joining analysis (neighbour -joining MEGA; Kumar et al., 1993).

The following is a discussion on the results of the cluster analysis conducted for the phylogenetic relationships between the two *Sargassum* species using the different primers.

## 5.4.3.1 OPA13

Two main clusters (S. binderi and S. oligocystum) (Figure 15) which consist of two and three subclusters, respectively. The two subclusters from S. binderi, that is, first subcluster: samples 1, 18, 20 and second subcluster: samples 2, 25, 15, 17, 22, 30, 26, 3, 4, 7, 8, 6, 9, 13, 12, 5, 10, 11, 27, 19, 16, 21, 14, 24, 23, 28, 29. Samples 31, 59, 32, 33, 34, 38, 35, 41, 36, 37, 45, 56, 55, 58, and 57 are under the first subcluster of S. oligocystum. The second subcluster consists of samples 40, 60, 48, 42, 46, 44, and 47. As for the last subcluster of S. oligocystum, the members are samples 39, 50, 53, 43, 49, 51, 52, and 54.

Samples 1 to 30 belonging to *S. binderi* are 81.4% similar. From the second subcluster, four groups of samples, that is, samples 3, 4, 7, 8, samples 6, 9, 13, samples 5, 10, 11, and samples 14, 24 give a 100% of similarity. There are two individual lineages, that is, samples 26 and 19 which show 85.7 % and 87.9% similarity to the main cluster of *S. binderi*, respectively.

Samples 31 to 60 belonging to *S. oligocystum* are 80% similar. There are eight groups of samples sharing a 100% similarity. Five out of eight are from the first subcluster, that is, samples 31, 59, samples 32, 33, 34, 38, samples 36, 37, samples 45, 56 and samples 55, 58, 57. Samples 42, 46 and samples 44, 47 from the second subcluster and samples 52, 54 from the last subcluster have a similarity level of 100%.

By comparing between the two main clusters, *S. binderi* and *S. oligocystum*, only 42.1 % similarity is shown. Therefore, by using OPA 13 as a primer, *S. binderi* is distantly related to *S. oligocystum* at the species level.

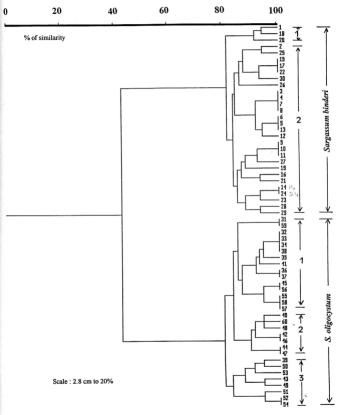
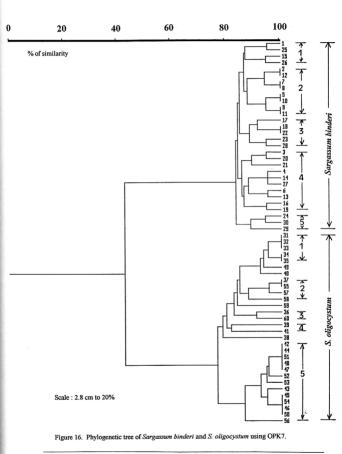


Figure 15. Phylogenetic tree of Sargassum binderi and S. oligocystum using OPA13.

similarity. For the individual lineages, sample 49 showed 93.57% similarity, sample 40 gave 92.14% similarity, sample 59 and 38 is 83.57% and 78.57% similar to the main cluster of *S. oligocystum*, respectively.

The two main clusters, that is, S. binderi and S. oligocystum are only 42.14% similar. Therefore, OPK7 as a primer indicates that S. binderi is also distantly related at the species level.



## 5.4.3.3 OPK9

The two main clusters, representing *S. binderi* and *S. oligocystum* (Figure 17), has six subclusters each. Samples 1, 3, 2 are grouped under the first subcluster of *S. binderi*. In the second subcluster, there are only three members, which include samples 5, 12, 13. Third subcluster contains samples 4, 22, 23. Fourth subcluster is made up by samples 10, 15, 14. Samples 8, 18, 27, 11, 20, 19, 29, 24, 30 are in the fifth subcluster and samples 6, 21, 7, 9, 16 are grouped under the last subcluster. Samples 13, 25, and 17 are individual lineages under the main cluster of *S. binderi*. As for *S. oligocystum*, the first subcluster consists of samples 31, 46. The second subcluster has six members, samples 36, 40, 37, 38, 39, 41. Samples 51, 53, 52,54 are grouped under the third subcluster. In the fourth subcluster, are samples 32, 47, 49, 35, 34, 50, 48. The fifth subcluster consists of samples 55, 57, 60, 58, 59, 56 and samples 42, 43, 45, 44 are separated into the sixth subcluster. There is only one individual lineage, that is, sample 33.

S. binderi has 80% similarity intraspecifically. There are two pairs of samples sharing a 100% similarity in the second subcluster, that is, samples 5 and 12 and the fifth subcluster, that is, samples 19 and 29. The individual lineages are sample 13, 17 and 25 which are 90%, 80.7% 82.9% similar to the main cluster of S. binderi, respectively.

For S. oligocystum, the intraspecific similarity level is 72.9%. Two pairs of samples under the second subcluster, that is, samples 36, 40, and samples 37, 38, 39, one pair under the third subcluster, samples 51, 53, one pair in the fourth subcluster, samples 47, 49 and one pair of sample grouped under the fifth subcluster, samples 57, 60, 58 indicated a 100% similarity intraspecifically. The individual lineage, sample 33 only showed 72.9% similarity to the main cluster of S. oligocystum.

By using OPK9 as a primer, the similarity level interspecifically is only 45.4%. Therefore, S. binderi is only distantly related to S. oligocystum genetically.

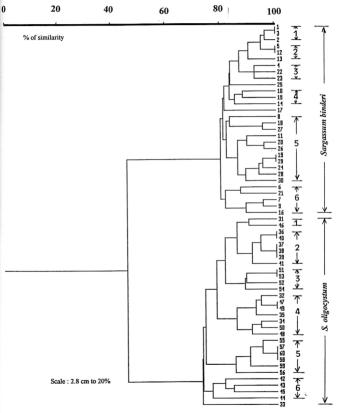


Figure 17. Phylogenetic tree of Sargassum binderi and S. oligocystum using OPK9.

#### 5.4.3.4 OPN6

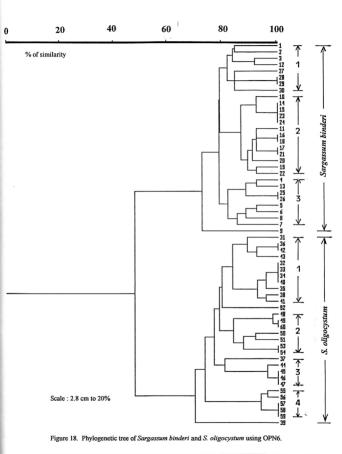
Two clusters are formed (Figure 18), that is, one from *S. binderi* and another from *S. oligocystum*. The cluster of *S. binderi* contained three subclusters, that is, samples 1, 2, 3, 12, 27, 28, 29, 30 (first subcluster), samples 10, 14, 15, 23, 24, 11, 16, 18, 17, 21, 20, 19, 22 (second subcluster), samples 4, 13, 25, 26, 5, 6, 8, 7 (third subcluster) and one individual lineage, that is, sample 9. Four subclusters can be found from the cluster of *S. oligocystum*, that is, samples 31, 36, 42, 43, 32, 33, 34, 40, 35, 38, 41 (first subcluster), samples 48, 49, 60, 50, 51, 53, 54 and samples 37, 44, 45, 46, 47 (second subcluster) samples 55, 56, 57, 58, 59 (third subcluster) and two individual lineages, that is, samples 52 and 39.

There are 77.2% similarity showed from samples 1 to 30 belonging to *S. binderi*. Four groups of samples showed the highest similarity level at 100%. Samples 28, 29 are grouped under the first subcluster, samples 14, 15, 23, 24, samples 16, 18, and samples 17, 21 from the second subcluster, and samples 25, 26 from the third subcluster. As for the individual lineage, that is, sample 9, the similarity level is 72.8% to the mian cluster of *S. binderi*.

Samples 31 to 60 belonging to S. oligocystum are 74.2% similar. From the dendrogram, there are four groups of samples showing 100% similarity, that is, samples 36, 42, and samples 32, 33, 34 from the first subcluster, samples 49, 60 and

samples 53, 54 from the second subcluster, samples 45, 46, 47 from the third subcluster and samples 57, 58, 59 from the last subcluster. The two individual lineages, samples 52 and 39 are 79.3% and 70% similar to the main cluster of *S. oligocystum*, respectively.

By comparing the clusters of *S. binderi* and *S. oligocystum* using OPN6 as a primer, these two species are distantly related at the similarity level of 47.9%.



## 5.4.3.5 OPN16

There are generally two main clusters (Figure 19) involved here, that is, one cluster from *Sargassum binderi* which contains two subclusters, that is, samples 1, 4, 8, 9, 14, 22, 3, 6, 12, 15, 18, 20, 21, 23, 19 25 (first subcluster), samples 2, 5, 7, 10, 11, 13, 16, 17, 26, 30, 27, 28, 24 (second subcluster) and one individual lineage, that is, sample 29. The other cluster, that is, *S. oligocystum*, has five subclusters, that is, samples 31, 52, 38, 33, 42, 46, 43, 47, 48, 50, 53, 51, 49 (first subcluster), samples 32, 58, 54, 45, 41, 60 (second subcluster), samples 57, 59 (third subcluster), samples 34, 55, 35, 39, 44, 37 (fourth subcluster), and samples 36, 40 (fifth subcluster) and one individual lineage, that is, sample 56.

Samples 1 to 30 belonging to *S. binderi* showed 91% similarity. From the first subcluster, there are two groups of samples which have 100% similarity, that is, samples 1, 4, 8, 9, 14, 22 and samples 3, 6, 12, 15, 18, 20, 21, 23. In the second subcluster, samples 2, 5, 7, 10, 11, 13, 16, 17, 26, and 30 appear to be 100% similar. For the individual lineage (sample 29), it showed 87.9% similarity to the cluster of *S. binderi*.

Samples 31 to 60 belonging to *S. oligocystum* showed 78.6% similarity. From the first subcluster of this species, there are three groups of samples sharing 100% similarity, that is, samples 31, 52, samples 42, 46, 43 and samples 48, 50, 53, 51, 49. In the

second subcluster, the 100% similarity are contributed by samples 32, 58, 54, and 45. Sample 56 was an individual lineage with 77.1% of similarity to the main cluster of S. oligocystum.

Between the two main clusters, there is only 52% similarity and this shows that *S. binderi* is only distantly related to *S. oligocystum* when primer OPN16 is used to obtain polymorphisms, for comparison.

As a result of too many pairs of samples having 100% similarity, OPN16 is not a good primer to separate the samples among the species. Therefore, this primer is not very suitable to use as a marker to generate DNA fingerprints to screen S. binderi and S. oligocystum.

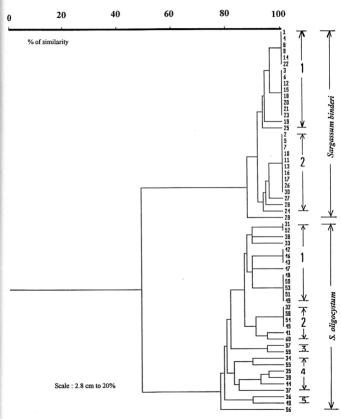


Figure 19. Phylogenetic tree of Sargassum binderi and S. oligocystum using OPN16.

## 5.4.4 Probe for DNA fingerprinting

All the primers, OPA13, OPK7, OPK9, OPN6 and OPN16 showed that *S. binderi* is only distantly related (similarity level in between 42 % to 52 %) to *S. oligocystum* based on the similarity level. The five primers can easily separate out the two *Sargassum* species into two main clusters. In between the individuals of *S. binderi*, all the samples showed a very high similarity level (71 % to 91 %). For all the individual samples of *S. oligocystum*, the similarity level obtained also gave high values (72.0 % to 80 %).

Among all the primers, OPK9 is the best primer to characterise the two Sargassum species. This primer can separate out very clearly even up to the individual level of all the samples tested (figure 17). Therefore, OPK9 may be suitable to use as a probe for DNA fingerprinting.

For OPK7, it is only to good use to separate out the individuals of *S. binderi* but not *S. oligocystum* although the separation between the two *Sargassum* species is clear (figure 16). This is because 16 out of 30 samples of *S. oligocystum* tested showed a similarity level of 100 %.

15 out of 30 samples of *S. binderi* and 19 out of 30 samples showed a similarity level of 100 % indicating that OPA13 is not a good primer to use to separate out the

individuals between S. binderi and S. oligocystum although the separation between the two Sargassum species is clearly obtained (figure 15).

When OPN6 was used, 12 out of 30 samples of *S. binderi* and 16 out of 30 samples showed a similarity level of 100 % (figure 18). Although the relationship of the two *Sargassum* species is clearly showed, it is not a good primer to separate the samples at the individual level.

OPN16 is the most unsuitable primer if samples were to sort out by individuals as 24 out of 30 samples of *S. binderi* tested showed 100 % similar to the main cluster of *S. binderi* (figure 19). 14 out of 30 samples belonging to *S. oligocystum* also showed 100 % similar to the main cluster of *S. oligocystum* (figure 19). Therefore, it is not suitable to use as a probe for DNA fingerprinting.

# 5.5 Molecular data versus morphology

The results obtained from this study using molecular technique, that is, RAPDs clearly showed that S. binderi and S. oligocystum are two different species. They are not synonyms as reported by Womersley and Bailey (1970). This study supported the hypothesis proposed by Ajisaka et al. (1998) that S. binderi is different from S. oligocystum by two characters, that is, the vesicles and the receptacles of the plant. The vesicles of S. binderi are winged and S. oligocystum are not. The receptacles of S. binderi are twisted and with sharply dentate margins but S. oligocystum usually have spines only at the tips of the receptacles.