# INDUCED MUTATION OF *IN VITRO* AQUATIC PLANT, *CRYPTOCORYNE XWILLISII* REITZ BY USING GAMMA IRRDIATION AND IRAP ANALYSES TO DISTINGUISH THE SPORTS (CLONAL MUTATION)

NORHANIZAN BINTI SAHIDIN

INSTITUTE OF BIOLOGICAL SCIENCES FACULTY OF SCIENCE UNIVERSITY OF MALAYA KUALA LUMPUR

2012

# INDUCED MUTATION OF *IN VITRO* AQUATIC PLANT, *CRYPTOCORYNE XWILLISII* REITZ BY USING GAMMA IRRDIATION AND IRAP ANALYSES TO DISTINGUISH THE SPORTS (CLONAL MUTATION)

### NORHANIZAN BINTI SAHIDIN

# DISSERTATION SUBMITTED IN FULFILMENT OF THE REQUIREMENTS FOR THE DEGREE OF MASTER OF SCIENCE

# INSTITUTE OF BIOLOGICAL SCIENCES FACULTY OF SCIENCE UNIVERSITY OF MALAYA KUALA LUMPUR

2012

# INDUCED MUTATION OF *IN VITRO* AQUATIC PLANT, *CRYPTOCORYNE XWILLISII* REITZ BY USING GAMMA IRRADIATION AND IRAP ANALYSES TO DISTINGUISH THE SPORTS (CLONAL MUTATION)

### NORHANIZAN BT. SAHIDIN

# DISSERTATION SUBMITTED IN FULFILMENT OF THE REQUIREMENTS FOR THE DEGREE OF MASTER OF SCIENCE

# INSTITUTE OF BIOLOGICAL SCIENCES FACULTY OF SCIENCE UNIVERSITY OF MALAYA KUALA LUMPUR

2012

### **ORIGINAL LITERARY WORK DECLARATION**

Name of Candidate: NORHANIZAN SAHIDIN (I.C/Passport No:720520015886)

**UNIVERSITI MALAYA** 

Registration/Matric No: SGR060027

Name of Degree: MASTER OF SCIENCE

Title of Project Paper/Research Report/Dissertation/Thesis ("this Work"):

### INDUCED MUTATION OF IN VITRO AQUATIC PLANT, CRYPTOCORYNE XWILLISII REITZ BY USING GAMMA IRRADIATION AND IRAP ANALYSIS TO DISTINGUISH THE SPORTS (CLONAL MUTATION)

Field of Study: Plant Biotechnology

I do solemnly and sincerely declare that:

- (1) I am the sole author/writer of this Work;
- (2) This Work is original;
- (3) Any use of any work in which copyright exists was done by way of fair dealing and for permitted purposes and any excerpt or extract from, or reference to or reproduction of any copyright work has been disclosed expressly and sufficiently and the title of the Work and its authorship have been acknowledged in this Work;
- (4) I do not have any actual knowledge nor do I ought reasonably to know that the making of this work constitutes an infringement of any copyright work;
- (5) I hereby assign all and every rights in the copyright to this Work to the University of Malaya ("UM"), who henceforth shall be owner of the copyright in this Work and that any reproduction or use in any form or by any means whatsoever is prohibited without the written consent of UM having been first had and obtained;
- (6) I am fully aware that if in the course of making this Work I have infringed any copyright whether intentionally or otherwise, I may be subject to legal action or any other action as may be determined by UM.

Candidate's Signature

Date

Subscribed and solemnly declared before,

Witness's Signature

Date

Name: Designation:

#### ABSTRACT

One part of ornamental fish industry is the aquatic plants. Ornamental fish industry is identifying as one of National Key Economic Area (NKEA) for Malaysia, under business opportunity. *Cryptocoryne xwillisii* Engler ex Baum is one of the highly demanded aquatic plants in international market. Unfortunately, the plants take months to grow, seldom flowering and no viable seeds. The studies were done to mass-produce the plants by tissue culture and to develop new variety by mutation.

Tissue culture of water trumpet, *Cryptocoryne xwillisii* Engler ex Baum, was developed using Murashige and Skoog 1962 (MS) medium, which contained two different plant growth regulators, namely 6-benzyladenine purine (BAP) and  $\alpha$ -naphthalene acetic acid (NAA). Seven different concentrations of BAP (0, 0.5, 1.0, 5.0, 10.0, 20.0 and 40.0  $\mu$ M) and NAA (0, 0.5, 1.0, 5.0, 10.0, 20.0 and 40.0  $\mu$ M) were investigated using a two-factor factorial design with 10 replicates. Results of the experiments were collected and analysed after 40 days of culture.

The results showed that the effects of plant growth regulators on increasing the number of shoots were highly significant (p<0.01). The MS medium containing 1.0  $\mu$ M BAP alone was the optimum concentration producing 6.8 ± 1.75 shoots per explant. This was the minimum formula concentration of BAP used that produced the highest number of shoots. Results showed that there were twelve hormone combinations giving high mean values of between 4 to 6 shoots per explant. All of the explants cultured in these media produced shoot (100 %).

Although all the explants gave a positive response in terms of regeneration, they however differed in the number and size of shoots produced. Analysis of the data using ANOVA indicated that the number of shoots produced was significantly influenced by both BAP and NAA concentrations simultaneously. This was suggested by the significance of the interactions between BAP and NAA which showed that BAP concentrations affected the number of shoots differently for each concentrations of NAA tested and vice versa.

Two new varieties of *C*. *xwillisii* have been developed through mutagenesis in this work. Shoot tips explants of *C*. *xwillisii* were subjected to a range of  $^{60}$ Co gamma ray irradiation of 0, 100, 200, 300, 400, 500, 600, 700 and 800 Gray. Results from experiments showed that the LD<sub>50</sub> for the tissue culture plants of *C*. *xwillisii* was at 32 Gy dose. And was therefore, less than 32 Gray was used as the appropriate dosage for induced mutations of the plants.

About two thousand of the shoot tips explants were irradiated using the 25 Gy and variants from the  $M_1$ ,  $M_2$ ,  $M_3$  and  $M_4$  generations were screened for morphological differences. The variants shoots were subcultured repeatedly until the 4<sup>th</sup> generation ( $M_4$ ) to ensure stability of mutants. Although initially many regenerants with different morphological traits were produced, only two mutants were shown to remain stable. The mutants obtained were dwarf plants (D1) and plants of taller stature with pigmented leaves (G1) than the controls. This was verified from the significant value of the F test in the ANOVA where P<0.05.

The Inter-Retrotransposon Amplified Polymorphism (IRAP) markers distinguished the D1 and G1 genomes from normal *C. xwillisii* genomes. The analysis revealed two specific bands 325 bp and 420 bp using Nikita primer for the D1 mutant and 240 bp and 300 bp using combination of 3'LTR primer and LTR 6149 primer for the G1 mutant.

#### ABSTRAK

Tumbuhan akuatik merupakan salah satu bahagian dalam industri ikan hiasan. Industri ikan hiasan telah dikenalpasti sebagai salah satu bidang keutamaan negara (National Key Economic Area (NKEA)), dibawah peluang perniagaan (business opportunity). *Cryptocoryne xwillisii* Engler ex Baum merupakan salah satu tumbuhan akuatik yang mendapat permintaan yang tinggi dalam pasaran antarabangsa. Akan tetapi, tumbuhan ini mengambil masa yang lama untuk membesar, jarang berbunga dan tiada biji benih yang viable. Kajian yang dijalankan adalah untuk menghasilkan tumbuhan ini secara pukal melalui kaedah kultur tisu dan penghasilan variati baru melalui kaedah mutasi.

Tisu kultur trumpet air, *Cryptocoryne xwillisii* Engler ex Baum, telah dijalankan menggunakan medium Murashige dan Skoog 1962 (MS) yang mengandungi dua pengalak pertumbuhan pokok yang berlainan, iaitu 6-benziladenin purin (BAP) dan  $\alpha$ -naftalina asetik asid (NAA). Tujuh kepekatan yang berlainan BAP (0, 0.5, 1.0, 5.0, 10.0, 20.0 and 40.0  $\mu$ M) dan NAA (0, 0.5, 1.0, 5.0, 10.0, 20.0 and 40.0  $\mu$ M) telah dikaji menggunakan kaedah dua faktor faktorial dengan sepuluh replikasi. Keputusan kajian diambil dan dianalisa selepas 40 hari dikultur.

Keputusan menunjukkan bahawa kesan pengalak tumbesaran tumbuhan untuk meningkatkan bilangan pucuk adalah sangat signifikan (p<0.01). Medium MS yang mengandungi 1.0  $\mu$ M BAP sahaja adalah kepekatan yang optima yang menghasilkan 6.8 ± 1.75 pucuk bagi setiap eksplan. Ini adalah formula yang minima yang boleh meningkatkan bilangan pucuk. Keputusan kajian menunjukkan terdapat dua belas kombinasi hormon yang memberi nilai purata diantara 4 hingga 6 pucuk setiap eksplan. Kesemua eksplan yang dikultur di atas media menghasilkan pucuk (100%).

v

Walaupun, kesemua eksplan memberi tindakbalas yang positif dari segi pertumbuhan namun keputusannya berbeza dari segi bilangan dan saiz pucuk yang dihasilkan. Analisa data dibuat menggunakan ANOVA menunjukkan bilangan pucuk yang terhasil dipengaruhi oleh BAP dan NAA dengan serentak. Ini dicadangkan berdasarkan signifikan interaksi antara BAP dan NAA di mana ia menunjukkan kepekatan BAP memberi kesan yang berbeza ke atas bilangan pucuk pada setiap aras kepekatan NAA dan juga sebaliknya.

Dua variati baru *C. xwillisii* telah berjaya dibangunkan dalam kajian ini melalui kaedah mutagenesis. Pucuk eksplan *C. xwillisii* telah didedahkan kepada satu julat penyinaran Sinaran Gamma <sup>60</sup>Co iaitu 0, 100, 200, 300, 400, 500, 600, 700 dan 800 Gray. Keputusan kajian menunjukkan dos kematian 50% (LD50) untuk tumbuhan tisu kultur *Cryptocoryne xwillisii* ialah pada dos 32 Gray. Dan oleh itu, nilai dos yang lebih rendah dari dos tersebut telah digunakan untuk mengakibatkan mutasi pada tumbuhan.

Sebanyak dua ribu pucuk eksplan telah diradiasi menggunakan dos 25 Gray dan varian-varian dari generasi M<sub>1</sub>, M<sub>2</sub>, M<sub>3</sub> dan M<sub>4</sub> telah dipilih untuk perubahan morfologi. Pucuk-pucuk varian telah disubkultur berulang kali sehingga generasi ke 4 (M<sub>4</sub>), ini bagi memastikan kestabilan mutan tersebut. Walaupun banyak regeneran dengan pelbagai sifat morfologi yang berbeza dihasilkan tetapi hanya dua sahaja mutan yang stabil diperolehi. Mutan yang diperolehi ialah tumbuhan kerdil (D1) dan tumbuhan yang lebih tinggi dari tumbuhan kawalan. Ini telah diverifikasi dari nilai signifikan ujian F dalam ANOVA dimana P<0.05.

Penanda Inter-Retrotransposon Amplified Polymorphism (IRAP) dapat membezakan genome D1 dan G1 dari genome *C. xwillisii* normal. Analisis yang dijalankan mendapati untuk mutan D1, didapati ada dua jalur khusus iaitu jalur 325 bp dan jalur 420 bp dengan menggunakan primer Nikita manakala untuk mutan G1, didapati ada dua jalur khusus iaitu jalur 240 bp dan jalur 300 bp dengan menggunakan kombinasi primer 3'LTR dan primer LTR 6149.

#### ACKNOWLEDGEMENTS

I wish to thank both my supervisors Prof. Dr. Norzulaani Khalid and Prof. Dr. Rofina Yasmin Othman, Institute of Biological Sciences, Faculty of Science, University Malaya for the opportunity to further my study. My experiences doing master project under Prof. Norzulaani and Prof. Yasmin is the most wonderful and precious experience.

My thanks also go to Unit Penyelidikan DiTaja (UPDiT), Institut Pengurusan Penyelidikan dan Perundingan, University Malaya for financial support (Geran P0249/2007A) for my project.

I also wish to thank my employer, the Freshwater Fisheries Research Divison, FRI Glami Lemi, Department of Fisheries for the opportunity to further study and the Department of Public Services for the two years Scholarship 'Hadiah Latihan Persekutuan'.

Last but not least, my special thanks to Kak Lina, Lab Assistant, my fellow lab mates, Nuraini, Malissa, Diana (Dee), Nor Azma, Irma, Sher Ming, Foo, Wan Sin, Ting Teck Kai, Tamil and others. Dedicated to:

My loving husband M.Husni Sulor, and my wonderful children Aisyah, Izzah, Izzati, Firdaus and Safiyya, My both parents and parents in law, And my relatives.

## TABLE OF CONTENTS

	Page
ORIGINAL LITERARY WORK DECLARATION	ii
ABSTRACT	iii
ABSTRAK	v
ACKNOWLEDGEMENTS	viii
LIST OF FIGURES	xi
LIST OF PLATES	xii
LIST OF TABLES	xvi
LIST OF APPENDIX	xvii
ABBREVIATIONS AND ACRONYMS	xviii
CHAPTER 1 INTRODUCTION	XX
CHAPTER 2 LITERATURE REVIEW	XX
CHAPTER 3 MATERIALS AND METHODS	XX
CHAPTER 4 RESULTS	xxi
CHAPTER 5 DISCUSSION	xxi
REFERENCES	xxii
APPENDIX	xxii

#### **LIST OF FIGURES**

- Figure 2.4.1. Estimated production and export value in million Ringgit Malaysia 6 of ornamental fish and aquatic plants from year 2003-2008
- Figure 3.4.3.1. *In vitro* induced mutation for *C*. *xwillisii* adapted from Basiran *et* 21 *al.* (1998).
- Figure 4.21. Bar chart showing the effects of different BA and NAA 30 concentrations on shoot multiplication from single-shoot explants after forty days of culture. Each bar represents the mean (± SD) response from two experiments, each with five replicates per treatment.
- Figure 4.31. Neighbour Joining (NJ) bootstrap consensus tree (replicates = 36 1000) and the tree was unrooted.
- Figure 4.41. Percentage survival of *in vitro* shoot tips *C*. x*willisii* on different 38 high dosage of Gamma ray after 30, 40, and 60 days of irradiations.
- Figure 4.42. Percentage survival of *C. xwillisii* on different low dosage of 39 Gamma ray after 60 days irradiations.
- Figure 4.43. LD<sub>50</sub> for *C. xwillisii* was 25 Gray, best linear line plotted from the 40 different low dosage of Gamma ray after irradiations.

## LIST OF PLATES

Plate 2.3.1.	Cryptocoryne xwillisii	5
Plate 3.3.1	Excised shoot explant (about 1.0 cm) for shoot tip cultures initiation.	17
Plate 4.2.1.	Abnormal <i>Cryptocoryne</i> xwillisii with large petiole and more round leaves.	32
Plate 4.2.2.	Stunted shoots were produced when cultured in high concentration of BA.	32
Plate 4.2.3.	Developmental pathway for <i>C. xwillisii</i> . A. Shoot tip for culture in the MS media. B. Plant regenerates after 20 days culture. C. Plantlets developed after 40 days culture. D. Rooted plantlets of <i>C.</i> <i>xwillisii</i> .	32
Plate 4.4.2.1.	Shoots of Cryptocoryne xwillisii to used in tissue culture	42
Plate 4.4.2.2.	Variant showing albino morphology, 20 Gy $M_1$	42
Plate 4.4.2.3.	Variant show deformation of leaves, 25 Gy M <sub>1</sub>	42
Plate 4.4.2.4.	Variant showing abnormal leaves, wide with light green color leaves $(30 \text{ Gy M}_{\odot})$	42
Plate 4.4.2.5.	Normal plants of <i>Cryptocoryne</i> x <i>willisii</i> in 30 ml universal container, 60 days culture.	42
Plate 4.4.2.6.	Variant showing necrosis symptom when subjected to high dosage of gamma ray.	42
Plate 4.4.2.7.	Irradiated culture explants transferred singly into new 30 ml universal containers	43
Plate 4.4.2.8.	Variant showing small leaves size – with dark green color, the plant length ~ 4 cm, 15 Gy, $M_1$	43
Plate 4.4.2.9.	Variant showing small and spiral leaves – with dark green color, the plant length ~ 4 cm, 20 Gy $M_1$	43
Plate 4.4.2.10.	Variant showing small leaves size – with light green color and chocolate color at the tips, the plant length ~ $2.5$ cm, $30$ Gy, M <sub>1</sub>	43
Plate 4.4.2.11.	Variant showing long leaves size – with light green color, the plant length ~ 6 cm, 20 Gy, $M_1$	43

- Plate 4.4.2.12. Variant showing small leaves size with light green color, the plant 44 length ~ 3.5 cm, 25 Gy M<sub>1</sub>
- Plate 4.4.2.13. Variant showing small leaves size with dark green color, the plant 44 length ~ 5 cm, 25 Gy M1
- Plate 4.4.2.14. Variant showing two types of leaves: normal and spiral leaves 44 with dark green color, the plant length ~ 4 cm, 20 Gy M<sub>1</sub>
- Plate 4.4.2.15. Variant showing spiral leaves with dark green color, the plant 44 length ~ 4 cm, 20 Gy M1
- Plate 4.4.2.16. Variant showing normal leaves size with pinkish color petiole, the 44 plant length ~ 5 cm, 25 Gy  $M_1$
- Plate 4.4.2.17.Variant showing skinny and spiral leaves size with white color,44the plant length ~ 7 cm, 20 Gy M1
- Plate 4.4.2.18. Another variant showing skinny and spiral leaves size with white 45 color, the plant length ~ 7 cm, 20 Gy  $M_1$
- Plate 4.4.2.19. Variant showing small leaves sizes with light green color, the 45 plant length ~ 4 cm, 20 Gy M<sub>1</sub>
- Plate 4.4.2.20. Variant showing thin leaves sizes with brown and green color, the 45 plant length ~ 7 cm, 20 Gy  $M_2$
- Plate 4.4.2.21. Variant showing the leaves with brown veins and tips, the plant 45 length  $\sim$  7 cm, 20 Gy M<sub>1</sub>
- Plate 4.4.2.22. Variant showing the leaves with brownish color and brown veins, 45 the plant length  $\sim 10$  cm, 20 Gy M<sub>4</sub>
- Plate 4.4.2.23. Variant showing the small leaves with light green color, the plant 45 length ~ 4 cm, 30 Gy,  $M_4$
- Plate 4.4.2.24. Cryptocoryne xwillisii (normal plants), mean sizes  $5.18 \pm 1.57$  cm, 46 M<sub>4</sub> generation.
- Plate 4.4.2.25. Mutant D1, plants color are darker green and smaller than normal 46 plants, mean sizes  $3.53 \pm 0.89$  cm, M<sub>4</sub> generation.
- Plate 4.4.2.26. Mutants G1, plants color are brownish and taller than normal plants 46 and the mean sizes  $6.92 \pm 1.79$  cm, M<sub>4</sub> generation.
- Plate 4.4.3.1.IRAP profile of the D1 mutant, G1 mutants and C (control) plant of48Cryptocoryne xwillisiiobtained with primer LTR1 showing

monomorphic bands. L: 100 bp plus DNA ladder.

- Plate 4.4.3.2. IRAP profile of the D1 mutant, G1 mutants and C (control) plant of 48 *Cryptocoryne xwillisii* obtained with primer LTR 6150 showing monomorphic bands. L: 100 bp plus DNA ladder.
- Plate 4.4.3.3. IRAP profile of the D1 mutant, G1 mutants and C (control) plant of 48
   *Cryptocoryne xwillisii* obtained with primer LTR 6149 showing monomorphic bands. L: 100 bp plus DNA ladder.
- Plate 4.4.3.4. IRAP profile of the D1 mutant, G1 mutants and C (control) plant of 48 *Cryptocoryne xwillisii* obtained with primer LTR2 showing monomorphic bands. L: 100 bp plus DNA ladder.
- Plate 4.4.3.5. IRAP profile of the D1 mutant, G1 mutants and C (control) plant of 49
   *Cryptocoryne xwillisii* obtained with primer Sukula showing monomorphic bands. L: 100 bp plus DNA ladder.
- Plate 4.4.3.6. IRAP profile of the D1 mutant, G1 mutants and C (control) plant of 49
   *Cryptocoryne xwillisii* obtained with primer LTR1 x LTR 6150
   showing monomorphic bands. L: 100 bp plus DNA ladder.
- Plate 4.4.3.7. IRAP profile of the D1 mutant, G1 mutants and C (control) plant of 49 *Cryptocoryne xwillisii* obtained with primer LTR1 x LTR2 showing monomorphic bands. L: 100 bp plus DNA ladder.
- Plate 4.4.3.8. IRAP profile of the D1 mutant, G1 mutants and C (control) plant of 49 *Cryptocoryne xwillisii* obtained with primer LTR1 x Nikita showing monomorphic bands. L: 100 bp plus DNA ladder.
- Plate 4.4.3.9.IRAP profile of the D1 mutant, G1 mutants and C (control) plant of50Cryptocoryne xwillisiiobtained with primer LTR1 x Sukulashowing monomorphic bands. L: 100 bp plus DNA ladder.
- Plate 4.4.3.10. IRAP profile of the D1 mutant, G1 mutants and C (control) plant of 50
   *Cryptocoryne xwillisii* obtained with primer LTR 6150 x LTR 6149
   showing monomorphic bands. L: 100 bp plus DNA ladder.
- Plate 4.4.3.11. IRAP profile of the D1 mutant, G1 mutants and C (control) plant of 50
   *Cryptocoryne xwillisii* obtained with primer LTR 6150 x LTR 2
   showing monomorphic bands. L: 100 bp plus DNA ladder.
- Plate 4.4.3.12. IRAP profile of the D1 mutant, G1 mutants and C (control) plant of 50

*Cryptocoryne* x*willisii* obtained with primer LTR 6150 x LTR Nikita showing monomorphic bands. L: 100 bp plus DNA ladder.

- Plate 4.4.3.13. IRAP profile of the D1 mutant, G1 mutants and C (control) plant of 51
   *Cryptocoryne xwillisii* obtained with primer LTR 6150 x LTR
   Sukula showing monomorphic bands. L: 100 bp plus DNA ladder.
- Plate 4.4.3.14. IRAP profile of the D1 mutant, G1 mutants and C (control) plant of 51
   *Cryptocoryne xwillisii* obtained with primer LTR 6149 x LTR
   Nikita showing monomorphic bands. L: 100 bp plus DNA ladder.
- Plate 4.4.3.15. IRAP profile of the D1 mutant, G1 mutants and C (control) plant of 51
   *Cryptocoryne xwillisii* obtained with primer LTR 6149 x LTR
   Sukula showing monomorphic bands. L: 100 bp plus DNA ladder.
- Plate 4.4.3.16. IRAP profile of the D1 mutant, G1 mutants and C (control) plant of 51 *Cryptocoryne xwillisii* obtained with primer LTR 2 x LTR Nikita showing monomorphic bands. L: 100 bp plus DNA ladder.
- Plate 4.4.3.17. IRAP profile of the D1 mutant, G1 mutants and C (control) plant of 52
   *Cryptocoryne xwillisii* obtained with primer LTR 2 x LTR Sukula showing monomorphic bands. L: 100 bp plus DNA ladder.
- Plate 4.4.3.18. IRAP profile of the D1 mutant, G1 mutants and C (control) plant of 52 *Cryptocoryne xwillisii* obtained with primer LTR Nikita x LTR Sukula showing monomorphic bands for G1 and C but no band for D1. L: 100 bp plus DNA ladder.
- Plate 4.4.3.19. IRAP profile of the D1 mutant, G1 mutants and C (control) plant of 52
   *Cryptocoryne xwillisii* obtained with primer Nikita showing two polymorphic bands (arrows). L: 100 bp plus DNA ladder.
- Plate 4.4.3.20. IRAP profile of the D1 mutant, G1 mutants and C (control) plant of 52
   *Cryptocoryne xwillisii* Engler ex Baum obtained with primer LTR1
   x LTR 6149 showing two polymorphic bands (arrows). L: 100 bp
   plus DNA ladder.

### **LIST OF TABLES**

**Table 4.21** Table below showed the effect of different number of shoots at 31

 different level of BA concentrations on all concentrations of NAA

**Table 4.3.1** IRAP primer combinations and the respective annealing 34 temperatures. Ø indicates unsuccessful primer combinations for amplification screening of designated varieties.

**Table 4.3.2.** The degree of polymorphism of the IRAP products within the 35

 Cryptocoryne accessions, \*indicates primer combinations that were used in the experiments.

**Table 4.4.2.1** Analysis of variant (ANOVA) table for mean length (cm) of normal
 41

 and mutants C. xwillisii.
 41

# LIST OF APPENDIX

Table 3.2.1 Composition of culture medium, Murashige and Skoog (MS) Medium	71
(1962)	
Table 3.2.2 Table below showing forty-nine different Plant Growth Regulators	72
(PGRs) combinations in this 7 x 7 factorial experiment and (number) coded for	
the different PGRs combinations.	
<b>Table 3.5.1</b> List of Cryptocoryne sp. that been used in the study.	72
Table 3.5.2 The IRAP primers used in the phylogenetics study of selected	73
Cryptocoryne sp. experiment.	
<b>Table 3.5.3</b> Forty-five combinations of primer A and primer B for IRAP-PCR	73
phylogenetics of selected Cryptocoryne sp. experiment.	
<b>Table 3.6.1</b> The IRAP Primers used in the experiment for mutation detection.	74
Table 3.6.2 Twenty-one combinations of primer A and primer B for IRAP-PCR	74
for mutation detection	
Table 4.2.1 Result of mean number of shoots on various concentration of NAA	75
and BA	
Table 4.4.2.1 Results of mean number of lengths of normal type C. xwillisii.	76
Table 4.4.2.2 Results of mean number of lengths of mutant G1 C. xwillisii.	77
Table 4.4.2.3 Results of mean number of lengths of mutant D1 C. xwillisii.	78
<b>Table 4.4.1.a.</b> Data on number of survival of radiated C. xwillisii after 2 months.	79

# **ABBREVIATIONS AND ACRONYMS**

%	-	Percent
<sup>60</sup> Co	-	Cobalt 60
AFLP	-	Amplified fragment length polymorphism
BAP	-	N6-benzyladenine Purine
NAA	-	1-napthaleneacetic acid
bp	-	base pair
ĊAPs	-	cleaved amplified polymorphism sequences
CTAB	-	hexadecyltrimethylammonium bromide
DF	-	Dilution Factor
DNA	-	Deoxyribonucleic Acid
DOF	-	Department of Fisheries
EDTA	-	Ethylenediaminetetraacetic Acid
ELF	-	extremely low frequency
EtBr	-	Etidium Bromide
FRI	-	Fisheries Research Institute
g/L	-	gram per liter
GDP	-	Gross Domestic Product
Gv	-	Grav
IRAP	-	Inter-retrotransposon amplified polymorphism
kPa	-	kilopascal
LD50	-	Lethal Dose 50%
	-	Linsmaier and Skoog
MGI	-	Malaysia Gross Income
min	-	minute
mM	-	milimolar
MS	_	Murashige and Skoog medium
MSO	-	Murashige and Skoog medium without hormone
NaCl	-	Sodium chloride
NaOCl	-	Sodium Hypo chloride
NaOH	-	Sodium hydroxides
NH₄ acetate	-	Ammonium acetate
°C	-	Degree Celsius
PCR	-	Polymerase Chain Reaction
pmol	-	Pico mol
RAMP	-	randomly amplified microsatellite polymorphisms
RAPD	-	Random amplification of polymorphic DNA
RBIP	-	Retrotransposon-based insertion polymorphism
REMAP	-	REtrotransposon-microsatellite amplified polymorphism
RFLP	-	Restriction fragment length polymorphism
RNAse	-	Ribonuclease
rpm	-	round per minute
rxn	-	reaction mixture
S-SAP	-	sequence-specific amplification polymorphism
SCAR	-	sequence characterized amplified regions
SNPs	-	single nucleotide polymorphism

SRAP	-	sequence-related amplified polymorphism
SSCP	-	single strand conformation polymorphism
TRAP	-	target region amplification polymorphism
Tris HCL	-	Tris Hydrochloric
USD	-	United States Dollar
UV	-	Ultra violet
v/v	-	volume per volume
VLF	-	very low frequency
W/V	-	weight per volume
μg	-	microgram
μl	-	micro liter
μM	-	micro molar
•		

## **CHAPTER 1 INTRODUCTION**

1.0 Introduction	1
1.1 Objectives of the study	2
CHAPTER 2 LITERATURE REVIEW	
2.0 Literature Review	3
2.1 Introduction	3
2.2 Distribution	3
2.3 Morphological Description C. xwillisii	5
2.4 Economic Importance of C. xwillisii	5
2.5 Conventional Propagation of Aquatic Plants	6
2.6 Tissue Culture Aquatic Plants	7
2.7 Mutations	8
2.7.1 Gamma Irradiation	11
2.8 Molecular Markers	12
CHAPTER 3 MATERIALS AND METHODS	
3.0 Materials and Methods	15
3.1 Plants Materials	15
3.1.1 Surface Sterilisation Of Plants Materials	15
3.2 Preparation Of Culture Media	15
3.3 Establishment of Shoot Tips Culture	16
3.3.1 Media Optimisation With Different Plant Growth Regulator (PGR)	17
Combinations	
3.4 Mutagenesis Induction	18
3.4.1 Determination Of Lethal Dose LD <sub>50</sub> (Gamma Irradiation)	18
3.4.2 Dose Rate Calculation For ${}^{60}Co$ unit (Gammacell 220).	19
3.4.3 Gamma Irradiation Experiment	19
3.5 Phylogenetic Studies Using Inter-Retrotransposon Amplified Polymorphism	22
(IRAP)	
3.5.1 Plants Materials	22
3.5.2 DNA Extractions	22

3.5.3 DNA Quantification	24
3.5.4 Gel Electrophoresis Of Genomic DNA	24
3.5.5 IRAP Analysis On Selected Species Of Cryptocoryne	25
3.5.6 Phylogenetic Analyses	26
3.6 IRAP Analysis On Mutants	27
3.6.1 Plants Materials	27
3.6.2 DNA Extraction	27
3.6.3 DNA Quantification	27
3.6.4 IRAP Analysis On Mutant vs Wild Type Of C.xwillisii	27
3.7 Statistical Analysis For Media Optimisation With Different PGR	28
Combinations	
CHAPTER 4 RESULTS	
4.0 Results	29
4.1 Establishment Of Shoot Tip Culture	29
4.2 Determination Of Optimal PGR For Shoot Multiplication	29
4.3 Phylogenetic Study Of Selected Cryptocoryne sp. (Malaysia and Sri Lanka)	33
Using IRAP Analysis	
4.4 Mutation Induction	37
4.4.1 Determination Of Lethal Dose LD50	37
4.4.2 Evaluation Of Gamma Irradiated Plants	41
4.4.3 IRAP Analysis of Mutant and Wild Type Plants	47
<b>CHAPTER 5 DISCUSSION</b>	
5.0 Discussion	53
5.1 Sterilization Technique	53
5.2 PGR Optimization For Shoot Proliferation	54
5.2.1 Absence of BA and NAA	55
5.2.2 Absence of NAA	56
5.2.3 Abscence of BA	57
5.2.4 BA and NAA in combinations	57
5.3 Phylogenetic Study of Selected Cryptocoryne sp. Of Malaysia and Sri Lanka	59
Using IRAP Analysis	
5.4 Determination of Lethal Dose $LD_{50}$	62

5.5 Evaluation Of Gamma Irradiated Plants	63
5.6 IRAP Analysis For Identification Of Sports (Clonal Mutation) / Mutant	63
REFERENCES	65
LIST OF APPENDIX	70