

# DISCUSSION

## 0. Discussion

The ultimate objective of the present study was to evaluate the potential use of tropical microalgae as bioassay organisms for nitrogen and phosphorus enrichment in freshwater ecosystems. The approach used was to investigate the effects of minimal levels of nitrogen and phosphorus required to stimulate the growth of three chlorophytes within short growth duration (96 h). This approach differed from the previous study conducted by Foo *et al.* (2001) on the three species. In the previous study, the test organisms were subjected to high levels of  $\text{NaNO}_3$  and  $\text{NH}_4\text{Cl}$ , ranging from 2.9 to 250 mM, and the growth duration was 14 days. In comparison, the levels of nitrogen tested in the present study were in the range of 0.03 to 18.75 mM. The levels tested were close to that encountered in natural waters. For example, clean freshwater contains about 0.07 mM  $\text{NaNO}_3$  (Rainor, 1984) while agro-industrial wastewaters such as rubber effluent may contain 21 mM  $\text{NH}_3\text{-N}$  and 0.21 mM  $\text{NO}_3\text{-N}$  (Phang and Ong, 1988).

### 1. Selection of minimal culture medium for testing

Of the different strengths of BBM tested, 1% BBM was found to be the minimal medium that stimulated growth of *Chlorella vulgaris* UMACC 001. A comparison of nitrogen levels in various culture media and test media is shown in Table 66. The nitrogen level in the minimal medium was close to that found in natural freshwater. Thus, this medium was chosen as the minimal medium for further growth experiments. The 1% BBM was spiked with nitrogen and

phosphorus levels at an increasing factor of five-fold. The overall growth trends and cell numbers attained after 12 and 96 h, of the test organisms were then compared with that in Dilution Medium, which was without nitrogen and phosphorus.

Table 66: Nitrogen levels in various culture media and test media.

Medium	Nitrogen (mM)
Yamamoto <i>et al.</i>	3.54 mM
Yamamoto and Granick	7.5 mM
Yamamoto and Iwasa	3.0 mM
Yamamoto	2.9 mM
Yamamoto	0.5 mM
YAP (Provisional Algal Assay Procedure)	0.3 mM
Yamamoto	0.07 mM
1% BBM (Used in the present study)	0.03 mM

Reference: Trainor (1984).

## 2. Growth responses of the three chlorophytes to variations in nitrogen and phosphorus levels

In the previous study (Foo *et al.*, 2001), the growth responses, both stimulatory and inhibitory, of the chlorophytes to high nitrogen levels (up to 250 mM) were investigated. In comparison, the present study aimed to compare the

stimulatory growth response of the chlorophytes to low levels of nitrogen and phosphorus. It was found that *Chlorella vulgaris* UMACC 001 was the most tolerant to high nitrogen levels of the four chlorophytes tested by Foo *et al.* (2001). However, tests using low levels of nitrogen failed to detect distinct difference in growth response towards the range of nitrogen used. Summary of findings on the growth response of the three chlorophytes to different nitrogen and phosphorus levels is shown in Table 67.

There was no growth inhibition of the three chlorophytes when grown at  $\text{NH}_4\text{Cl}$  level as high as 18.75 mM. This finding contrasted with other studies which showed that high levels of ammonium, especially in the form of ammonia, are toxic to microalgae. For example, Kallqvist and Svenson (2003) reported that concentrations of  $\text{NH}_4^+$  and  $\text{NH}_3$  above 30  $\mu\text{M}$  inhibit growth of *Nephroselmis pyriformis*. Phytoplankton which are sensitive to high ammonia concentrations are usually those that inhabit environments with constant low levels of ammonia. The species used by Kallqvist and Svenson (2003) is a marine chlorophyte, and most probably isolated from marine waters with low ammonium level. And thus, this could probably account for the difference in sensitivity to ammonia. In contrast, the three chlorophytes tested in the present study were isolated from nitrogen-rich habitats. For example, *Chlorella vulgaris* UMACC 001 was isolated from an eutrophic lake at an experimental farm (Phang and Chu, 1996). This species was shown to be highly tolerant to high levels of nitrate and ammonium, growing well even at 250 mM  $\text{NaNO}_3$  or  $\text{NH}_4\text{Cl}$  (Foo *et al.*, 2001).

Expt	N & P levels	Growth response	Comments
1	NaNO <sub>3</sub> : 0.03, 0.3 & 3.0 mM	<ul style="list-style-type: none"> <li>0.03 mM (1% BBM) : minimal level of NaNO<sub>3</sub> that enhanced growth compared to Dilution Water, at least for the initial 4 days of growth</li> <li>Growth enhanced after 48 h for the three chlorophytes</li> <li>PGE-96 based on both cell numbers and OD<sub>620</sub> increased with increasing NaNO<sub>3</sub> levels for the three chlorophytes</li> </ul>	<ul style="list-style-type: none"> <li>1 % BBM was chosen as a 'minimal' medium for short-term (96 h) growth study</li> <li>Similar growth response of the three chlorophytes tested to the levels of NaNO<sub>3</sub> tested</li> </ul>
2	NaNO <sub>3</sub> : 0.03, 0.15, 0.75, 3.75 & 18.75 mM PO <sub>4</sub> <sup>3-</sup> :0.02 mM	<ul style="list-style-type: none"> <li><i>C. vulgaris</i> UMACC 001 &amp; <i>S. quadricauda</i> UMACC 041: Similar growth trends for NaNO<sub>3</sub> levels between 0 – 18.75 mM</li> <li><i>A. Convolutus</i> UMACC 101 : growth was enhanced at NaNO<sub>3</sub> ≥0.03 mM NaNO<sub>3</sub>; no marked difference in trends between 0.03 – 18.75 mM NaNO<sub>3</sub></li> <li>PGE-96 increased with increasing NaNO<sub>3</sub> levels for all species tested</li> </ul>	<ul style="list-style-type: none"> <li>PGE-96 was a better parameter for bioassays of NaNO<sub>3</sub> levels compared to growth trends</li> </ul>
3	NH <sub>4</sub> Cl : 0.03, 0.15, 0.75, 3.75 & 18.75 mM PO <sub>4</sub> <sup>3-</sup> :0.02 mM		
4	NaNO <sub>3</sub> : 0.03 mM PO <sub>4</sub> <sup>3-</sup> : 0.02, 0.1, 0.5, 2.5 & 12.5 mM	<ul style="list-style-type: none"> <li><i>C. vulgaris</i> UMACC 001: No marked difference in growth trends between 0 – 12.5 mM PO<sub>4</sub><sup>3-</sup></li> <li><i>S. quadricauda</i> UMACC 041: growth enhanced after 24 h, with respect to Dilution Water</li> <li><i>A. Convolutus</i> UMACC 101: cell numbers attained increased with increasing PO<sub>4</sub><sup>3-</sup> level</li> <li>Both PGE-12 and PGE-96 increased with increasing PO<sub>4</sub><sup>3-</sup> levels</li> </ul>	<ul style="list-style-type: none"> <li>The levels of PO<sub>4</sub><sup>3-</sup> tested did not enhance growth of <i>C. vulgaris</i> UMACC 001 when NaNO<sub>3</sub> level used was low (0.03 mM)</li> <li>Low NaNO<sub>3</sub> level sufficient for growth to use the excess PO<sub>4</sub><sup>3-</sup></li> </ul>

Table 67: A summary of growth responses of the three chlorophytes to nitrogen and phosphorus levels in the medium.

Expt	N & P levels	Growth response	Comments
5	NH <sub>4</sub> Cl : 0.03 mM PO <sub>4</sub> <sup>3-</sup> : 0.02, 0.1, 0.5, 2.5, & 12.5 mM	<ul style="list-style-type: none"> <li>• <i>C. vulgaris</i> UMACC 001: no growth enhancement between 0 – 12.5 mM PO<sub>4</sub><sup>3-</sup></li> <li>• <i>S. quadricauda</i> UMACC 041 &amp; <i>A. Convolvulus</i> UMACC 101: growth enhanced in PO<sub>4</sub><sup>3-</sup> containing medium after 48 h, no marked difference in growth trends between 0.02 – 12.5 mM</li> <li>• PGE-96 increased with increasing PO<sub>4</sub><sup>3-</sup> levels, PGE-12 not affected by PO<sub>4</sub><sup>3-</sup> levels</li> </ul>	<ul style="list-style-type: none"> <li>• PGE-96 was a good parameter for assessing growth enhancement due to increasing phosphate levels</li> </ul>
6	NaNO <sub>3</sub> : 18.75 Mm PO <sub>4</sub> <sup>3-</sup> : 0.02, 0.1, 0.5, 2.5, & 12.5 mM	<ul style="list-style-type: none"> <li>• Growth enhance after 24 h in PO<sub>4</sub><sup>3-</sup> containing medium for all species tested</li> <li>• PGE-96 increased with increasing PO<sub>4</sub><sup>3-</sup> levels</li> </ul>	<ul style="list-style-type: none"> <li>• PGE-96 was a good parameter for assessing growth enhancement due to increasing phosphate levels</li> </ul>
7	NH <sub>4</sub> Cl : 18.75 mM PO <sub>4</sub> <sup>3-</sup> : 0.02, 0.1, 0.5, 2.5, & 12.5 mM	<ul style="list-style-type: none"> <li>• Growth enhancement throughout the growth duration in PO<sub>4</sub><sup>3-</sup> containing medium compared to Dilution Water for all species tested</li> <li>• PGE-96 increased with increasing PO<sub>4</sub><sup>3-</sup> levels but not PGE-12</li> </ul>	<ul style="list-style-type: none"> <li>• PGE-96 was a good parameter for assessing growth enhancement due to increasing phosphate levels</li> </ul>

This species has also been successfully used to treat rubber effluent and palm oil mill effluent, as high percentages removal of nitrogen and phosphorus were achieved (Phang and Ong, 1988; Geetha *et al.*, 1994; Phang *et al.*, 2001). While species sensitive to high ammonium levels are suitable for toxicity testing, tolerant species apart from its use in treating wastewater, will also be useful as a bioassay organism to detect the high levels of ammonium in agro-industrial wastewater.

At low levels of  $\text{NaNO}_3$  (0.03 mM, 1%BBM), the growth of *Chlorella vulgaris* UMACC 001 was not enhanced even at a phosphate level ten-fold of that in BBM. However, at high  $\text{NaNO}_3$  level (18.75 mM), growth of this species was markedly enhanced with increasing phosphate levels. Thus, the cultures grown at 0.03 mM  $\text{NaNO}_3$  might be nitrogen-limited, and therefore, unable to use the excess phosphate efficiently.

Results showed that the cell number attained after 96 h was the most useful parameter for comparing the stimulatory effect of growth due to addition of low levels of nitrogen or phosphorus (Table 67). In comparison, cell number attained after 12 h may not show a consistent trend with respect to the levels of nitrogen or phosphorus in the medium. Thus, Percentage Growth Enhancement at 96 h (PGE-96) is a new parameter that we are introducing for use in bioassay of nitrogen and phosphorus. The overall growth trends and specific growth rates did not seem to be good parameters for bioassay. One good example from the present study was the response of *Ankistrodesmus convolutus* UMACC 101 to increasing phosphate levels. While  $\mu$  did not vary markedly (6.04 – 9.51  $\text{d}^{-1}$ ), the

GE-96 increased from 111 to 216% with increasing phosphate levels from 0.02 to 12.5 mM.

### 3. Potential use of the chlorophytes for bioassay

Algal bioassays are used to study the biological responses to the changes in water quality and to determine the levels of nutrients such as nitrogen and phosphorus, and pollutants such as heavy metals and pesticides in the water samples. One of the criteria in choosing an organism for assay purposes is that it should be sensitive to low concentrations of the material (Pipe *et al.*, 1984). One of the common species used in bioassay is *Selenastrum capricornutum* because it can be easily cultured, with minimum morphological changes during the population growth phase (Walsh *et al.*, 1984). This species is also highly sensitive and can be used to assay down to 0.01 mM nitrogen (Maestrini *et al.*, 1984). There have been efforts to search for new, more sensitive test species that respond to both growth inhibitors and stimulators. For instance, De Vries and Amphof (1984) explored the possible use of *Stigeoclonium* for bioassay of nitrogen in eutrophic waters. Recently, Kallqvist and Svenson (2003) used *Chlorella pyrenoidosa* in toxicity testing of ammonia in marine waters. Most of the species used in bioassays are temperate species, although there have been efforts to screen tropical phytoplankton for use in toxicity testing of heavy metals (Melor *et al.*, 2002).

This present study was probably the first to assess the potential use of tropical microalgae for bioassay of nitrogen and phosphorus enrichment. The



Three chlorophytes seem to be useful as bioassay organisms based on their response of PGE-96 to the increasing levels of nitrogen and phosphorus. However, if *Chlorella vulgaris* UMMAC 001 was to be used as bioassay for phosphorus, high nitrogen should be added in the test medium. Other features that make these three species useful bioassay include homogeneity of the cultures and minimum changes in morphology during the growth test.

In general, testing of natural waters is usually based on  $IC_{50}$  and Algal Growth Potential (AGP). The first parameter is more useful for toxicity testing rather than assessing growth stimulation due to nutrient enrichment (e.g. Hallqvist and Svenson, 2003). The AGP test is based on the principle that the maximum cell yield or biomass attained by the test alga is proportional to amount of nutrient present in the water sample (Skulberg, 1995). For example, the AGP assay based on *Selenastrum capricornutum* (NIVA-CHL-1) was used to detect the phosphorus level in agricultural runoff (Skulberg, 1995). The water samples were first sterilised by gamma-radiation before being inoculated with the test organism and the algal assay was performed for 9–12 days under standard growth conditions. Thus, longer time is required for AGP test compared to the bioassays of nitrogen and phosphorus enrichment demonstrated in the present study, which took only 96 h.

#### 4. Areas for further studies

The blooming of microalgae in natural environments is usually due to the variations of the ratio of nitrogen and phosphorus, and very unlikely to due solely

a single factor. For instance, Camacho and De Wit (2003) reported that adding nitrogen or phosphorus alone, or both nutrients at N: P ratio of 16 can cause blooming of different benthic microalgae in a hypersaline lake. Thus, one of the areas worth further studies is to investigate the interactive effects of both nitrogen and phosphorus, using factorial experimental design, on the growth of the three microphytes. Thus, it is worthwhile to investigate the growth response of the microalgae to variations in N: P ratios. The N: P ratios can be varied in batch cultures or by manipulating the growth rates of the test species in continuous culture.

The uptake of nitrogen and phosphorus is another area worthy for further studies. The uptake studies can be conducted using radiotracers such as  $N^{15}$  and  $P^{32}$ . Both bioassays and physiological tests should be conducted concomitantly (Hameed *et al.*, 1999). Apart from algal growth, the effects of nitrogen and phosphorus on physiological processes such as photosynthesis and accumulation of cell materials such as lipids, proteins, carbohydrates, nucleic acids and ATP should be further characterised.

Investigations on the activities of enzymes involved in nitrogen and phosphorus utilisation by microalgae is another avenue of further studies. For example, assaying of the alkaline phosphatase activity of symbiotic alga in coral can serve as a useful bioindicator for nitrogen status of the surrounding sea waters (Annis and Cook, 2002). Recently, Newman *et al.* (2002) reported that phosphatase activity is useful as an early warning indicator of wetland eutrophication. The potential use of molecular markers as bioindicator for

phosphorus enrichment is worth exploring. For example, a luciferase gene that is inducible by alkaline phosphatase has been cloned in *Synechococcus harveyi* (Schreiter *et al.*, 2001). Under phosphorus-limitation, the gene is induced and this causes bioluminescence.