

ACL - 3761

PERPUSTAKAAN UNIVERSITI MALAYA

INVN 31/1/2001

DIGESTION OF CELLULOSIC MATERIALS FROM
LOCAL FORAGE (*PENNISETUM PURPUREUM*)
AND PALM OIL MILL EFFLUENT (POME) BY THE
RUMEN BACTERIAL POPULATION FROM GOAT

BY

MOHD FUAT BIN ABD RAZAK

JABATAN GENETIK DAN BIOLOGI SEL
INSTITUT SAINS BIOLOGI
FAKULTI SAINS
UNIVERSITI MALAYA

Perpustakaan Universiti Malaya



A510033439

DISSSERTATION PRESENTED FOR THE
DEGREE OF MASTER OF SCIENCE
UNIVERSITY MALAYA

KUALA LUMPUR

Dimikrofiksikan pada..... 08.03.2002
No. Mikrofis..... 15367
(1999) Jumlah Mikrofis..... 3

HAMSIAH BT. MOHAMAD ZAHARI
UNIT REPROGRAFI
PERPUSTAKAAN UTAMA
UNIVERSITI MALAYA
U.P.R.

TO

NUR HANISAH & NUR HAZIRAH

ACKNOWLEDGEMENTS

I would like to express my sincere and utmost gratitude and thanks to my supervisor, Associate Prof. Shaiful Azni Abdul Aziz, for all his guidance, supervision, criticisms and encouragement that have enabled the completion of this research.

I am also indebted to the Dean of Institute for Post-graduate Studies and Research, University of Malaya (IPSP), for allowing me to use the various facilities available in the laboratory. I would like to extend my thanks and appreciation to the Manager of Bukit Rajah Plantation, Sime Darby Malaysia, for his kindness in providing the POME throughout the course of my research and to the members of Genetic & Cellular Biology Department, especially Mr. Rizan Daud, for all their help.

Last but not least, I also wished to thank my beloved wife for her help, patience and encouragement throughout my research.

This research has been sponsored by IRPA, R&D Vote 4/07/04/50, "Rumen Microbiology" (Ministry of Science, Technology and Environment, Malaysia).

ABSTRACT

Rumen samples were obtained from goats fed on *Pennisetum purpureum* (Rumen A), *P. purpureum* supplemented with Vigna bean sprout waste (Rumen B) and *P. purpureum* supplemented with palm oil mill effluent [POME] (Rumen C). The average total viable bacteria per gram of rumen content were 1.96×10^8 , 1.24×10^9 and 1.10×10^8 for rumen A, B and C respectively. Bacterial number was lowest in the reticulum of all types of rumen, but was highest in the dorsal-anterior part of rumen A and rumen B and in the dorsal-center part of rumen C. The occurrence of cellulolytic bacteria was highest in rumen B and lowest in rumen C. Addition of bean waste caused an increase in percentage of cellulolytic bacteria whereas the usage of POME caused eight-fold decrease in cellulolytic bacteria population.

Total soluble sugar and total volatile fatty acids concentrations were highest in rumen B followed by rumen A and rumen C. However, the concentration of total soluble sugar decreased from reticulum to the ventral part of rumen C. In rumen A and B, although the soluble sugar concentration was lowest in the ventral part of the rumen, the highest sugar concentration was in the dorsal-anterior part of the rumen A and in the ventral part of the rumen B. However, the distribution of VFA in the different part of

rumen A and C was the reverse of the pattern for soluble sugar. In rumen B, VFA concentration decreased from reticulum to the dorsal-anterior part of rumen and then increased to the ventral part of rumen.

Between the three rumens samples, rumen A showed highest cellulolytic activity against grass fibre, POME fibre, microcrystalline cellulose and acid-swollen cellulose followed by rumen B, and rumen C. In rumen A and rumen C digestion of grass fibre and microcrystalline cellulose was more active in the ventral part of rumen whereas the digestion of acid-swollen cellulose and POME fibre was more active in the dorsal part of rumen. On the contrary, the digestion of grass fibre, microcrystalline cellulose and POME fibre was more active in the dorsal part of rumen whereas digestion of acid-swollen cellulose was more active in the ventral part of rumen B.

Eleven percent of the cellulolytic bacteria isolated from the rumen of goat fed with *P. purpureum* showed positive cellulolytic activity against acid-swollen cellulose and microcrystalline cellulose. *Ruminococcus flavefaciens* was the most dominant (34.62%) cellulolytic bacteria isolated followed by *Fibrobacter succinogenes* (30.77%), *Ruminococcus albus* (11.54%), *Butyrivibrio fibrisolvens* (11.52%), *Eubacterium cellulosolvens* (7.69%) and *Clostridium cellobioparum* (3.85%). This was in agreement with general trend for rumen cellulolytic bacterial population. However, the presence of only 6.8% of

cellulolytic bacteria in rumen fed with POME as compared to grass could be attributed to the different in the nutrient status of the two substrates. Although the concentration of nitrogen-free extract was approximately equal in both rumens, *P. purpureum* was characterised by higher fibre content (29.40%) compared to POME (16.80%) but POME contained higher fat, protein and soluble sugar.

The suitability of POME as medium for growth and cellulolytic activity of pure cultures of ruminal cellulolytic bacteria was studied using whole POME, POME extracts, and POME extracts added with acid-swollen and microcrystalline cellulose. *R. albus*, *F. succinogenes*, *R. flavefaciens*, *C. celliciboparum* and *B. fibrisolvans* grew in all the POME or POME-based media. However, *B. fibrisolvans* and *C. celliciboparum* showed better growth in all media compared to the other cellulolytic bacteria but the former had much lower cellulolytic activity than *R. albus* and *R. flavefaciens*. *R. flavefaciens* produced extracellular cellulase in POME extract in the absence of cellulose whereas *R. albus* require the presence of cellulose for higher production of extracellular cellulase. *F. succinogenes* showed poor growth and low extracellular cellulase production in POME or POME-based media.

Fermentation of POME and *P. purpureum* by rumen bacterial population from grass-fed goats was studied in a semi-batch fermentation system simulating the rumen system. Soluble sugar in both substrates was rapidly utilised

followed by rapid increase in total rumen bacteria, total cellulolytic bacteria and volatile fatty acid concentration. However, molar ratio of fatty acids and fatty acid utilisation pattern were different in both systems. The acetic acid to propionic acid ratio indicated the presence of active cellulolytic bacterial population in both systems. However, the population adapted to grow on *P. purpureum* were better in cellulolytic activity against grass fibre, POME fibre and microcrystalline cellulose, whereas the population adapted to grow on POME were better in cellulolytic activity against acid-swollen cellulose. This study suggested that there was a change of bacterial population grown on POME.

ABSTRAK

Sampel-sampel rumen telah diperolehi dari kambing yang makan *Pennisetum purpureum* (Rumen A), *P. purpureum* yang ditambah dengan sisa kacang Vigna bertunas (Rumen B), dan *P. purpureum* yang ditambah dengan efluen kilang kelapa sawit [POME] (Rumen C). Purata jumlah bakteria hidup setiap gram kandungan rumen ialah 1.98×10^5 bagi rumen A, 1.24×10^5 bagi rumen B, dan 1.10×10^5 bagi rumen C. Retikulum kesemua jenis rumen mengandungi bilangan bakteria terendah, tetapi tertinggi di bahagian anterior dorsal rumen A dan rumen B dan di bahagian tengah dorsal rumen C. Rumen B mengandungi bilangan bakteria selulolitik tertinggi dan rumen C mengandungi bilangan bakteria selulolitik terendah. Sisa kacang yang ditambah dalam makanan kambing mengakibatkan peningkatan peratusan bilangan bakteria selulolitik manakala penambahan POME menyebabkan penurunan bilangan bakteria selulolitik sebanyak 6 kali ganda.

Jumlah gula larut dan kepekatan asid lemak meruap adalah tertinggi dalam kandungan rumen B, diikuti oleh rumen A dan rumen C. Kepekatan gula larut menurun dari retikulum ke bahagian ventral rumen C. Walaupun kepekatan gula juga terendah di bahagian ventral rumen A dan rumen B tetapi kepekatan gula paling tinggi di bahagian anterior dorsal rumen A dan di bahagian ventral rumen B. Walau bagaimanapun, corak taburan VFA dalam berbagai bahagian rumen A dan rumen

C adalah berlawanan dari corak taburan kepekatan gula larut di dalam rumen-rumen tersebut. Dalam rumen B, kepekatan VFA berkurangan dari retikulum ke bahagian anterior dorsal dan kemudiannya meningkat di bahagian ventral rumen.

Diantara ketigá-tiga rumen, rumen A menunjukkan aktiviti selulolitik tertinggi terhadap serabut rumput *P. purpureum*, serabut POME, selulosa mikrokristal dan selulosa kembang-asid. Aktiviti selulotik kandungan rumen B lebih tinggi dari rumen C. Dalam rumen A dan rumen C, penghazaman serabut rumput dan selulosa mikrokristal adalah lebih aktif di bahagian ventral rumen manakala penghazaman serabut POME dan selulosa kembang-asid lebih aktif di bahagian dorsal rumen. Sebaliknya, penghazaman serabut rumput, selulosa mikrokristal dan serabut POME adalah lebih aktif di bahagian dorsal rumen B manakala penghazaman selulosa kembang-asid di dalam rumen B paling aktif di bahagian ventral.

Hanya sebelas peratus dari bakteria yang diasingkan dari rumen kambing yang makan *P. purpureum* sahaja menunjukkan aktiviti selulolitik terhadap selulosa mikrokristal atau selulosa kembang-asid. *Ruminococcus flavefaciens* adalah bakteria selulolitik paling banyak diasingkan dari rumen ini (34.62%) diikuti oleh *Fibrobacter succinogenes* (30.77%), *Ruminococcus albus* (11.54%), *Butyrivibrio fibrisolvens* (11.52%), *Eubacterium cellulosolvens* (7.69%) dan *Clostridium cellobioparum*

(3.85%). Keputusan ini bersamaan dengan corak normal populasi bakteria selulolitik dalam rumen. Walau bagaimanapun, peratusan bakteria selulolitik di dalam rumen kambing yang makan POME lebih rendah iaitu sebanyak 6.8% dan ini adalah disebabkan perbezaan status nutrien diantara *P. purpureum* dengan POME. Walaupun kepekatan ektrak bebas-nitrogen hampir sama dalam kedua-dua *P. purpureum* dan POME, *P. purpureum* mengandungai selulosa lebih tinggi kepekatannya (29.40%) berbanding dengan POME (16.80%). POME pula lebih tinggi kandungan lemak, protein, dan gula larut berbanding dengan *P. purpureum*.

Kesesuaian POME sebagai medium pertumbuhan dan penghasilan aktiviti selulolitik beberapa kultur tulin bakteria selulolitik rumen telah dikaji menggunakan POME, ektrak POME, dan ektrak POME yang ditambah dengan selulosa mikrokristal dan selulosa kembang-asid. *R. albus*, *R. flavefaciens*, *C. cellobioparum* dan *B. fibrisolvens* boleh tumbuh dalam POME atau medium berdasarkan POME. Walaupun *B. fibrisolvens* dan *C. cellobioparum* menunjukkan pertumbuhan lebih baik di dalam kesemua media pertumbuhan berbanding dengan bakteria selulolitik yang lain tetapi aktiviti selulolitik kedua-duanya adalah jauh lebih rendah dari *R. albus* dan *R. flavefaciens*. *R. flavefaciens* menghasilkan selulase luar-sel di dalam ektrak POME walaupun dalam ketiadaan selulosa manakala *R. albus* memerlukan kehadiran selulosa bagi menghasilkan selulase luar-sel yang lebih

banyak. *F. succinogenes* menunjukkan kadar pertumbuhan dan penghasilan selulase luar-sel paling rendah dalam kesemua media pertumbuhan.

Fermentasi POME dan *P. purpureum* oleh populasi bakteria rumen dari rumen kambing yang makan *P. purpureum* telah dikaji dalam sistem fermentasi separa-kelompok yang meniru sistem rumen asli. Didapati bahawa gula larut dalam kedua-dua substrat digunakan secara pantas dan diikuti dengan pertambahan pantas bilangan jumlah bakteria, bilangan bakteria selulolitik, dan kepekatan asid lemak meruwap. Walau bagaimanapun, nisbah molar asid-asid lemak dan corak penggunaan semula asid-asid lemak berbeza di dalam sistem fermentasi POME dengan sistem fermentasi *P. purpureum*. Nisbah asid asetik dengan asid propionik menunjukkan terdapatnya populasi bakteria selulolitik yang aktif di dalam kedua-dua sistem fermentasi. Walau bagaimanapun, populasi bakteria yang telah menyesuaikan diri hidup menggunakan substrat *P. purpureum* menunjukkan aktiviti selulolitik lebih tinggi terhadap serabut *P. purpureum*, selulosa mikrokristal dan serabut POME berbanding dengan populasi bakteria yang telah menyesuaikan diri hidup mengunakan POME. Walau bagaimanapun, populasi bakteria yang telah menyesuaikan diri hidup menggunakan POME menunjukkan aktiviti selulolitik yang baik terhadap selulosa kembang-asid berbanding dengan populasi bakteria yang telah menyesuaikan diri hidup menggunakan substrat *P. purpureum*. Kajian ini menunjukkan terdapatnya perubahan populasi bakteria setelah tumbuh menggunakan substrat POME.

CONTENTS

ACKNOWLEDGEMENT	iii
ABSTRACT	iv
ABSTRAK	viii
LIST OF FIGURES	xvii
LIST OF TABLES	xix

CHAPTER 1: INTRODUCTION

1.1.	The ruminants	1
1.2.	Microbial activity in the rumen	2
1.3.	Rumen microbial population	3
1.3.1.	Rumen bacteria	4
1.3.2.	Rumen fungi	8
1.3.3.	Rumen protozoa	10
1.4.	Influence of diet on the microorganisms in the rumen	13
1.5.	Degradation of feedstuff in the rumen	16
1.6.	Enzymatic hydrolysis of cellulose	17
1.6.1.	Basic mechanism	17
1.6.2.	Factors affecting the complexity of Cellulase system	18
1.6.3.	Cellulases of the rumen bacteria	20
1.6.4.	Factors affecting the cellulolytic activity of the cellulolytic bacterial population	24
1.7.	Present programme	28
1.8.	Objectives of research	30

CHAPTER 2: CHARACTERIZATION OF RUMEN INGESTA FROM GOATS FED
WITH *P. PURPUREUM*, *P. PURPUREUM* WITH VIGNA BEAN
WASTES AND *P. PURPUREUM* WITH POME-BASED
CONCENTRATE

2.0.	INTRODUCTION	31
2.1.	BACTERIAL COUNT	32
2.1.1.	Materials and methods	32
2.1.1a.	Rumen samples	32
2.1.1b.	Preparation of oxygen-free CO ₂	32
2.1.1c.	Preparation of pre-reduced dilution solution	33
2.1.1d.	Media for roll-tube	34
2.1.1e.	Rumen content sampling and inoculum preparation	36
2.1.1f.	Total viable and total cellulolytic bacterial count	37
2.1.2	Results	37
2.2.	ANALYSES ON RUMEN INGESTA	40
2.2.1.	Total soluble sugar	40
2.2.1a.	Materials and methods	40
2.2.1b.	Results	41
2.2.2.	Volatile fatty acids contents	41
2.2.2a.	Materials and methods	41
2.2.2b.	Results	42
2.2.3.	Cellulolytic activity	46
2.2.3a.	Materials and methods	46
2.2.3a.1.	Cellulosic substrates	46
2.2.3a.2.	Measurement of cellulolytic activity	47

2.2.3b.	Results	49
2.2.4.	pH	54
2.2.4a.	Materials and methods	54
2.2.4b.	Results	54
2.3.	DISCUSSION	56

**CHAPTER 3: ISOLATION AND IDENTIFICATION OF CELLULOLYTIC
RUMEN BACTERIA**

3.0.	INTRODUCTION	63
3.1.	ISOLATION OF CELLULOLYTIC BACTERIA	64
3.1.1.	Materials and methods	64
3.2.	DETERMINATION OF CELLULOLYTIC ACTIVITY	66
3.2.1.	Materials and methods	66
3.2.2.	Results	66
3.3.	IDENTIFICATION OF CELLULOLYTIC BACTERIA	68
3.3.1.	Cellular morphology, Gram reaction, spore, motility and catalase production	68
3.3.1.1.	Materials and methods	68
3.3.1.2.	Results	69
3.3.2.	VFA and alcohol production	71
3.3.2.1.	Materials and methods	71
3.3.2.2.	Results	71
3.3.3.	Carbohydrate fermentation	73
3.3.3.1.	Materials and methods	73
3.3.3.2.	Results	74
3.3.4.	Characterization of isolates	74
3.4.	DISCUSSION	78

CHAPTER 4: GROWTH AND CELLULOLYTIC ACTIVITY OF PURE
CULTURES OF *R. ALBUS*, *R. FLAVEFACIENS*,
F. SUCCINOGENES, *B. FIBRISOLVENS*, AND
C. CELLOBIOPARUM IN POME, POME EXTRACTS
AND POME EXTRACTS PLUS CELLULOSE

4.0.	INTRODUCTION	82
4.1.	NUTRIENT ANALYSIS OF POME	82
4.1.1.	Materials and methods	82
4.1.1a	Ash	83
4.1.1b	Fat	83
4.1.1c	Fibre	84
4.1.1d	Protein	85
4.1.1e	Nitrogen-free extracts	86
4.1.1f	Soluble sugar	86
4.1.2.	Results	87
4.2.	GROWTH AND CELLULOLYTIC ACTIVITY	88
4.2.1	Materials and methods	88
4.2.1.1	Media preparation	88
4.2.1.2	Determination of Soluble sugar in POME media	90
4.2.1.3	Preparation of inoculum	90
4.2.1.4	Inoculation and incubation procedure	91
4.2.1.5	Analyses of samples	91
4.2.2	Results	92
4.2.2.1	Soluble sugar in POME media	92
4.2.2.2	Bacterial count	93
4.2.2.3	Cellulolytic activity	97
4.3.	DISCUSSION	98

CHAPTER 5: DIGESTION OF *P. PURPUREUM* AND POME IN
SEMI-BATCH FERMENTATION SYSTEM

5.0	INTRODUCTION	107
5.1	NUTRIENT ANALYSIS OF <i>PENNISETUM PURPUREUM</i>	108
5.1.1	Materials and methods	108
5.1.2	Results	108
5.2.	SEMI-BATCH FERMENTATION	109
5.2.1.	Materials and methods	109
5.2.1.1	Culture apparatus	109
5.2.1.2	Media preparation	113
5.2.1.3	Preparation of inoculum	115
5.2.1.4	Inoculation and fermentation procedure	115
5.2.1.5	Analyses of samples	116
5.2.2	RESULTS	117
5.2.2.1	Total viable and cellulolytic counts	117
5.2.2.2	Total soluble sugar	118
5.2.2.3	VFA concentration and pattern	122
5.2.2.4	Cellulolytic activity	123
5.3.	DISCUSSION	133
CHAPTER 6:	GENERAL DISCUSSION AND CONCLUSION	143
REFERENCES		155
COMMUNICATION		199

LIST OF FIGURES

Figure 2.1: Total viable bacteria	39
Figure 2.2: Total soluble sugar	43
Figure 2.3: Total volatile fatty acids	44
Figure 2.4a: Ruminal crude cellulase extracts on swollen cellulose	50
Figure 2.4b: Ruminal crude cellulase extracts on crystalline cellulose	51
Figure 2.4c: Crude cellulase extracts on POME fibres	52
Figure 2.4d: Crude cellulase extracts on grass fibres	53
Figure 4.1a: Growth of bacteria in POME extracts	94
Figure 4.1b: Growth of bacteria in POME extracts plus celluloses	95
Figure 4.1c: Growth of bacteria in POME	96
Figure 4.2a: Cellulolytic activity in POME extracts	99
Figure 4.2b: Cellulolytic activity in POME extracts plus cellulose	100

Figure 4.2c: Cellulolytic activity in POME	101
Figure 5.1 Set-up of semi-batch fermentation	112
Figure 5.2: Total viable bacterial count in semi-batch fermentation	119
Figure 5.3: Total viable celluolytic count in semi-batch fermentation	120
Figure 5.4: Total soluble sugar in semi-batch fermentation	121
Figure 5.5: Total volatile fatty acids in semi-batch fermentation	124
Figure 5.6: Acetic to propionic acid ratio	125
Figure 5.7a: Cellulolytic activity for swollen cellulose	128
Figure 5.7b: Cellulolytic activity for POME fibre	129
Figure 5.7c: Cellulolytic activity for crystalline cellulose	130
Figure 5.7d: Cellulolytic activity for grass fibre	131

LIST OF TABLES

Table 2.1: Count of total and cellulolytic bacteria and percentage of cellulolytic bacteria	38
Table 2.2: molar % of major VFA at the different part of rumen	45
Table 2.3: pH at the different part of rumen	55
Table 3.1: Cellulolytic activity of cellulolytic bacterial isolates	67
Table 3.2: Cell shape, Gram reaction, endospores, motility and catalase reaction	70
Table 3.3: Fermentation end products	72
Table 3.4: Fermentation of carbohydrates	75
Table 3.5: Characterization of isolates	77
Table 3.6: Occurrence of cellulolytic bacteria in the rumen of sheep with different diets	81
Table 4.1: Chemical constituents of POME	87
Table 4.2: Total soluble sugar in POME media	92

Table 5.1: Chemical constituents of <i>P. purpureum</i>	109
Table 5.2a: VFA in grass semi-batch system	126
Table 5.2b: VFA in POME semi-batch system	126
Table 5.3: Relative rate of digestion of different forms of cellulose by cellulase enzyme produced in semi-batch fermentation	132