AG 805

GAS CHROMATOGRAPHIC-MASS SPECTROMETRIC ANALYSIS OF CAPTOPRIL AND ITS APPLICATION IN PHARMACOKINETIC STUDIES IN HEALTHY SUBJECTS

ZAMRI BIN CHIK

Institute of Postgraduate Studies and Research (IPSR)

University of Malaya

Kuala Lumpur



Dissertation Submitted for the Degree of Master of Philosophy University of Malaya Kuala Lumpur

November 2000

CONFERENCE PRESENTATIONS

Zamri Chik, Toong C. Lee, Zahurin Mohamed, Mustafa A. Mohd, & Chim C. Lang (1999). Bioequivalence study of two tablet formulations of captopril (Brand A vs. Brand leader) in healthy volunteers. *Proceedings of the Annual Scientific Sessions, National Heart Association of Malaysia*, Kuala Lumpur, Malaysia. February, 2000.

Zamri Chik, Toong C. Lee, Zahurin Mohamed, Mustafa A. Mohd, & Chim C. Lang (1999). Bioequivalence study of two tablet formulations of captopril (Brand B vs. Brand leader) in healthy volunteers. Proceedings of the 15th Scientific Meeting of the Malaysian Society of Pharmacology and Physiology(MSPP), USM Kubang Kerian, Kelantan, Malaysia. May, 2000.

ACKNOWLEDGEMENTS

In the name of Allah, Most Gracious, Most Merciful. None of this work could have been done without your help.

First and foremost I would like to express my heartfelt gratitude to my supervisor, Assoc. Prof. Mustafa Ali Mohd. and to my co-supervisor Prof. Zahurin Mohamed for their beneficial guidance, invaluable advice and the commitment throughout this study. Rich flowing tributes are attributed to Assoc. Prof. Dr. Lee Toong Chow, Clinical Investigation Centre, University Malaya Medical Centre for his continuous guidance, suggestion, help and full commitment until the end of these thesis written, especially in bioequivalence studies and statistical analysis. Thanks also for his ideas and comments through out this thesis.

I am also thankful to the following individuals for their invaluable contributions to specific parts of this study especially bioequivalence studies: Dr. Astrit, Dr. Behzad, Dr. Anila Doci and Dr. Rokiah. My appreciation also goes to the following nurses and staff of UMMC: Sr. Noriah, Sr. Sofea, Sr. Rozana, Pakcik Mail, Mr. Voo Yam Por, and also to all my colleagues in bioequivalence studies: Ms. Pang, Ms. Hanan, Ms. Tan, Ms. Zuraini, Ms. Noraliza, Mr. Anuar and Ms. Azlina. I am also greatly indebted to all the volunteers who had participated in bioequivalence studies.

My thanks also goes to my colleagues in SUCXeS laboratory and Department of Pharmacology for their friendship and help. I am deeply thankful to my parents and parents in-law, for their ongoing support which has always made me strive to finish the sudy. Finally, I owe very much to my family, especially my wife, Asiah for her love, support, encouragement, patient and trust has given me and for blessing us with a daughter, Aida Shazwina and a son, Muhammad Haikal Haziq.

ABSTRACT

Captopril is a highly specific competitive inhibitor of the angiotensin-converting enzyme (ACE). This enzyme converts angiotensin I, a relatively inactive decapeptide, to angiotensin II, a potent endogenous vasoconstrictor substance. Captopril is widely used for the treatment of hypertension and congestive heart failure. This drug, which contains a sulphydryl group binds readily to albumin and to other plasma proteins. Captopril is unstable in blood and plasma ex vivo because of the reactivity of sulphydryl group which results in oxidation and rapid formation of disulphides. Therefore a fixative or a stabiliser must be added to each blood sample immediately following collection.

In this assay, N-ethylmaleimide (NEM) was used as a stabilizer in the collection tubes, prior to a rapid extraction technique by solid phase extraction (SPE). An enhanced and sensitive Gas Chromatography-Mass Spectrometry (GCMS) assay was developed for the quantitation of captopril. A commercially available internal standard, thiosalicylic acid (TSA) was used to minimise error in quantitation by GCMS. To enhance the volatility and gas chromatography elution properties, both captopril and TSA were derivatised to ester products by using pentafluorobenzylbromide. Captopril and TSA are quantitated as their bis-pentafluorobenzyl derivatives. The assay was linear from 1 to 160 ng/ml with mean recoveries of 104% and 99% for captopril and TSA, respectively, when the assay was carried out at 1.5, 75 and 150 ng/ml captopril. At the three concentrations of captopril mentioned, the coefficient of variation (CV) for inter–assay precision was 7.4, 9.5, and 4.4% respectively while accuracy was 6.1, 7.4, and 5.7%, respectively.

Four sets of bioequivalence studies were conducted for four generic captopril products with three products containing 25 mg captopril and one product containing 12.5 mg captopril. Bioequivalence studies were carried out in accordance with the Malaysian guideline on Good Clinical Practice (GCP) (adopted from ICH) which incorporates the Declaration of Helsinki as well as being accordance with standard operating procedures (SOP). All the generic products were found to be bioequivalent with the reference products based on statistical assessment of three parameters, namely $T_{\rm em}$ C_{em} and AUC.

The ratio test over reference (T/R) for T_{me}, C_{me}, and AUC were very close to 1 with the values ranging from 0.9 to 1.4, 0.9 to 1.1 and 1.0 to 1.1 for T_{me}, C_{me}, and AUC, respectively. The mean values for T_{me}, C_{me}, and AUC for 25 mg dose of brand leader and generic products was very similar. The values were 0.7, 117.1 and 212.6 for brand leader and 0.7, 120.5 and 212.6 for generic products. This is the same with 12.5 mg dose, where the mean value for T_{me}, C_{me}, and AUC were 0.8, 55.6 and 115.7 for Brand leader, while the generic products were 0.8, 48.9, and 106.4, respectively.

A standard two-stage analysis of pharmacokinetic parameters obtained from the oequivalence studies carried out, indicated that the clearance (CL), half-life (t_{10}) and elimination rate constant (K) in the 66 subjects studied, were within the range obtained from published data.

TABLE OF CONTENTS	
CONFERENCE PRESENTATIONS	i
ACKNOWLEDGEMENTS	ii
ABSTRACT	iv
TABLE OF CONTENTS	vi
LIST OF TABLES	xii
LIST OF FIGURES	xvii
ABBREVIATIONS	xxi
CHAPTER 1 INTRODUCTION	
1.1 ANGIOTENSIN-CONVERTING ENZYME (ACE) INHIBITORS	1
1.1.1 Mechanism of action	2
1.1.2 Chemical features of ACE inhibitors	5
1.2 CLINICAL USES OF ACE INHIBITORS	6
1.2.1 Hypertension	6
1.2.2 Congestive Heart Failure (CHF)	7
1.3 THE USE OF CAPTOPRIL FOR THE TREATMENT OF HYPERTENSION AND CONGESTIVE HEART FAILURE	9
1.3.1 General characteristics	9
1.3.2 Dosage and Administration	10
1.4 PHARMACOKINETICS OF CAPTOPRIL	1
1.4.1 Absorption	1

1.4.1.1 Effect of food

13

1.4.1.2 Effect of age	14
1.4.1.3 Gender effect	14
1.4.2 Distribution	14
1.4.3 Metabolism and excretion	16
1.4.4 Clearance	18
1.4.5 Elimination half life	19
1.5 PHARMACOKINETICS OF CAPTOPRIL IN PATIENTS WITH	
HYPERTENSION AND CONGESTIVE HEART FAILURE	20
1.5.1 Hypertension	20
1.5.2 Congestive heart failure	21
1.6 ADVERSE EFFECTS OF CAPTOPRIL	23
1.7 AIM OF THE PRESENT STUDY	24
1.8 POSSIBLE OUTCOMES OF THE STUDY	25
CHAPTER 2 ANALYTICAL METHODOLOGY FOR CAPTOPRIL	27
2.1 INTRODUCTION	27
2.1.1 The existence of a sulphydryl group in the captopril structure	27
2.1.2 Chemical structure for the detection of captopril by GCMS	28
2.1.3 Internal standard	28
2.2 PUBLISHED METHODS FOR CAPTOPRIL ANALYSIS	30
2.2.1 High performance liquid chromatography	30
2.2.2 Gas chromatography-mass spectrometry	32

2.3 METHOD DEVELOPMENT FOR CAPTOPRIL FOR THE PRESENT	
STUDY	33
2.3.1 Introduction	33
2.3.2 Extraction method of captopril	40
2.4 EXPERIMENTAL	43
2.4.1 Chemicals	43
2.4.2 Stock solutions	43
2.4.3 Standard / control solutions	43
2.4.4 Sample preparation	43
2.4.5 Gas Chromatograph – Mass Spectrometer	44
2.4.6 Method validation for the captopril assay	45
2.4.6.1 Specificity	45
2.4.6.2 Calibration curve	45
2.4.6.3 Precision, accuracy and recovery	46
2.4.6.4 Stability	47
2.5 RESULT AND DISCUSSION	48
2.6 CONCLUSION	58
CHAPTER 3 BIOEQUIVALENCE STUDY OF 4 BRANDS OF GENERIC CAPTOPRIL	59
3.1 INTRODUCTION	59
3.1.1 The concept of Bioequivalence	59
3.1.2 Bioavailability	61
3.1.3. Requirement for bioequivalence	61

3.1.4 Design of bioequivalence study	62
3.1.4.1 Parallel-Groups design	62
3.1.4.2 Two Period Crossover Design	63
3.1.4.3 Subjects and number of subject	63
3.1.5 Relevant pharmacokinetic parameters	64
3.1.6 Statistical analysis	66
3.2 STUDY OBJECTIVES	69
3.2.1 Primary Objective	69
3.2.2 Secondary Objective	69
3.3 STUDY TREATMENTS	69
3.3.1 Reference (R)	69
3.3.2 Test (T)	70
3.4 FACILITIES	71
3.5 STUDY METHODS	71
3.5.1 Study Design	71
3.5.2 Subject Number	72
3.5.3 Subject Enrolment	72
3.5.4 Restriction	74
3.5.5 Drug Accountability and Dosing	74
3.5.6 Admission and Procedure	75
3.5.7 Blood Sampling	77
3.5.8 Washout Period	79

3.6 ANALYTICAL METHOD	80
3.7 PHARMACOKINETICS AND STATISTICAL EVALUATION	81
3.7.1 Pharmacokinetics	81
3.7.2 Statistical	82
3.8 CLINICAL AND SAFETY RECORDS	82
3.9 DATA AND SAMPLES	83
3.10 GOOD CLINICAL PRACTICE AND ETHICAL CONSIDERATION	84
3.10.1 Ethics Approval	84
3.10.2 Informed Consent	84
3.10.3 Volunteer Compensation	85
3.10.4 Termination of the Study	85
3.10.5 Confidentiality	85
3.11 INTERIM ANALYSIS	85
3.12 RESULTS AND DISCUSSION	85
3.13 CONCLUSIONS	131
CHAPTER 4 POPULATION ANALYSIS OF PHARMACOKINETIC	
PARAMETERS OF CAPTOPRIL	132
4.1 INTRODUCTION	132
4.2 METHOD	133
4.3 RESULT	135
4.4 DISCUSSION AND CONCLUSION	144

	Table of contents		
CHAPTER 5 GENERAL CONCLUSION	145		
REFERENCES	150		
APPENDICES	163		

LIST OF TABLES

CH	A	PΊ	ж	ĸ	-1

1.1	Chemical-Physical properties of some ACE inhibitors in use	6
1.2	Captopril pharmacokinetic parameters in males and females after a 100 mg oral	
	dose (Massana E. et al. 1997)	14
1.3	Literature values of some body clearance for captopril	19
1.4	Literature values of some half-life for captopril	20
CHA	APTER 2	
2.1	Within-day precision for the assay of captopril in plasma using TSA as the	
	internal standard	54
2.2	Between-day precision for the assay of captopril in plasma using TSA as the	
	internal standard	54
2.3	Accuracy for assay of captopril in plasma using TSA as internal standard	55
2.4	Percentage recovery at 3 concentrations of captopril	56
2.5	Percentage Recovery of the Internal Standard	56
2.6	Long-term stability test of captopril at concentrations in the lower and higher part of	f
	the concentration range determined from mean of two readings	57
2.7	Published data of the values of accuracy, precision and LOD.	58
CH	APTER 3	
3.1	Crossover design with 10 subjects	63
3.2	Demographic and anthropometric data of the study participants (Study 1)	86

3.3	Demographic and anthropometric data of the study participants (Study 2)	87
3.4	Demographic and anthropometric data of the study participants. (Study 3)	88
3.5	Demographic and anthropometric data of the study participants. (Study 4)	89
3.6	Pertinent information for the calculation of $AUC(0\text{-}\infty)$ including the elimination	rate
	constant, K and the AUC fraction for reference product (brand leader) for Study 1	97
3.7	Pertinent information for the calculation of AUC(0- ∞) including the elimination rate	
	constant, K and the AUC fraction for reference product (brand leader) for Study 2	98
3.8	Pertinent information for the calculation of AUC(0- ∞) including the elimination rate	
	constant, K and the AUC fraction for reference product (brand leader) for Study 3	99
3.9	Pertinent information for the calculation of $AUC(0-\infty)$ including the elimination rate	
	constant, K and the AUC fraction for reference product (brand leader)for Study 4	100
3.10	Pertinent information for the calculation of $AUC(0-\infty)$ including the elimination rate	
	constant, K and the AUC fraction for test product (brand A) for $\mbox{\ study\ } l$	101
3.11	Pertinent information for the calculation of AUC(0- ∞) including the elimination rate	
	constant, K and the AUC fraction for test product (brand B) for Study 2	102
3.12	Pertinent information for the calculation of AUC(0-∞) including the elimination rate	
	constant, K and the AUC fraction for teat product (brand C) for Study 3	103
3.13	Pertinent information for the calculation of AUC(0-xx) including the elimination	rate
	constant, K and the AUC fraction for teat product (brand D) for Study 4	104
3.14	Individual values of T for both reference and test products for Study 1	105
3.15	Individual values of AUC (0-xx) for both reference and test products for Study 1	106
3.16	$_{\rm 5}$ Individual values of $C_{\rm ms}$ for both reference and test products for Study 1	106
3 17	7. Analysis of Variance for AUC (0-∞) for Study 1	107

3.18	Analysis of Variance for C _{nax} for Study 1	108
3.19	Individual values of $T_{\text{\tiny max}}$ for both reference and test product (Brand B) for Study	110
3.20	Individual values of AUC (0- ∞) for both reference and test product (Brand B) for	
	Study 2	111
3.21	Individual values of $C_{\mbox{\tiny max}}$ for both reference and test product (Brand B) for Study 2	112
3.22	Analysis of Variance for Log_{10} AUC $(0\text{-}\infty)$ for Study 2	113
3.23	Analysis of Variance for Log $_{10}$ C $_{\scriptsize \tiny max}$. for Study 2	114
3.24	Individual values of $T_{\text{\tiny max}}$ for both reference and test products (Brand C) for Study 3	115
3.25	Individual values of AUC (0- ∞) for both reference and test product (Brand C) for	
	Study 3	116
3.26	Individual values of $C_{\mbox{\tiny max}}$ for both reference and test product (Brand C) for Study 3	116
3.27	Analysis of Variance (Type III) for Log $_{10}$ AUC (0- \propto) for Study 3	117
3.28	Analysis of Variance (Type III) for Log $_{10}$ C $_{max}$ for Study 3	118
3.29	Individual values of $T_{\mbox{\tiny max}}$ for both reference and test products (Brand D) for Study 4	120
3.30	Individual values of AUC (0- ∞) for both reference and test product (Brand D) for	
	Study 4	121
3.31	Individual values of $C_{\mbox{\tiny max}}\mbox{for both reference}$ and test product (Brand D) for Study 4	122
3.32	Analysis of Variance (Type III) for Log_{10} AUC (0- $\!\propto$) for Study 4	123
3.33	Analysis of Variance (Type III) for $Log_{10}\ C_{\mbox{\tiny max}}$ for Study 4	124
3.34	Mean values of $T_{\mbox{\tiny max}},$ AUC, $C_{\mbox{\tiny max}},$ $t_{\mbox{\tiny 1/2}}$ and K for the Reference products (Brand le	ader
	used in Study 1, 2, 3 and 4	128
3.35	Mean values of $T_{\mbox{\tiny max}},$ AUC, $C_{\mbox{\tiny max}},$ $t_{\mbox{\tiny 1/2}}$ and K for the Test products (Brand A,B,C, and	D)
	used in Study 1, 2, 3 and 4	129

130

CHA	PT	ER 4	
4.1	De	scriptive statistics for CL / F, t_{12} and K	136
4.2	Ca	ptopril pharmacokinetic parameters for all subjects (n = 66) in the 4 sets of	
	bic	sequivalence studies carried out	139
4.3	Pu	blished data of the values of clearance (CL), half-life $(t_{\imath\alpha})$ and elimination rate	
	coı	nstant (K)	140
APP	EN	DIX A2.1	
A2.1	.1	Individual plasma captopril concentrations (ng/ml) following a single oral dose	of 25
		mg brand leader (Reference). (Study 1)	181
A2.1	.2	Individual plasma captopril concentrations (ng/ml) following a single oral dose	of 25
		mg brand A(Test).(Study 1)	182
A2.1	.3	Individual plasma captopril concentrations (ng/ml) following a single oral dose	of 25
		mg brand leader(Reference). (Study 2)	183
A2.1	.4	Individual plasma captopril concentrations (ng/ml) following a single oral dose	of 25
		mg brand B (Test). (Study 2)	184
A2.1	.5	Individual plasma captopril concentrations (ng/ml) following a single oral do	se of
		12.5 mg brand leader (Reference). (Study 3)	185
A2.1	.6	Individual plasma captopril concentrations (ng/ml) following a single oral do	se of
		12.5 mg brand C (Test). (Study 3)	186

3.36

Published data for T_{max}, AUC and C_{max}

A2.1.7	Individual plasma captopril concentrations (ng/ml) following a single ora	l dose o	1
	25 mg brand leader (Reference). (Study 4)	187	

A2.1.8 Individual plasma captopril concentrations (ng/ml) following a single oral dose of 25 mg brand D (Test). (Study 4) 188

LIST OF FIGURES

1.1 Sites of action of ACE inhibitor

CHAPTER 1

1.2	Chemical structure of captopril	9			
CHA	CHAPTER 2				
2.1.	Chemical structure for captopril and related substances	29			
2.2	Extraction of plasma using solid phase extraction	36			
2.3	Derivatisation reaction of captopril	36			
2.4	Derivatisation reaction of TSA	37			
2.5	Mass spectra of (A) bis-pentafluorobenzyl captopril and (B) bis-				
	pentafluorobenzyl TSA	38			
2.6	Total ion chromatogram for (A) captopril and (B) thiosalysilic acid	39			
2.7	Schematic diagram for extraction procedure of captopril	41			
2.8	Schematic diagram for SPE	42			
2.9	Chemical structure of fragment ions for bis-pentafluorobenzyl captopril	49			
2.10	Chemical structure of fragment ion for bis-pentafluorobenzyl TSA	50			
2.11	Chemical structure of free pentafluorobenzyl group	50			
2.12 GCMS chromatograms of plasma extracts: (A) blank plasma, (B) plasma spiked with 20					
	ng/ml IS (TSA) and 1 ng/ml captopril (LOQ)	51			
2.13	Standard curve for captopril from 1 to 160 ng/ml in plasma	52			

CHAPTER 3

3.1	Measurement of blood pressure and heart rate	76
3.2	Intravenous in-dwelling cannula were kept in a large anticubital vein	76
3.3	Dosing of subjects with the Test or Reference drug	76
3.4	Standardised food was served during the study	77
3.5	Sampling of blood from a subject	77
3.6	Blood samples were placed in centrifuge to separate out the plasma	79
3.7	Plasma was transfered to cryo vials	79
3.8	Sample preparation at the SUCXeS laboratory	80
3.9	Analysis of captopril by GCMS at the SUCXeS laboratory	81
3.10	Plot of mean plasma captopril concentration vs. time following a single dose of	f the
	reference drug and of Brand A, both containing 25 mg captopril. Values are expre	essed
	as mean \pm standard deviation	90
3.11	Plot of mean plasma captopril concentration vs. time following a single dose of	f the
	reference drug and of Brand B, both containing 25 mg captopril. Values are express	ed as
	mean ± standard deviation	91
3.12	Plot of mean plasma captopril concentration vs time following a single dose o	f the
	reference drug and of Brand C, both containing 12.5 mg captopril. Values are expre	essed
	as mean \pm standard deviation	91
3.13	Plot of mean plasma captopril concentration vs time following a single dose o	f the
	reference drug and of Brand D, both containing 25 mg captopril. Values are expre	essed
	as mean \pm standard deviation	92
3.14	Plot of mean plasma captopril concentration vs. time for all Reference products	93

94

95

CHAPTER 4					
4.1	Superimposed Histogram plot with fitted normal curve of frequency of observe	ation			
	versus clearance	141			
4.2	Superimposed Histogram plot with fitted normal curve of frequency of observe	ation			
	versus half-life	142			
4.3	Superimposed Histogram plot with fitted normal curve of frequency of observ	ation			
	versus elimination constant	143			
APF	PENDIX A1				
A1.2.1 Representative chromatogram of 1 ng/ml captopril standard. Peak identity: 2 = TSA,					
	16 = captopril	177			
A1.2	2.2 Representative chromatogram of 160 ng/ml captopril standard. Peak identity: 1=	TSA,			
	30 = captopril	177			
A1.2	2.3 Representative chromatogram of 1.5 ng/ml captopril control. Peak identity: 3=	TSA,			
	22 = captopril	178			
A1.2	2.4 Representative chromatogram of 150 ng/ml captopril control. Peak identity: 2=	TSA,			
	22 = captopril	178			
A1.2	2.5 Representative chromatogram of captopril sample from bioequivalence study	(40			
	minutes sampling). Peak identity: 3= TSA, 26 = captopril	179			

3.15 Plot of mean plasma captopril concentration vs. time for all Test products

3.16 Plot of mean plasma captopril concentration for all Reference and Test product

A1.2.6	Representative chromatogram of captopril sample from bioequivalence	study (10
	hours sampling). Peak identity: 3= TSA, 22 = captopril	179
APPE	NDIX A2	
A2.2.1	Pair wise presentation of individual plasma captopril concentration versus tin	me curves
	following a single dose of 25 mg captopril. (• = reference, brand leader,	\Box = test.
	Brand A). Study 1	189
A2.2.2	Pair wise presentation of individual plasma captopril concentration versus tir	me curves
	following a single dose of 25 mg captopril. (\bullet = reference, brand leader,	\Box = test,
	brand B). study 2	193
A2.2.3	Pair wise presentation of individual plasma captopril concentration versus tin	me curves
	following a single dose of 12.5 mg captopril. (• = reference, Brand leader,	\Box = test.
	Brand C). Study 3	201
A2.2.4	Pair wise presentation of individual plasma captopril concentration versus tin	me curves

following a single dose of 25 mg captopril. (• = reference, Brand leader, □ = test,

Brand D) . Study 4

206

ABBREVIATIONS

ACE angiotensin converting enzyme

RAAS renin – angiotensin-aldosterone system

FDA food and drug administration

SH sulphydryl

CHF congestive heart failure

CONSENSUS cooperative north Scandinavian enalapril survival study

SOLVD studies of left ventricular dysfunction

MI myocardial infarction

GCMS gas chromatography-mass spectrometry

HPLC high performance liquid chromatography

LOQ limit of quantitation

SD standard deviation

CV coefficient of variation

SEM standard error of the mean

MRT mean residence time

C_{max} maximum plasma concentration

T_{max} time to achieve peak plasma concentration

AUC area under the plasma concentration-time curve

AUC_{pst} area under the plasma concentration-time curve from time zero to the

last measurable concentration

AUC_{0-∞} area under the plasma concentration curve from time zero to infinity

K elimination rate constant

t_{1/2}: elimination/terminal half-life

V_d Volume of distribution

CL clearance

BE bioequivalence

DCA drug control authority

T test

R reference

ANOVA analysis of variance

CI confidence interval

α significance level

CIC clinical investigation centre

SUCXeS Shimadzu UMMC centre for xenobiotic studies

GMP good manufacturing practice

ICH International Conference of Harmonisation

GCP good clinical practice

MESC University Malaya Medical Ethics Sub-Committee

SOP standard operating procedure

COA certificate of analysis

CRFs case record form