

CLOSED STACKS

Larvicidal Principles From Some Insecticidal Plants of Sarawak

Gwendoline Ee Cheng Lian

A Dissertation
Submitted for the Degree of
Doctor of Philosophy

Doctor of Philosophy

April, 1996 Department of Chemistry Faculty of Science University of Malaya Kuala Lumpur _{Dimitrofistan peda...}

Perpustakaan Universiti Malaya

HAMSIAH BT. MOHAMAD ZAHARI UPR UNIT REPPORPAFI PERPUSTAKAAN UTAM UNIVERSITI MALAYA

PERPUSTAKAAN UNIVERSITI MALAYA

16.08.2000

ACKNOWLEDGEMENTS

I would like to express my deepest gratitude and sincere appreciation to Professor Dr Goh Swee Hock for his invaluable guidance, constant supervision, intellectual advice and constructive criticisms throughout the course of this study and during the preparation of this thesis. I also wish to thank Dr. Chuah Cheng Hock and Professor Yong Hoi Sen for their supervision during Professor Goh's sabbatical leave. I also benefited from Dr. Chuah's supervision and help towards the later part of this study.

My sincere thanks and deepest gratitude are also extended to Mr. Lee Han Lin of the Institute of Medical Research, Kuala Lumpur for guidance on bioassay work involving Aedes aegypti. I am also grateful to Mr. K. T. Wong from the Pharmacology Department, National University of Singapore for conducting in vitro bioassays. Many thanks also go to Gnanamalar Panandam for typing this thesis. To K. Yoganathan, Lee Kee Huat, Ooi Kay Eng. Hew Nam Fong and Dr. Richard Wong thank you for all the help you have given me throughout my course of study.

I also wish to thank my parents, my two sisters Evelyn and Susan and my two sons Joey and Lance for their patience, tolerance and moral support. To them I dedicate this work. Finally, I would like to thank my husband Jegak Uli for encouraging me to undertake this study. I also thank him for collecting plant samples and for preparation of this thesis for without him I would never have achieved this piece of work.

ACKNOWLEDGEMENTS

I would like to express my deepest gratitude and sincere appreciation to Professor Dr Goh Swee Hock for his invaluable guidance, constant supervision, intellectual advice and constructive criticisms throughout the course of this study and during the preparation of this thesis. I also wish to thank Dr. Chuah Cheng Hock and Professor Yong Hoi Sen for their supervision during Professor Goh's sabbatical leave. I also benefited from Dr. Chuah's supervision and help towards the later part of this study.

My sincere thanks and deepest gratitude are also extended to Mr. Lee Han Lin of the Institute of Medical Research, Kuala Lumpur for guidance on bioassay work involving Aedes aegypti. I am also grateful to Mr. K. T. Wong from the Pharmacology Department, National University of Singapore for conducting in vitro bioassays. Many thanks also go to Gnanamalar Panandam for typing this thesis. To K. Yoganathan, Lee Kee Huat, Ooi Kay Eng. Hew Nam Fong and Dr. Richard Wong thank you for all the help you have given me throughout my course of study.

I also wish to thank my parents, my two sisters Evelyn and Susan and my two sons Joey and Lance for their patience, tolerance and moral support. To them I dedicate this work. Finally, I would like to thank my husband Jegak Uli for encouraging me to undertake this study. I also thank him for collecting plant samples and for preparation of this thesis for without him I would never have achieved this piece of work.

ABSTRACT

Chemical and cytotoxicity studies were made on twelve plant species of the genus *Goniothalamus* and one plant species each of the genera *Disepalum* and *Mezzetia* (Annonaceae). These plants were mostly used as insect repellents by the Iban community in Sarawak, East Malaysia.

The chemical investigations on the plant species covered larvicidal principles such as annonaceous acetogenins, styrylpyrones, flavonoids, alkaloids and essential oils (mainly sesquiterpenes). Detail studies were carried out on four Goniothalamus species (G. andersonii, G. dolichocarpus, G. malayanus and G. velutinus), Disepalum anomalum and Mezzetia umbellata. Natural products were isolated using chromatographic techniques and identified using spectroscopic methods (including 2-D NMR). Eight other Goniothalamus species (G. ridleyi, G. macrophyllus, G. uvarioides, G. macranii, G. sinclairinius, G. gigantifolius, G. umbrosus and G. montanus) were screened for selected larvicidal principles using thin layer chromatography, gas chromatography and gas chromatography-mass spectrometry.

The four Goniothalamus species which were studied in detail provided styrylpyrone derivatives (+)-goniothalamin, (+)-goniothalamin epoxide, (+)-goniodiol, (+)-goniothalamol, two new natural products (-)-iso-5-deoxygoniopypyrone and (+)-5β-hydroxygoniothalamin and essential oils which mainly contained a mixture of sesquiterpenes. The two new natural products were characterised spectroscopically.

The structure assigned for (-)-iso-5-deoxygoniopypyrone is consistent with a stereoisomer of the known (+)-5-deoxygoniopypyrone previously isolated from *G. giganteus*. NOE difference spectra of the acetylated compound show NOE enhancement between the phenyl protons and H-7 and H-8, these effects suggesting that H-7 and the phenyl group are in close proximity. For an assumed preferential average conformation of the phenyl group in a quasi-equatorial position H-7 and H-8 will assume a transoid geometry. A coupling constant value of 10.4 Hz between H-7 and H-8 is in agreement of an almost antiparallel geometry of the two hydrogens. In contrast, a singlet is reported for H-8 in the compound (+)-5-deoxygoniopypyrone due to an almost orthogonal H-7 - H-8 orientation. This new compound was synthesised from the *erythro*-(6R, 7S, 8S)-goniodiol using a catalytic amount of DBU in THF.

The structure of (+)-5β-hydroxygoniothalamin was assigned using ¹H-¹H COSY, ¹H-¹³C HETCOR and NOE difference spectra. A coupling constant value of 2.9 Hz between H-5 and H-6 indicates a dihedral angle of approximately 50°. Moreover, the acetate derivative also shows a similar J_{5,6} coupling of 2.9 Hz. There is no NOE enhancement between H-7 and H-5. This new compound and its diastereomer (+)-5α-hydroxygoniothalamin were synthesised by reacting

goniothalamin with selenium dioxide and refluxing in dioxane for three and a half

Goniothalamus dolichocarpus furnished two very cytotoxic flavonoids naringenin and pinocembrin. The four Goniothalamus species also provided very cytotoxic annonaceous acetogenins, annonacin and goniothalamicin along with the phenanthrene lactam, aristolactam BII and the dioxoaporphine ouregidione. Annonacin and aristolactam BII were also present in Mezzetia umbellata. Disepalum anomalum gave a common oxoaporphine liriodenine and a new annonaceous C-39 acetogenin was found for the first time in this genus. The new natural product disepalin has a mono-THF ring with an adjacent acetate group at the C-20 position. The structural assignments of this compound were determined using 2D-NMR (HMBC, COSY, H-C HETCOR, NOESY), NOE difference spectra and mass spectrometric techniques. Stereochemical assignments were by correlations with reported NMR data in addition to the new NMR data obtained.

The eight other *Goniothalamus species* were also found to have (+)-goniothalamin, (+)-goniothalamin epoxide, (+)-goniothalamin, (+)-goniodiol, (+)-

v

goniodiol diacetate and the complex mixture of essential oils containing sesquiterpenes. Other natural products such as (-)-iso-5-deoxygoniopypyrone, naringenin and pinocembrin, aristolactam BII and ouregidione were also found to be present in some of these plants. Annonacin was detected only in *Goniothalamus macranii*. The various styrylpyrones, alkaloids, flavonoids and acetogenins isolated were tested on the larvae of *Aedes aegypti* using the WHO (1981a) standard procedures with slight modifications. The pure compounds tested showed moderate to good larvicidal activity, the most cytotoxic compounds being naringenin and annonacin (LCs0 values 3.7 and 9.5 µg/ml respectively).

Crude stem bark extracts of all twelve Goniothalamus species, Disepalum anomalum and Mezzetia umbellata were screened for their larvicidal activity against the larvae of Aedes aegypti. Most crude hexane extracts were moderately cytotoxic except for that from G. gigantifolius, while that of Disepalum anomalum and Mezzetia umbellata mildly cytotoxic. The crude ethanol extracts of G. sinclairinius and G. montanus were not cytotoxic to the larvae of Aedes aegypti, while the rest of the extracts showed larvicidal activity. The mosquito larvae of Aedes aegypti were susceptible to the methanol-soluble fractions partitioned from the ethanol extracts and good activity was found. Preliminary in vitro cytotoxicity screening against P388 cell lines was also carried out on the crude extracts and some bioactivity was detected which corroborates the presence of bioactive acetogenins and some of the styrylpyrone derivatives.

TABLE OF CONTENTS

r	AGE
Acknowledgements	ii
Abstract	111
Table of Contents	vii
List of Tables	vi
List of Tables	XI
List of Figures	xiv
I int of Plates	-xviii
Acronyms/Abbreviations used in text	xix
Acronyms/Addreviations used in text	
CHAPTER ONE: INTRODUCTION	1
1.1 On incompres on Carliny	1
1.2. DOTANY DISTRIBUTION AND ETHNOROTANY OF GENERA STUDIED	4
1.2 I Annonaceae family	2
1.2.1 Annonaceae jamity	3
1.2.2 The genus Goniothalamus	5
1.2.2.1 Goniothalamus andersonii	7
1.2.2.2 Goniothalamus dolicnocarpus 1.2.2.3 Goniothalamus malayanus	7
1.2.2.3 Goniothalamus maiayanus	10
1.2.2.4 Goniothalamus vetutinus 1.2.2.5 Goniothalamus macrophyllus	10
1.2.2.6 Goniothalamus macrophytius 1.2.2.6 Goniothalamus ridleyi	11
1.2.2.6 Goniothalamus rialeyi 1.2.2.7 Goniothalamus uvarioides	11
1.2.2.8 Goniothalamus unbrosus	13
1.2.3 The Genus Mezzetia	13
1.2.3.1 Mezzetia umbellata	14
1.2.4 The Gamus Disanglum	14
1.2.4.1 Disepalum anomalum	17
1.2. CHEMISTRY OF THE ANNONACEAE: A LITERATURE REVIEW	17
1.3.1 Non-alkaloidal constituents of Annonaceae	17
1 2 1 1 Towns and a semantic compounds	10
Ditarranae	
Tritamonas starole and sanonins	17
A comptic compounds	20
1 2 2 411 -1-14	20
Overnombines	20
4.6 Di	23
1.2.2.1 Conjetholomin	
1.2.4.4	27
1.2.4.1. Standard alonger of acetogening	
A A categorine with adjacent his-tetrahydrofuran ring	40
D. Mana tatachuda furan annonaceous acetogenins	50
1.3.4.2 Extraction, isolation and dosage methods.	30

1.3.4.3 Structural elucidation of acetogenins	57
1.3.4.5 Streetina electration of accognition. 1.3.4.5 Biosynthesis	63
1.3.4.5 Biosynthesis	67
and a second state of the commence of the comm	74
1.2.5. A	/0
1.3.7 Compounds isolated from the genera Disepalum and Mezzetia	79
1.3.7 Compounds isolated from the general 2 top allow a series and a series as	01
CHAPTER TWO: BIOASSAY TECHNIQUES	01
.1 THE PURPOSE OF BIOLOGICAL ASSAYS	81
2. A NEED TO STUDY BIOACTIVITIES OF NATURAL PRODUCTS	82
2 DENOTETOR PLOASEAVS COMMONI VIISED - A REVIEW	84
2.2.1. D.:	82
	86
2.2.2. I (duelaward): A biogesmy for inhibitors and promotors of plant growth	
(McLaughlin et al. 1991)	88
2.2.4. The support billing an magistance of magazita langua to insecticides tests	
(WHO/VPC/81 807)	89
A Establishing the hore line	89
B. Procedure for carrying out bioassays C. Results.	07
C. Results D. Estimation of the median effective dose	92
D. Estimation of the median effective dose E. Fitting a probit regression line	94
E. Fitting a probit regression line	98
2.5 BIOACTIVITIES OF ACETOGENINS FROM THE GENUS GONOTHALAMUS	101
BIOACTIVITIES OF ACETOGENING FROM THE GENES COMMEMBER OF	105
CHAPTER THREE: EXPERIMENTAL	105
3.1 PLANT MATERIALS	105
2 INCTRINGUIS	105
2.2.1. Malting points	105
2.2.2 Ontical notations	103
2.2.2. Infrared enactroscomy	105
2.2.4. Illumidat enactroscom	103
2.2.5 NMP anastroscomy	103
3.2.6 Mass spectroscom	100
2.2.7 Uigh norformance liquid chromatography	100
2.2.9. Can abrawatagraphy	100
2.2.0. Can Chromatography Mass Spectrometry	100
2. CURRELLITORS ARTHORS	107
2.2.2 This lower abromatography	100
3 3 3 Propagative Inver chromatography (PLC)	100
2.2.4. Contributed this lower chromatography (Chromatotron)	109
2.2.5.5	110
A DIOACCAY PROCEDURES	110
2.4.1. Culturing of Andre geometi mosquitoes	110
2.4.2. December for assuming out the susceptibility or resistance of mosquito larvae to	
!	111
2.4.2. December for assembled out on in vitro anti-cancer bioassay	112
2.5. Exemplication of control NDS	113
A C T	114
2.6.1. I - I - i - v - f bio activo principlos from Gonjothalamus andersonii	114
2 C 2 I - I - I C L	110
3.6.5 Isolation of bioactive principles from Gondonatamus vetatimas	124
3.0.3 Isolation of bioactive principles from Diseptium anomalum	

3.6.6 Isolation of bioactive principles from Mezzetia umbellata	. 126
3.6.7 Gas chromatography screening of the stem bark of the various Gonioinalamus species	. 12/
3.7 SYNTHESES	. 128
3.7.1 Synthesis of goniothalamin epoxide and isogoniothalamin epoxide	. 128
3.7.2 Synthesis of goniodiol	. 128
3.7.2 Synthesis of goniodiol	. 129
3.7.4 Synthesis of goniodial diacetate	. 129
3 7 5 Synthesis of (-)-iso-5-degrygonionynyrone	. 129
3.7.6. Synthesis of 5 β-hydroxygoniothalamin and 5α-hydroxygoniothalamin	. 129
2.7.7. Acaptation of 5 c. hydroxygoniothalamin and 5B-hydroxygoniothalamin	. 130
3.7.8 Sibilation of disensiin	. 130
2.7.0 Application of disonalin	. 131
3.7.10 Acetylation of (-)-iso-5-deoxygoniopypyrone	. 131
CHAPTER FOUR: RESULTS AND DISCUSSION-LARVICIDAL PRINCIPLES	132
	. 132
4.1 ISOLATION OF LARVICIDIAL PRINCIPLES FROM THE STEM BARK OF THE VARIOUS SPECIES	
STUDIED	132
4.2 ISOLATION OF LARVICIDAL PRINCIPLES FROM THE STEM BARK OF VARIOUS INSECTICIDAL	122
PLANTS FROM THE ANNONACEAE FAMILY	133
4.2.1 Goniothalamus andersonii	. 133
4.2.2 Goniothalamus dolichocarpus	. 134
4.2.3 Gonjothalamus malayanus	134
4.2.4 Goniothalamus velutinus	133
4.2.5 Mezzetia umbellata	135
4.2.6 Disepalum anomalum	135
4.2.7 Characterisation of GA-1 as goniothalamin	136
4.2.9. Characterisation of G.4.2 as (+)-agniothalamin enoxide	144
4.2.0. Characterisation of (+) isogoniothalamin enoxide	149
4.2.10 Synthesis of goniothalamin enoxide and isogoniothalamin epoxide	138
4.2.1.1 Characterisation of GA-A as (+)-ganiedial	138
4.2.11.1 Synthesis of goniodiol	165
4.2.12 Characterisation of GA-5 as (+)-goniodiol diacetate	1/4
4.2.13 Characterisation of LB5 as pinocembrin	180
4.2.1.4 Champatania ation of I P6 as navinganin	188
4.2.15 Characterisation of EELA 25 as (-)-iso-5-deoxygoniopypyrone	197
4.2.15.1 Synthesis of (-)-iso-3-decoxygoniopypytotic 4.2.16 Characterisation of GMg and GV9b as (+)-goniothalenol	213
4.2.16.1 Synthesis of goniothalenol	224
4.2.16.1 Synthesis of goniothaletot	223
4.2.19 () 58 Anatomagniothalamin	231
4.2.18.1 Syntheses of 5β-hydroxygoniothalamin and 5α-hydroxygoniothalamin	236
4.2.10 (+)-50-Hydroxygoniothalamin	238
3 2 19 1 Mechanism of reaction	248
4.2.20 () See Agatomygoniothalamin	249
4.2.2.1. Characterisation of LRMF.8b. GMC KHP13b and GV260R as (+)-annonacin	233
4.2.22 Characterization of E7 as a new acatogonin disensiin	2/0
4.2.22 Characterisation of GM5-7 and GV3h-2 as aristolactam RII	289
4 2 24 Characterisation of LS13-4v-13 as liriodenine	290
4.2.25 Characterisation of GV26SF2 and GMf as ouregidione	301
CHAPTER FIVE: RESULTS AND DISCUSSION-BIOASSAY RESULTS	
CHAPTER FIVE: RESULTS AND DISCUSSION-BIOASSAT RESULTS	211
5.1. INTRODUCTION	211
6.2 DIOACCAY OF CRUIDE BLANT EYTRACTS	311
5.3. LARVICIDAL ACTIVITY FOR CRUDE PLANT EXTRACTS OF VARIOUS PLANT SPECIES OF THE	212
GENERA GONIOTHALAMUS, DISEPALUM AND MEZZETIA	312
5.3.1. Crude hexane extracts of the stem bark from the plants of the genera Goniothalamus,	212
Disapplym and Morratia	313

5.3.1.1 Goniothalamus andersonii	314
5.3.1.2 Conjuthalamus dalichocarnus	314
5.2.1.2 Caniathalamus valutimus	314
5.3.1.4 Goniothalamus malayanus	314
5.2.1.5 Conjuthalamus magraphyllus	313
5.2.1.6 Carrieth alamon widleni	313
5.2.1.7 Conjuthalamus ajaantifolius	310
5.2.1.9 Contathalamus sinclairinius	310
5.3.1.9 Goniothalamus montanus	310
5.3.1.10 Goniothalamus macranii	317
5.3.1.11 Goniothalamus macrami	317
5.3.1.12 Disepalum anomalum	317
5 2 1 12 Marratia umballata	310
5.2.2 Conclusions	318
5.2.2 Crude athanol extracts of the stem bark from the plants of the genera Goniothalamus,	
Disanglum and Mazzatia	320
5 2 2 1 Conjethalamus andersonii	321
5.2.2.2 Conjust alamus delichecarnus	321
5.2.2.2 Coming the lamping resolutions	321
5.2.2.4 Conjethelamus malayanus	321
5 2 3 5 Conjuthalamus macrophyllus	344
5.2.2.6 Caniathalamus vidlavi	322
5 3 3 7 Conjothalamus ajaantifolijus	322
5 2 2 9 Conjethalamus singlairinius	322
5.2.2.0 C-wieth-lamus montanus	323
5 3 3 10 Conjothalamus macranii	323
5 2 2 11 Conjuthalamus marioides	323
5 3 3 12 Marzetia umbellata	323
5.3.3.13 Disenglum anomalum	324
5.3.4 Conclusions	324
5.2.5. Could make and caluble fractions of the ethanol bark extracts of the genera	
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera	326
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalamus, Mezzetia and Disepalum	326
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalamus, Mezzetia and Disepalum 5.3.5.1 Goniothalamus andersoni	326 328 328
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalamus, Mezzetia and Disepalum	326 328 328
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalamus, Mezzetia and Disepalum. 5.3.5.1 Goniothalamus anderroni. 5.3.5.2 Goniothalamus doictocarpus. 5.3.5.3 Goniothalamus velutinus	326 328 328 328
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalamus, Mezzetia and Disepalum	326 328 328 329 329
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalams, Mezzetia and Disepalum. 5.3.5.1 Goniothalamus anderroni. 5.3.5.2 Goniothalamus dolichocarpus. 5.3.5.3 Goniothalamus selutinus. 5.3.5.4 Goniothalamus selutinus. 5.3.5.5 Goniothalamus malayanus.	326 328 328 328 329 329
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalamus, Mezzetia and Disepalum 5.3.5.1 Goniothalamus andersonii. 5.3.5.2 Goniothalamus dolichocarpus. 5.3.5.3 Goniothalamus andupanus 5.3.5.4 Goniothalamus malayanus 5.3.5.5 Goniothalamus marcoppyllus. 5.3.5.6 Goniothalamus ridleyi.	326 328 328 328 329 329 329 330
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalams. Mezzetia and Disepalum. 5.3.5.1 Goniothalams and derronii. 5.3.5.2 Goniothalams volichocarpus. 5.3.5.3 Goniothalams velutinus. 5.3.5.4 Goniothalams unlayarus. 5.3.5.5 Goniothalams marcaphyllus. 5.3.5.6 Goniothalams marcaphyllus. 5.3.5.7 Goniothalams gigantifolius.	326 328 328 329 329 329 330 330
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalamus, Mezzetia and Disepalum 5.3.5.1 Goniothalamus andersonii. 5.3.5.2 Goniothalamus dolichocarpus. 5.3.5.3 Goniothalamus malayanus 5.3.5.4 Goniothalamus malayanus 5.3.5.5 Goniothalamus malayanus 5.3.5.5 Goniothalamus ridleyi 5.3.5.6 Goniothalamus ridleyi 5.3.5.7 Goniothalamus ridleyi 5.3.5.8 Goniothalamus sinclairinius	326 328 328 329 329 329 330 330 330
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalams Mezzetia and Disepalum. 5.3.5.1 Goniothalams and chronii. 5.3.5.2 Goniothalams valiothocarpus. 5.3.5.3 Goniothalams velutinus. 5.3.5.4 Goniothalams maraphyllus. 5.3.5.5 Goniothalams maraphyllus. 5.3.5.6 Goniothalams ridey. 5.3.5.6 Goniothalams sinclarivius. 5.3.5.7 Goniothalams sinclarivius. 5.3.5.8 Goniothalams sinclarivius. 5.3.5.8 Goniothalams sinclarivius.	326 328 328 329 329 329 330 330 330
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalamus, Mezzetia and Disepalum. 5.3.5.1 Goniothalamus andersonii 5.3.5.2 Goniothalamus solichocarpus. 5.3.5.3 Goniothalamus malayanus. 5.3.5.4 Goniothalamus malayanus. 5.3.5.5 Goniothalamus marcophyllus. 5.3.5.6 Goniothalamus ridleyi 5.3.5.7 Goniothalamus sirileyi 5.3.5.8 Goniothalamus sirileyi 5.3.5.9 Goniothalamus sirileivi	326 328 328 329 329 329 330 330 330 330
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalamus, Mezzetia and Disepalum 5.3.5.1 Goniothalamus andersonii. 5.3.5.2 Goniothalamus dolichocarpus. 5.3.5.3 Goniothalamus malayanus 5.3.5.4 Goniothalamus malayanus 5.3.5.5 Goniothalamus marcappyllus 5.3.5.6 Goniothalamus ridleyi 5.3.5.7 Goniothalamus gigantifolius 5.3.5.9 Goniothalamus gigantifolius 5.3.5.9 Goniothalamus montanus 5.3.5.9 Doniothalamus montanus 5.3.5.1 Disepalum anomalum 5.3.6 Conclusions	326 328 328 329 329 329 330 330 330 330
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalamus, Mezzetia and Disepalum. 5.3.5.1 Goniothalamus andersoni. 5.3.5.2 Goniothalamus andersoni. 5.3.5.3 Goniothalamus welutinus. 5.3.5.4 Goniothalamus malayanus. 5.3.5.5 Goniothalamus malayanus. 5.3.5.5 Goniothalamus ridleyi. 5.3.5.6 Goniothalamus ridleyi. 5.3.5.7 Goniothalamus sinclarinius. 5.3.5.8 Goniothalamus sinclarinius. 5.3.5.9 Goniothalamus sinclarinius. 5.3.5.9 Goniothalamus sinclarinius. 5.3.5.9 Goniothalamus sinclarinius. 5.3.5.0 Mezzetia umbellata 4.3.5.11 Disepalum anomalum. 5.3.6 Conclusions.	326 328 328 329 329 330 330 330 331 331
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalamus, Mezzetia and Disepalum. 5.3.5.1 Goniothalamus andersoni. 5.3.5.2 Goniothalamus andersoni. 5.3.5.3 Goniothalamus welutinus. 5.3.5.4 Goniothalamus malayanus. 5.3.5.5 Goniothalamus malayanus. 5.3.5.5 Goniothalamus ridleyi. 5.3.5.6 Goniothalamus ridleyi. 5.3.5.7 Goniothalamus sinclarinius. 5.3.5.8 Goniothalamus sinclarinius. 5.3.5.9 Goniothalamus sinclarinius. 5.3.5.9 Goniothalamus sinclarinius. 5.3.5.9 Goniothalamus sinclarinius. 5.3.5.0 Mezzetia umbellata 4.3.5.11 Disepalum anomalum. 5.3.6 Conclusions.	326 328 328 329 329 330 330 330 331 331
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalamus, Mezzetia and Disepalum 5.3.5.1 Goniothalamus andersonii. 5.3.5.2 Goniothalamus solichocarpus. 5.3.5.3 Goniothalamus malayanus 5.3.5.4 Goniothalamus malayanus 5.3.5.5 Goniothalamus malayanus 5.3.5.5 Goniothalamus ridleyi 5.3.5.5 Goniothalamus ridleyi 5.3.5.7 Goniothalamus sinclairinius 5.3.5.9 Goniothalamus sinclairinius 5.3.5.9 Goniothalamus montanus 5.3.5.1 Disepalum anomalum 5.3.6 Conclusions 5.3.6 Conclusions 5.3.6 Conclusions 5.3.6 Conclusions 5.3.6 Conclusions	326 328 328 329 329 330 330 330 331 331
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalamus, Mezzetia and Disepalum 5.3.5.1 Goniothalamus andersonii. 5.3.5.2 Goniothalamus aloichocarpus. 5.3.5.3 Goniothalamus malayanus 5.3.5.4 Goniothalamus malayanus 5.3.5.5 Goniothalamus malayanus 5.3.5.5 Goniothalamus ridleyi 5.3.5.7 Goniothalamus ridleyi 5.3.5.7 Goniothalamus gigantifolius 5.3.5 Goniothalamus montanus 5.3.5 Goniothalamus montanus 5.3.5.1 Disepalum anomalum 5.3.5.1 Disepalum anomalum 5.3.6 Conclusions 5.4 LaRVICIDAL ACTIVITY OF SOME COMPOUNDS ISOLATED FROM THE VARIOUS PLANT SPECIES STUDIED	326 328 328 329 329 329 330 330 331 331 331
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalamus, Mezzetia and Disepalum 5.3.5.1 Goniothalamus andersonii. 5.3.5.2 Goniothalamus aloichocarpus. 5.3.5.3 Goniothalamus malayanus 5.3.5.4 Goniothalamus malayanus 5.3.5.5 Goniothalamus malayanus 5.3.5.5 Goniothalamus ridleyi 5.3.5.7 Goniothalamus ridleyi 5.3.5.7 Goniothalamus gigantifolius 5.3.5 Goniothalamus montanus 5.3.5 Goniothalamus montanus 5.3.5.1 Disepalum anomalum 5.3.5.1 Disepalum anomalum 5.3.6 Conclusions 5.4 LaRVICIDAL ACTIVITY OF SOME COMPOUNDS ISOLATED FROM THE VARIOUS PLANT SPECIES STUDIED	326 328 328 329 329 329 330 330 331 331 331
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalamus, Mezzetia and Disepalum. 5.3.5.1 Goniothalamus andersonii. 5.3.5.2 Goniothalamus andersonii. 5.3.5.3 Goniothalamus subutinus 5.3.5.4 Goniothalamus malayanus 5.3.5.5 Goniothalamus malayanus 5.3.5.5 Goniothalamus ridleyi 5.3.5.7 Goniothalamus sigantifolius 5.3.5.8 Goniothalamus sinclarinius 5.3.5.9 Goniothalamus sinclarinius 5.3.5.9 Goniothalamus sinclarinius 5.3.5.1 Disepalum anomalum 5.3.5.1 Disepalum anomalum 5.3.6 Conclusions 5.4 LARVICIDAL ACTIVITY OF SOME COMPOUNDS ISOLATED FROM THE VARIOUS PLANT SPECIES STUDIED STUDIED S.5 CONCLUSIONS	326 328 328 329 329 330 330 331 331 333 334
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalamus, Mezzetia and Disepalum. 5.3.5.1 Goniothalamus andersonii. 5.3.5.2 Goniothalamus andersonii. 5.3.5.3 Goniothalamus subutinus 5.3.5.4 Goniothalamus malayanus 5.3.5.5 Goniothalamus malayanus 5.3.5.5 Goniothalamus ridleyi 5.3.5.7 Goniothalamus sigantifolius 5.3.5.8 Goniothalamus sinclarinius 5.3.5.9 Goniothalamus sinclarinius 5.3.5.9 Goniothalamus sinclarinius 5.3.5.1 Disepalum anomalum 5.3.5.1 Disepalum anomalum 5.3.6 Conclusions 5.4 LARVICIDAL ACTIVITY OF SOME COMPOUNDS ISOLATED FROM THE VARIOUS PLANT SPECIES STUDIED STUDIED S.5 CONCLUSIONS	326 328 328 329 329 330 330 331 331 333 334
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalamus, Mezzetia and Disepalum. 5.3.5.1 Goniothalamus andersonii 5.3.5.2 Goniothalamus andersonii 5.3.5.3 Goniothalamus andersonii 5.3.5.4 Goniothalamus malayanus. 5.3.5.5 Goniothalamus malayanus. 5.3.5.6 Goniothalamus ridleyi 5.3.5.6 Goniothalamus ridleyi 5.3.5.7 Goniothalamus sinclarinius. 5.3.5.8 Goniothalamus sinclarinius. 5.3.5.9 Goniothalamus sinclarinius. 5.3.5.9 Goniothalamus sinclarinius. 5.3.5.9 Goniothalamus sinclarinius. 5.3.5.0 Mezzetia umbellata. 4.3.5.11 Disepalum anomalum 5.3.6 Conclusions. 5.4 LARVICIDAL ACTIVITY OF SOME COMPOUNDS ISOLATED FROM THE VARIOUS PLANT SPECIES STUDIED. 5.5 CONCLUSIONS. REFERENCES.	326 328 328 329 329 329 330 330 331 331 334 334
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalamus, Mezzetia and Disepalum. 5.3.5.1 Goniothalamus andersonii 5.3.5.2 Goniothalamus andersonii 5.3.5.3 Goniothalamus andersonii 5.3.5.4 Goniothalamus malayanus. 5.3.5.5 Goniothalamus malayanus. 5.3.5.6 Goniothalamus ridleyi 5.3.5.6 Goniothalamus ridleyi 5.3.5.7 Goniothalamus sinclarinius. 5.3.5.8 Goniothalamus sinclarinius. 5.3.5.9 Goniothalamus sinclarinius. 5.3.5.9 Goniothalamus sinclarinius. 5.3.5.9 Goniothalamus sinclarinius. 5.3.5.0 Mezzetia umbellata. 4.3.5.11 Disepalum anomalum 5.3.6 Conclusions. 5.4 LARVICIDAL ACTIVITY OF SOME COMPOUNDS ISOLATED FROM THE VARIOUS PLANT SPECIES STUDIED. 5.5 CONCLUSIONS. REFERENCES.	326 328 328 329 329 329 330 330 331 331 334 334
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalamus, Mezzetia and Disepalum. 5.3.5.1 Goniothalamus andersonti 5.3.5.2 Goniothalamus andersonti 5.3.5.3 Goniothalamus andersonti 5.3.5.4 Goniothalamus malayanus. 5.3.5.4 Goniothalamus malayanus. 5.3.5.5 Goniothalamus ridleyi 5.3.5.6 Goniothalamus ridleyi 5.3.5.7 Goniothalamus sinclariritus. 5.3.5.8 Goniothalamus sinclariritus. 5.3.5.9 Goniothalamus sinclariritus. 5.3.5.9 Goniothalamus sinclariritus. 5.3.5.9 Goniothalamus sinclariritus. 5.3.5.0 Mezzetia umbellata. 4.3.5.11 Disepalum anomalum. 5.3.6 Conclusions. 5.4 LARVICIDAL ACTIVITY OF SOME COMPOUNDS ISOLATED FROM THE VARIOUS PLANT SPECIES STUDIED. 5.5 CONCLUSIONS. REFERENCES.	326 328 328 329 329 330 330 331 331 337 3346
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalams. Mezzetia and Disepalum. 5.3.5.1 Goniothalams and officencerpus. 5.3.5.2 Goniothalams maloyanus. 5.3.5.3 Goniothalams maloyanus. 5.3.5.4 Goniothalams maloyanus. 5.3.5.5 Goniothalams maloyanus. 5.3.5.5 Goniothalams ridley. 5.3.5.6 Goniothalams ridley. 5.3.5.7 Goniothalams sinclarinius. 5.3.5.8 Goniothalams sinclarinius. 5.3.5.9 Goniothalams sinclarinius. 5.3.5.9 Goniothalams sinclarinius. 5.3.5.0 Mezzeta umbellata. 4.3.5.11 Disepalum anomalum. 5.3.6 Conclusions. 5.4 LARVICIDAL ACTIVITY OF SOME COMPOUNDS ISOLATED FROM THE VARIOUS PLANT SPECIES STUDIED. 5.5 CONCLUSIONS. REFERENCES. COMPOUND/STRUCTURE INDEX.	326 328 328 329 329 329 330 330 330 331 331 334 334 334
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalamus, Mezzetia and Disepalum. 5.3.5.1 Goniothalamus andersonii. 5.3.5.2 Goniothalamus andersonii. 5.3.5.3 Goniothalamus andersonii. 5.3.5.4 Goniothalamus malayanus. 5.3.5.5 Goniothalamus malayanus. 5.3.5.5 Goniothalamus ridleyi 5.3.5.6 Goniothalamus sinclarinius. 5.3.5.7 Goniothalamus sinclarinius. 5.3.5.8 Goniothalamus sinclarinius. 5.3.5.9 Goniothalamus sinclarinius. 5.3.5.9 Goniothalamus sinclarinius. 5.3.5.0 Mezzetia umbellata 4.3.5.11 Disepalum anomalum 5.3.6 Conclusions. 5.3.6 Conclusions. 5.3.6 Conclusions. 5.5 CONCLUSIONS. REFERENCES. COMPOUND/STRUCTURE INDEX.	326 328 328 328 329 329 329 329 329 330 330 331 331 331 331 331 331 331 331
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalamus Mezzetia and Disepalum. 5.3.5.1 Goniothalamus and circonii. 5.3.5.2 Goniothalamus valichocarpus. 5.3.5.3 Goniothalamus valichocarpus. 5.3.5.3 Goniothalamus malayanus. 5.3.5.5 Goniothalamus macraphyllus. 5.3.5.6 Goniothalamus radiey. 5.3.5.7 Goniothalamus sinclarivius. 5.3.5.8 Goniothalamus sinclarivius. 5.3.5.8 Goniothalamus sinclarivius. 5.3.5.9 Goniothalamus sinclarivius. 5.3.5.10 Mezzetia umbellata. 4.3.5.11 Disepalum anomalum. 5.3.6 Conclusions. 5.4 Laxviculal Activity OF SOME COMPOUNDS ISOLATED FROM THE VARIOUS PLANT SPECIES STUDIED. 5.5 CONCLUSIONS. REFERENCES. COMPOUND/STRUCTURE INDEX	3268 3288 3288 3298 3299 3299 3299 3300 3300 3310 331 331 331 331 331 331
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalamus, Mezzetia and Disepalum. 5.3.5.1 Goniothalamus andersoni. 5.3.5.2 Goniothalamus solichocarpus. 5.3.5.3 Goniothalamus solichocarpus. 5.3.5.4 Goniothalamus solichocarpus. 5.3.5.5 Goniothalamus radieyu. 5.3.5.6 Goniothalamus radieyu. 5.3.5.6 Goniothalamus sinclarinius. 5.3.5.7 Goniothalamus sinclarinius. 5.3.5.8 Goniothalamus sinclarinius. 5.3.5.9 Goniothalamus sinclarinius. 5.3.5.9 Goniothalamus sinclarinius. 5.3.5.0 Mezzetia umbellata. 4.3.5.11 Disepalum anomalum. 5.3.6 Conclusions. 5.4 LARVICIDAL ACTIVITY OF SOME COMPOUNDS ISOLATED FROM THE VARIOUS PLANT SPECIES STUDIED. 5.5 CONCLUSIONS. REFERENCES. COMPOUND/STRUCTURE INDEX.	326 328 328 328 329 329 329 329 329 329 329 329 329 330 330 331 331 331 331 331 331 331 331
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalamus, Mezzetia and Disepalum. 5.3.5.1 Goniothalamus andersonii 5.3.5.2 Goniothalamus andersonii 5.3.5.3 Goniothalamus andersonii 5.3.5.4 Goniothalamus malayanus. 5.3.5.5 Goniothalamus radayanus. 5.3.5.5 Goniothalamus raderyiius. 5.3.5.6 Goniothalamus rideryi 5.3.5.7 Goniothalamus sinclarinius. 5.3.5.8 Goniothalamus sinclarinius. 5.3.5.9 Goniothalamus sinclarinius. 5.3.5.9 Goniothalamus sinclarinius. 5.3.5.9 Goniothalamus sinclarinius. 5.3.5.0 Mezzetia umbellata. 4.3.5.11 Disepalum anomalum. 5.3.6 Conclusions 5.4 LARVICIDAL ACTIVITY OF SOME COMPOUNDS ISOLATED FROM THE VARIOUS PLANT SPECIES STUDIED 5.5 CONCLUSIONS. REFERENCES. COMPOUND/STRUCTURE INDEX APPENDICES APPENDICS 4.7 PENDICS 4.7 PENDICS 4.7 PENDICS 4.7 PENDICS 4.7 PENDICS 4.7 PENDICS	326 328 328 328 328 328 328 329 329 329 329 330 330 331 331 331 331 331 331 331 331
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalamus, Mezzetia and Disepalum. 5.3.5.1 Goniothalamus anderroni. 5.3.5.2 Goniothalamus velutinus. 5.3.5.3 Goniothalamus velutinus. 5.3.5.4 Goniothalamus velutinus. 5.3.5.5 Goniothalamus maloyanus. 5.3.5.6 Goniothalamus sindoponus. 5.3.5.6 Goniothalamus sindoponus. 5.3.5.7 Goniothalamus sindoponus. 5.3.5.8 Goniothalamus sindoponus. 5.3.5.9 Goniothalamus sindoponus. 5.3.5 Goniothalamus sindoponus. 5.3.5 Goniothalamus sindoponus. 5.3.5 Goniothalamus sindoponus	326 328 328 329 329 329 329 329 330 330 331 333 331 333 334 335 335 335 335 335 335 335 335
5.3.5 Crude methanol soluble fractions of the ethanol bark extracts of the genera Goniothalamus, Mezzetia and Disepalum. 5.3.5.1 Goniothalamus andersoni. 5.3.5.2 Goniothalamus solichocarpus. 5.3.5.3 Goniothalamus solichocarpus. 5.3.5.4 Goniothalamus solichocarpus. 5.3.5.5 Goniothalamus radieyu. 5.3.5.6 Goniothalamus radieyu. 5.3.5.6 Goniothalamus sinclarinius. 5.3.5.7 Goniothalamus sinclarinius. 5.3.5.8 Goniothalamus sinclarinius. 5.3.5.9 Goniothalamus sinclarinius. 5.3.5.9 Goniothalamus sinclarinius. 5.3.5.0 Mezzetia umbellata. 4.3.5.11 Disepalum anomalum. 5.3.6 Conclusions. 5.4 LARVICIDAL ACTIVITY OF SOME COMPOUNDS ISOLATED FROM THE VARIOUS PLANT SPECIES STUDIED. 5.5 CONCLUSIONS. REFERENCES. COMPOUND/STRUCTURE INDEX.	326 328 328 329 329 329 329 329 330 330 331 333 331 333 334 335 335 335 335 335 335 335 335

LIST OF TABLES

Table 1. 1: Goniothalmus species available in Malaysia	. 5
T-LI- 1 2. Occurrence of excepterhines in Appenageae	21
Table 1. 3: Goniothalamin contents in different morphological parts of Goniothalamus species	33
Table 1. 4: Diagnostic ¹ H and ¹³ C NMR chemical shifts for the γ-lactone moiety	58
T-blad S. Diagnostic NIMP chemical chiffs of different subunits with various relative	
	60
Table 1 6: Cytotoxicity and antitumour activity of annonaceous acetogenins	71
man 1 1 7 1 1 1 2 1 1 1 1 2 1 1 1 1 1 1 1 1	
	13
	74
Table 2. 1: Some novel antitumour prescreens	07
Table 2. 2: A test of the toxicity of rotenone to Macrosiphoniella sanborni	94
Table 2. 3: Empirical probits and approximate expected probits for data of Table 2.2	97
Table 2. 4: Transformation of percentages to probits	,,
Table 2. 5: Comparison of observed and expected mortality in eye estimation for rotenone -	97
Macrosiphoniella sanborni test. Table 4. 1: Larvicidal principles present in the stem bark of various plant species studied	32
Table 4. 1: Larvicidal principles present in the stem bark of various plant species studied	37
Table 4. 2: ¹ H NMR (δ) assignments and J(Hz) values for goniothalamin (270 MHz, CDCl ₃)	31
Table 4. 3: Proton-proton connectivities obtained from H-H COSY experiment for	42
(+)-goniothalamin (270 MHz and 67.8 MHz)	72
Table 4. 4: Proton-proton connectivities obtained from H-H-OST experiment to (+)-goniothalamin (270 MHz and 67 s. MHz). Table 4. 4: Proton and carbon connectivities obtained from H-H-OST experiment for the control of the control o	42
(+)-goniothalamin (2/0 MHz and 6/.8 MHz)	
Table 4. 5: ¹³ C NMR (δ) assignments for goniothalamin (67.8 MHz, CDCl ₃)	45
Table 4. 6: ¹ H NMR (δ) assignments and coupling constant values (Hz) for goniothalamin epoxide 1	45
Table 4. 7: ¹ H - ¹ H connectivities (¹ H - ¹ H COSY) for goniothalamin epoxide	43
Table 4. 8: ¹ H - ¹³ C connectivities obtained from ¹ H - ¹³ C HETCOR experiment for	10
goniothalamin epoxide	40
Table 4. 9: ¹³ C NMR (δ) assignments for goniothalamin epoxide	47
Table 4. 10: ¹ H NMR (δ) assignments and J(Hz) values for (+)-isogoniothalamin epoxide	150
Table 4. 11: 'H - 'H connectivities ('H - 'H cOSY) for (+)-isogoniothalamin epoxide	156
Table 4. 12: ¹ H - ¹³ C connectivities (¹ H - ¹³ C HETCOR) for (+)-isogoniothalamin epoxide	156
Table 4. 13: ¹³ C NMR (δ) assignments for isogoniothalamin epoxide	56
Table 4 14. It NIAD (S) assignments and (Hz) values for (+)-ganiodial (270 MHz, CDCl ₂)	159
Table 4 15. III III connectivities (III - III COSV) for (+)-geniodial	160
Table 4 16: H-13C connectivities ('H-13C HETCOR) for (+)-goniodiol	100
Table 4, 17: 13C NMR (δ) assignments for (+)-goniodiol (67.8 MHz, CDCl ₃)	160
Table 4 19. LINIAR (8) assignments and I(Hz) values for three-goniodiol (270 MHz, CDCl ₃)	167
Table 4 19. H - H connectivities obtained from H - H COSY experiment for threo-goniodiol	167
Table 4 20, 1H 13C connectivities obtained from 1H 13C HETCOR experiment for	
three geniodial	167
Table 4. 21: ¹³ C NMR assignments (δ) for three-goniodiol (67.8 MHz, CDCl ₃)	172
Table 4.22. HANAR (8) assignments and I values (Hz) for goniodial diacetate	175
Table 4 23: U - U connectivities (U - U COSY) for goniodial diacetate	1/3
Table 4 24 H - 13C connectivities (H - 13C HETCOR) for goniodiol diacetate	1/3
Table 4. 25: ¹³ C NMR (δ) assignments for goniodiol diacetate (67.8 MHz, CDCl ₃)	175
T-11- 4.26. ILLNIAD (S) essignments and WHz) values for pinocembrin (270 MHz, CD ₂ COCD ₃)	181
Table 4. 26: 'H NMR (6) assignments and J(Hz) values for pinocentorin (276 MHz) Cosystems and J(Hz) Cosystems an	181
Table 4. 22: H - H connectivities obtained from H - 13C HETCOR experiment for pinocembrin	181

Table 4. 29: ¹³ C NMR assignments for pinocembrin (67.8 MHz, CD ₃ COCD ₃)	. 187
T-bl- 4, 20, 14 NMP (S) assignments and I/Hz) values for naringenin (270 MHz, CD ₂ COCD ₁)	. 189
Table 4.31: H - 'H connectivities obtained from 'H - 'H COSY experiment for naringenin	. 189
Table 4. 32: ¹ H - ¹³ C connectivities obtained from ¹ H - ¹³ C HETCOR experiment for naringenin	. 189
Table 4 33: 13C NMR assignments for paringenin (67.8 MHz, CD ₂ COCD ₃)	. 197
Table 4 34. H NMR assignments for (-)-iso-5-deoxygoniopypyrone acetate (270 MHz, CDCl ₃)	. 201
T-ble 4 25. H L connectivities obtained from H - H COSY experiment for	
() i f di-m-mana acatata	. 201
Table 4 36. H NMR assignments for (-)-iso-5-deoxygoniopypyrone (270 MHz, CDCl ₃)	. 207
(-)-iso-5-deoxygoniopypyrone acetate	. 207
Table 4 38. 13C NMR assignments for (-)-iso-5-deoxygoniopypyrone acetate (0/.8 MHz, CDCl3)	. 207
Table 4. 20. It NIMP (S) assignments and I(Hz) values for (+)-goniothalenol (270 MHz, CDCl ₃)	. 216
Table 4. 40: H- H connectivities (H-13C HETCOR) for (+)-goniothalenol	. 216
Table 4. 41: 'H - 'I'C connectivities ('H - 'I'C HETCOR) for (+)-goniothalenol	
(+)-goniothalenol	. 223
Table 4.42: ¹³ C NMR (8) assignments for goniothalenol (67.8 MHz, CDCl ₃)	. 223
Table 4 43. H NMR (δ) assignments for (+)-5β-hydroxygoniothalamin (270 MHz, CDCl ₃)	. 226
many 4 44 ltr ltrtimes shading from 'U 'U COSV experiment for (+)-58-	
hydroxygoniothalamin	. 226
hydroxygoniothalamin	. 227
Table 4.46: 13C NMR (8) assignments for (+)-58-hydroxygoniothalamin (67.8 MHz, CDCl ₃)	227
many and transport of the control of	
Table 4. 48: 'H- 'H connectivities obtained from 'H-'H COSY experiment for	233
Table 4 48: ¹ H ₂ ¹ H connectivities obtained from ¹ H - ¹ H COSY experiment for	
() 58 sectory/geniothalamin	233
Table 4. 49: ¹ H - ¹ C connectivities obtained from ¹ H - ¹ C CHETCOR experiment for	
(-)-5β-acetoxygoniothalamin	233
(-)-5β-acetoxygoniothalamin (67.8 MHz, CDCl ₃)	236
Table 4. 50: C NMR assignments for (-)-5p-acetoxygoniothalamin (7)-6 MHz, $\sigma = 0.00$, Table 4. 51: H NMR (δ) assignments and J values for (+)-5 α -hydroxygoniothalamin	
Table 4. 51: 'H NMR (6) assignments and J values for (+)-50-flydroxygorifodiadamin	. 239
1able 4. 52: 'H- 'H connectivities obtained from 'H-'H COSY experiment for	
Table 4. 52: H- H connectivities obtained from 11-11 COST experiment of	239
Table 4, 52: 'H- H connectivities obtained from 'H-'B'C HETCOR experiment for Table 4, 53: 'H-'B'C connectivities obtained from 'H-'B'C HETCOR experiment for	
(+)-5α-hydroxygoniothalamin	240
(+)-5α-hydroxygoniothalamin	246
Table 4. 54: ¹³ C NMR (δ) assignments for (+)-5α-hydroxygoniotnalamin (67.8 MHz, CDCl ₃)	250
Table 4. 55: ¹ H NMR (δ) assignments and J(Hz) values for (-)-5α-acetoxygoniothalamin	230
Table 4. 56: ¹ H- ¹ H connectivities obtained from ¹ H - ¹ H COSY experiment for	254
(-)-50-actoxygoniothalamin. Table 4. 57: ¹ H - ¹ C connectivities obtained from ¹ H - ¹ C HETCOR experiment for	234
Table 4. 57: 'H - 13C connectivities obtained from 'H - 13C HETCOR experiment for	254
() 5 a scetovy goniothalamin	254
Table 4 58: ¹³ C NMR (δ) assignments for (-)-5α-acetoxygoniothalamin (67.8 MHz, CDCl ₃)	254
The second secon	260
Table 4. 60: H - H connectivities obtained from H - H COSY experiment for (+)-annonacin	260
Table 4. 61: Structure and 'H- ¹ C connectivities obtained from 'H- 'H COSY experiment for (+)-annonacin Table 4. 61: Structure and 'H- ¹ C connectivities obtained from 'H- ¹ C HETCOR experiment	~~
for (+) apparagin	202
Table 4. 62: ¹³ C NMR (δ) assignments for (+)-annonacin (67.8 MHz, CDCl ₃)	270
Table 4, 63: Structure and proton NMR spectral data for disepalin (CDCl3, 500 MHz) and	
annonacin (CDCl2, 470 MHz) (McCloud et al., 1987)	277
m 11 4 64 Poster process compactivities obtained from 'H-'H COSV experiment for disepalin	287
¹ H - ¹³ C HETCOR and HMBC experiments for disepalin	287
(McCloud et al., 1987)	28

Table 4. 67: Structure and comparison of ¹ H NMR values of the acetate derivatives of disepalin	288
and annonacin	
Table 4. 68: ¹ H NMR assignments (δ) and J values (Hz) for aristolactam BII (270 MHz, CDCl ₃)	290
Table 4. 69: 'H - 'H connectivities obtained from 'H - 'H COSY experiment for aristolactam BII Table 4. 70: 'H - '3C connectivities obtained from 'H - '3C HETCOR experiment for	290
aristolactam BII	295
anstolaciam bil	295
Table 4. 71: ¹³ C NMR (δ) assignments for aristolactam BII (67.8 MHz, CDCl ₃)	207
Table 4. 72: ¹ H NMR (δ) assignments and J(Hz) values for liriodenine (270 MHz, CDCl ₃)	. 291 . 297
Table 4. 73: ¹ H - ¹ H connectivities (¹ H - ¹ H COSY) for liriodenine	
Table 4. 74: ¹³ C NMR (δ) assignments for liriodenine (67.8 MHz, CDCl ₃)	. 301
Table 4. 75: Structure and ¹ H NMR (δ) assignments and J(Hz) values for ouregidione in	
different solvents (270 MHz)	. 303
Table 4 76: 1H - 1H connectivities obtained from 1H - 1H COSY experiment for ouregidione	. 304
Table 4. 77: Proton and carbon connectivities and their ² J and ³ J interactions obtained from	
HMQC and HMBC experiments for ouregidione (CDCl ₃ , 270 MHz and 67.8 MHz)	. 305
Table 4. 78: ¹³ C NMR (δ) assignments for ouregidione	. 305
Table 5. 1: Larvicidal (Aedes aegypti) activity of crude hexane extract	. 313
Table 5. 1: Larvicidal (Aedes degypti) activity of crude flexalle extracts	320
Table 5. 2: Larvicidal (Aedes aegypti) activity of crude ethanol extracts	. 326
Table 5. 3: In vitro cytotoxicity activity of crude ethanol extracts	. 520
Table 5. 4: Larvicidal (Aedes aegypti) activity of methanol soluble fractions of crude ethanol	. 327
extracts	
Table 5. 5: In vitro cytotoxicity activity of methanol soluble fractions	
Table 5. 6: Larvicidal (Aedes gegypti) activity of compounds	. 534

LIST OF FIGURES

Figure 1. 1: Some recently identified oxoaporphines	22
Figure 1. 2: Structures of dihydropontevedrine and corydione	23
Figure 1. 3: Structures of some 4,5-dioxoaporphines	24
Figure 1. 4: Originally assigned and revised structures for pontevedrine	25
Figure 1. 5: Synthesis of aristolactam	20
Figure 1. 5. Synthesis of aristolactani	20
Figure 1. 7: Structures of some aristolactams	20
Figure 1. 7: Structures of some an isolactains	20
Figure 1. 9: Structures of five phenanthrene derivatives	30
Figure 1. 10: Structures of some aristolactam alkaloids	21
Figure 1. 11: Structure of (+)-goniothalamin	37
Figure 1. 11: Structure of (+)-gonitotial anim	34
Figure 1. 12: Synthesis of 6R-(+)-goniothalamin	35
Figure 1. 14: Syntheses of (6S)-goniothalamin and (6R)-goniothalamin	38
Figure 1. 15: Structures of some styrylpyrone derivatives	41
Figure 1. 16: Structures of five tetrahydrofuran subunits	42
Figure 1. 10. Structures of γ-lactone units of acetogenins	43
Figure 1. 18: Structure of outstactmone	44
Figure 1. 20: Structure of reticulatacin	45
Figure 1. 20: Structures of some novel bis- and mono-THF acetogenins	40
Figure 1. 22: Two novel acetogenins with epoxy rings	47
Figure 1. 23: Acetogenins with adjacent bis-THF rings	50
Figure 1. 24: Some mono-THF annonaceous acetogenins	54
Figure 1. 25: Structure of goniocin, a tri-THF acetogenin	54
Figure 1. 26: Structures of some non-adjacent bis-THF acetogenin	55
Figure 1. 27: Structures of some non-THF acetogenins.	62
Figure 1. 27: Structures of some non-TTT accognition	02
Figure 1. 29: The MTPA plane of an MTPA ester and H _{k,b,c} and H _{x,y,z} on the right and left sides of the plane, respectively	62
of the plane, respectively	02
Figure 1. 30: Model A for the determination of the absolute configuration of secondary alcohols	62
(Ohtani et al., 1991)	64
Figure 1. 31: Hypothesis for the biosynthesis of the THF rings of annonaceous accognitis. Figure 1. 32: Biogenetic pathways leading to the acetogenins from G. giganteus.	65
Figure 1. 32: Biogenetic pathways leading to the acetogenins from G. giganteus. Figure 1. 33: Biogenetic pathways leading to the acetogenins from G. giganteus.	66
Figure 1. 33: Biogenetic pathways leading to the acetogenins from G. giganieus	00
Figure 1. 34: A possible role of diepomuricanin in the biogenesis of solamin (Laprevote et al., 1992)	67
(Laprevote et al., 1992)	77
Figure 1. 35: Structures of (-)- anolobine, (-)-anonaine and ilriodenine	78
Figure 1. 36: Structures of goniopedaline, aristolactam AII and taliscanine	78
Figure 1. 37: Structures of velutinam and aristolactam Bit	79
Figure 1. 38: Structure of scorazanone Figure 1. 39: Structures of asimilobine and (-)-scoulerine	80
Figure 1. 39: Structures of asimilotine and (-)-scottlerine. Figure 2. 1: Relation between percentage kill of <i>Macrosiphoniella sanborni</i> and doses of	
Figure 2. 1: Relation between percentage kill of <i>Macrostphometa sanborni</i> and doses of rotenone, showing normal sigmoid curve represented by equation $Y = 2.25 + 4.01x$	
(Source: Finney, 1971: 30)	. 95
Figure 2. 2: Relation between probit of kill of <i>Macrosiphoniella sanborni</i> and dose of rotenone,	
showing probit regression line from equation Y = 2.25 + 4.01x (Source: Finney, 1971: 31)	. 96
Figure 2. 3: Styrylpyrones and their biological activities (McLaughlin et al., 1991).	. 99
Figure 2. 3: Styrylpyrones and their biological activities (McLaughini et al., 1991)	102
Figure 2. 4: Annonaceous acetogenins from Goniothalamus giganieus and their Cytotoxiches	116
Figure 3. 1: Scheme of extraction of the bark of Goniothalamus andersonii Figure 3. 2: Scheme of extraction of the bark of Goniothalamus dolichocarpus	119
Figure 3. 2: Scheme of extraction of the bark of Goniothalamus malayanus	121

	122
Figure 3. 4: Scheme of extraction for the bark of Goniothalamus velutinus	125
Figure 3. 5: Scheme of extraction for the bark of Disepalum anomalum	123
Figure 3. 5. Scheme of extraction for the bark of <i>Mezzetia umbellata</i>	126
Figure 4. 1: Structure of (+)-goniothalamin	130
Figure 4. 1: Structure of (+)-goniothalamin (270 MHz, CDCl ₃)	130
Figure 4 3: 'H = 'H COSV spectrum of (+)-goniothalamin	139
Figure 4. 4: ¹³ C NMR spectrum of (+)-goniothalamin (67.8 MHz, CDCl ₃)	140
Figure 4. 5: ¹ H - ¹³ C HETCOR spectrum of (+)-goniothalamin	141
Figure 4. 5: H = C HE PCOR spectrum of (+)-goniothalamin	143
Figure 4. 7: FIMS fragmentation of (+)-goniothalamin	144
Figure 4. 9. Structure of (+) goniotholomin enovide	144
Figure 4. 9: H NMR spectrum of (+)-goniothalamin enoxide (270 MHz, CDCl ₃)	146
Figure 4 10: H - H COSY spectrum of (+)-gonjothalamin epoxide	14/
Figure 4. 11: 13C NMP exectrum of (+)-goniothalamin enoxide	148
Figure 4 12. Structure of (+) inagoniothalamin enovide	149
Figure 4, 13: H NMR spectrum of (+)-isogoniothalamin epoxide (270 MHz, CDCl ₃)	151
Figure 4 14: H = H COSV spectrum of (+)-isogoniothalamin epoxide	152
Figure 4 15: ¹³ C NMR spectrum of (+)-isogoniothalamin epoxide (67.8 MHz, CDCL ₃)	153
Figure 4 16. LH 13C HETCOP enectrum of (+)-isogoniothalamin enoxide	154
Figure 4 17: FIMS spectrum of (+) isogoniothalamin enovide	155
Figure 4, 19: A perspective view of (+) isogoniothalamin enoxide	157
Figure 4. 20: Structure of (+) geniodial	158
Figure 4 21: 'H NMR spectrum of (+)-goniodiol (270 MHz, CDCl ₂)	101
Figure 4. 22: ¹ H- ¹ H COSY spectrum of (+)-goniodiol	162
Ti and Board of the second of	163
Figure 4. 23: "C NMR spectrum of (+)-goniodiol Figure 4. 24: 'H- ¹³ C HETCOR spectrum of (+)-goniodiol	164
Figure 4 26. Combasis of (+) againstic	100
Figure 4 26. Machanism for the synthesis of (+) ganiedial	100
Figure 4. 29: HNMP enectrum of three-ganiedial (270 MHz, CDCls)	168
Figure 4. 29: H-H COSY spectrum of threo-goniodiol	169
Figure 4. 30: ¹³ C NMR spectrum of <i>threo</i> -goniodiol (67.8 MHz, CDCl ₃)	170
Figure 4 21, IL IIC HETCOP spectrum of three-ganisdial	171
Figure 4. 32: Cyclisation of of threo-goniodiol and erythro-goniodiol	172
Figure 4. 33: Synthesis of (+)-goniopypyrone	173
Figure 4 24. Structure of (+) conjudial discretate	174
Figure 4. 34: Structure of (+)-goniodiol diacetate (270 MHz, CDCl ₃)	176
Figure 4. 36: 'H-'H COSY spectrum of (+)-goniodiol diacetate	177
Figure 4. 37: ¹³ C NMR spectrum of (+)-goniodiol diacetate (67.8 MHz, CDCl ₃)	178
Figure 4. 37: C NMR spectrum of (+)-goniodiol diacetate (07.6 MHz, CDC13)	179
Figure 4. 39: Structure of pinocembrin	180
Figure 4. 40: ¹ H NMR spectrum of pinocembrin (270 MHz, CD ₃ COCD ₃)	182
Figure 4. 40: 'H NMK spectrum of pinocembrin (270 MHz, CD ₃ COCD ₃)	183
Figure 4. 41: 'H-'H COSY spectrum of pinocemorin	184
Figure 4. 42: "C NMR spectrum of pinocembrin (67.8 Mriz, CD ₃ COCD ₃)	185
Figure 4. 43: 'H-'C HETCOR spectrum of pinocembrin	186
Figure 4. 45: EIMS spectrum of pinocembrin Figure 4. 45: EIMS fragmentations of pinocembrin	187
Figure 4. 45: EIMS fragmentations of pinocembrin	107
Figure 4. 46: Structure of naringenin	100
Figure 4. 46: Structure of naringenin (270 MHz, CD ₃ COCD ₃	101
Figure 4. 48: H-1H COSY spectrum of naringenin	102
Figure 4. 49: ¹³ C NMR spectrum of naringenin (67.8 MHz, CD ₃ COCD ₃)	102
Figure 4. 50: ¹ H- ¹³ C HETCOR spectrum of naringenin	104
Figure 4. 51: DEPT spectra of naringenin	194
Figure 4. 52: EIMS spectrum of naringenin	106
Fig. 4 52, CC MC anastrum of paringonin	. 196
Figure 4. 54: Structure of (-)-iso-5-deoxygoniopypyrone Figure 4. 55: Structure of (-)-iso-5-deoxygoniopypyrone	. 200

Figure 4. 56: Structure of (+)-5-deoxygoniopypyrone	200
Figure 4 58. H NMR spectrum of (-)-iso-5-deoxygoniopypyrone (crude) (2/0 MHz, CDCl ₃)	202
Figure 4. 59: H-1H COSY spectrum of (-)-iso-5-deoxygoniopypyrone (crude)	203
Figure 4. 60: ¹³ C NMR spectrum of (-)-iso-5-deoxygoniopypyrone (crude) (67.8 MHz, CDCl ₃)	204
Figure 4. 61: H-13C HETCOR spectrum of (-)-iso-5-deoxygoniopypyrone (crude)	205
Figure 4. 62: EIMS spectrum of (-)-iso-5-deoxygoniopypyrone accetate (270 MHz, CDCl ₃)	208
Figure 4. 63: 'H NMR spectrum of (-)-iso-5-deoxygoniopypyrone acetate (276 NH2, CDC43)	209
Figure 4. 64: H- H COSY spectrum of (-)-iso-5-deoxygoniopypyrone acetate (67.8 MHz, CDCl ₃)	210
Figure 4. 66: ¹ H- ¹³ C HETCOR spectrum of (-)-iso-5-deoxygoniopypyrone acetate	211
Figure 4. 67: DEPT spectra of (-)-iso-5-deoxygoniopypyrone acetate	212
Figure 4 69: Symthesis of (-) iso-5-degyygonionypyrone	213
Fig. 4 60. Destulated synthesis of gonjonynyrone	214
Pi 4 71. ILLNIMB encettum of (+) goniothalenol (400 MHz CDCla)	21/
E: 4 72. II II COSV spectrum of (+) goniothalenol (400 MHz CDCls)	210
Figure 4 73: Long-range H-H COSV spectrum of (+)-goniothalenol	219
Figure 4. 74: ¹³ C NMP enectrum of (+)-goniothalenol (400 MHz, CDCl ₂)	. 220
Figure 4.75, USOC spectrum of (+) conjutbalenol	. 221
Figure 4. 76: FIMS enectrum of (+)-goniothalenol	. 222
Figure 4, 77: Cyclication of 58-hydroxyisogoniothalamin epoxide	. 224
Figure 4. 79. Custination of 58 hydroxygoniothalamin enoxide	. 224
Figure 4 79: Structure of (+)-58-hydroxygoniothalamin	. 225
Figure 4. 80: ¹ H NMP spectrum of (+)-58-hydroxygoniothalamin (270 MHz, CDCl ₃)	. 228
Figure 4. 81: ¹³ C NMR spectrum of (+)-5B-hydroxygoniothalamin (67.8 MHz, CDCl ₃)	. 229
Figure 4 82: FIMS spectrum of (+)-5β-hydroxygoniothalamin	. 230
Figure 4, 83: Structure of (-)-5B-acetoxygoniothalamin	. 231
Figure 4. 84: ¹ H NMR spectrum of (-)-5β-acetoxygoniothalamin (270 MHz, CDCl ₃)	. 234
Figure 4. 85: 13C NIMP enectrum of (-)-5R-acetoxygoniothalamin (67.8 MHz. CDCl ₂)	. 235
Figure 4. 86: Attempted syntheses of 5-hydroxygoniothalamin (Smith, 1994)	. 237
Figure 4. 87: Structure of (+)-5α-hydroxygoniothalamin	. 238
Figure 4. 88: NOE enhancement for 5α-hydroxygoniothalamin	. 240
Figure 4. 88: NOE enhancement for 3α-hydroxygoniothalamin (270 MHz, CDCl ₃)	241
Figure 4. 89: ¹ H- ¹ H COSY spectrum of (+)-5α-hydroxygoniothalamin (270 MHz).	242
Figure 4. 90: 'H-'H COSY spectrum of (+)-5α-hydroxygoniothalamin (67.8 MHz, CDCl ₃)	243
Figure 4. 91: "C NMR spectrum of (+)-5α-hydroxygoniotnaiamin (67.8 MHz, CDCi3)	244
Figure 4. 92: ¹ H- ¹³ C HETCOR spectrum of (+)-5α-hydroxygoniothalamin	245
Figure 4. 93: NOE difference spectra of (+)-5α-hydroxygoniothalamin	243
Figure 4. 94: A perspective view of (+)-5α-hydroxygoniothalamin	. 247
Figure 4. 95: Mechanism for the oxidation of goniothalamin to (+)-5β-hydroxygoniothalamin	240
Figure 4. 96: Structure of(-)-5\(\alpha\)-acetoxygoniothalamin	. 249
Figure 4. 97: ¹ H NMR spectrum of (-)-5\(\alpha\)-acetoxygoniothalamin (270 MHz, CDCl ₃)	. 251
Figure 4. 98: ¹³ C NMR spectrum of (-)-5\(\alpha\)-acetoxygoniothalamin (67.8 MHz, CDCl ₃)	. 252
Figure 4, 00: NOE difference spectrum of (-)-50-acetoxygoniothalamin	253
Figure 4 100: Structure of (+) appropria	255
T' 4 101 FING 1 CIN (C f (1) announcin	25/
Figure 4, 102: FIMS for the silated derivative of (+)-annonacin	236
E' 4 102. Ctture of the or B unceturated lectons	239
Figure 4, 104: ¹ H NMR spectrum of (+)-annonacin (270 MHz, CDCl ₂)	201
Figure 4 105: 'H-'H COSY spectrum of (+)-annonacin	202
Figure 4. 106: ¹³ C NMR spectrum of (+)-annonacin (67.8 MHz, CDCl ₃)	264
Figure 4. 107: DEPT spectra of (+)-annonacin.	265
Figure 4. 107: DEP1 spectra of (+)-annonacin Figure 4. 108: ¹ H- ¹³ C HETCOR spectrum of (+)-annonacin	266
Figure 4. 109: CIMS spectrum of (+)-annonacin	267
Figure 4. 110: EIMS spectrum of (+)-annonacin	201

Figure 4. 111: EIMS spectrum of the silated derivative of (+)-annonacin	268
Figure 4. 112: Structure of disepalin	270
Figure 4. 113: FAB MS of disepalin	273
Figure 4 114: FAB MS and EIMS for the silated derivative of disepalin	213
Figure 4, 115: NOE enhancements observed for disensin	2/3
Figure 4. 116: ¹ H NMR spectrum of disepalin (500 MHz, CDCl ₃)	278
Figure 4, 117: H NMR spectrum of disepalin (500 MHz, CDCl ₃) (expanded)	219
Figure 4. 118: ¹³ C NMR spectrum of disepalin (67.8 MHz, CDCl ₃)	280
Figure 4. 119: ¹ H- ¹ H COSY spectrum of disepalin (500 MHz, CDCl ₃)	281
Figure 4. 120: NOE difference spectrum of disepalin	282
Figure 4 121: NOF difference spectra of disepalin (500 MHz, CDCl ₃)	283
Figure 4 122: HMBC spectrum of disepalin (500 MHz, CDCl ₃)	284
Figure 4 122: FAD MS enectrum of disensin	285
Figure 4 124: FAB MS spectrum of the silvlated derivative of disepalin	280
Pi 4 126. Companies of original patent BII	209
Figure 4 126: ¹ H NMR spectrum of aristolactam BII (270 MHz, CDCl ₃)	291
Figure 4 127: 'H-'H COSY spectrum of aristolactam BII	292
Figure 4. 128: ¹³ C NMR spectrum of aristolactam BII (67.8 MHz, CDCl ₃)	293
Figure 4, 120: FIMS spectrum of aristolactam BII	. 294
Figure 4 130: Structure of liriodenine	. 290
Figure 4 131: H NMR spectrum of liriodenine (270 MHz, CDCl ₃)	. 298
Figure 4 132: ¹³ C NMR spectrum of liriodenine (67.8 MHz, CDCl ₃)	. 299
Figure 4 133: FIMS spectrum of liriodenine	. 300
Figure 4 134: Structure of our egidione	. 302
Figure 4. 135: ¹ H NMR spectrum of ouregidione (270 MHz, CDCl ₃)	. 306
Figure 4 126: LU LU COSV enectrum of ouregidione	. 30/
Figure 4 137: 13C NMR spectrum of ouregidione (67.8 MHz, CDCl ₃)	. 308
Figure 4 138: NOE difference experiments for ouregidione	. 309
Figure 4. 139: EIMS spectrum of ouregidione	. 310

LIST OF PLATES

Plate 1: Fruits, flowers, leaves (top), and stem (bottom) of Goniothalamus andersonii
Plate 2: Flower (top), fruits (middle), and leaves (bottom) of Goniothalamus dolichocarpus
Plate 3: Fruits and flowers of Goniothalamus malayanus
Plate 4: Leaves of Goniothalamus velutinus (top), and leaves and shoot of Goniothalamus
macrophyllus (bottom)
Plate 5: Fruits, leaves and stem of Goniothalamus umbrosus (top), flowers and leaves of
Disepalum anomalum (bottom)
Ploto 6: Flowers on branches (top) and leaves (bottom) of Mezzetia umbellata

ACRONYMS/ABBREVIATIONS USED IN TEXT

'n alpha β heta

δ chemical shift in ppm

γ gamma micro gram degree in celsius adenosine triphosphate ATP

broad doublet brd brs broad singlet

N.O-bis (trimethylsilyl)-trifluoroacetamide BSTFA Chemical ionization mass spectroscopy CIMS

centimetre cm

COSY Correlated Spectroscopy

13C carbon-13 confidence limit CL

doublet d DRII 1.8-diazabicyclo [5.4.0] undec-7-ene

doublet of doublet dd

doublet of doublet of doublet ddd

doublet of doublet of doublet dddd

Distortionless enhancement by polarization transfer DEPT

dimethylsulfoxide DMSO DNP dinitrophenol dt doublet of triplet ED effective dose

Electron impact-mass spectroscopy EIMS

Fast Atom Bombardment FAR Fe-S ferrous sulphide

FΙ fluorescence inhibition

FP frond proliferation gram(s)

g GC Gas Chromatography

Gas Chromatography-mass spectroscopy GC-MS

growth inhibition GI

1H proton

HETCOR Heteronuclear chemical shift-correlation

Heteronuclear Multiple Bond Connectivity by 2D Multiple Quantum HMBC

NMR

¹H-Detected Heteronuclear Multiple-Quantum Coherence via Direct HMOC

Coupling

acetic acid HOAc HPLC high performance liquid chromatography

HRMS high resolution mass spectrum

Hz. hertz

internal diameter id

IR infrared coupling constant in Hz

.I

1

LC lethal concentration

lethal dose LD

literature Lit.

LPRI. log-probit regression lines

I.R long range coupling constant

m multiplet

m-chloroperoxybenzoic acid MCPBA

megahertz MHz

ml millilitre(s) millimetre(s) mm

mol moles

molecular weight Mol. wt.

melting point m.p.

mass spectrum/spectra/spectrometer/spectroscopy MS

methyl tert-butyl ether MTRE

MTPA methoxytrifluoromethylphenylacetate

NA not available

nicotinamide adenine dinucleotide, reduced form NADH

National Cancer Institute NCI

nanometre(s) nm

Nuclear Magnetic Resonance NMR

Nuclear Overhauser Enhancement NOE

Nuclear Overhauser and Exchange Spectroscopy NOESY

OAc acetate

PCN propionitrile Ph phenyl

preparative layer chromatography PLC

parts per million ppm

quartet a

dq doublet of quartet

[R] corresponding alkyl group

refractive index RΙ

S singlet standard error SE

t triplet

td triplet of doublets

tetrahvdrofuran THF

thin layer chromatography TLC TMS tetramethylsilane

N,N,N,N'-tetramethyl-p-phenylenediamine TMPD

ultra violet UV

WHO World Health Organization