

**MICROPROPAGATION, ANTIOXIDANT AND
ANTIMICROBIAL ACTIVITIES OF
ASPARAGUS OFFICINALIS cv. MARY WASHINGTON *IN
VIVO* AND *IN VITRO***

ARASH KHORASANI ESMAEILI

**DISSERTATION SUBMITTED IN FULFILMENT OF
THE REQUIRMENTS FOR THE DEGREE
OF MASTER OF SCIENCE**

**INSTITUTE OF BIOLOGICAL SCIENCES
FACULTY OF SCIENCE
UNIVERSITY OF MALAYA
KUALA LUMPUR
2012**

This Thesis is dedicated to my Parents
Who supported me for each and every day of my
life since birth,
Enabling such a study to take place today

**UNIVERSITI MALAYA
ORIGINAL LITERARY WORK DECLARATION**

Name of Candidate: Arash Khorasani Esmaeili (I.C/Passport No: D13139190)

(I.C/Passport No: D13139190)

Registration/Matric No: SGR080120

Name of Degree: Master of Science

Title of Project Paper/Research Report/Dissertation/Thesis (“this Work”): Micropropagation, Antioxidant and Antimicrobial Activities of *Asparagus officinalis* cv. Mary Washington *In Vivo* and *In Vitro*

Field of Study: Plant Biotechnology

I do solemnly and sincerely declare that:

- (1) I am the sole author/writer of this Work;
 - (2) This Work is original;
 - (3) Any use of any work in which copyright exists was done by way of fair dealing and for permitted purposes and any excerpt or extract from, or reference to or reproduction of any copyright work has been disclosed expressly and sufficiently and the title of the Work and its authorship have been acknowledged in this Work;
 - (4) I do not have any actual knowledge nor do I ought reasonably to know that the making of this work constitutes an infringement of any copyright work;
 - (5) I hereby assign all and every rights in the copyright to this Work to the University of Malaya (“UM”), who henceforth shall be owner of the copyright in this Work and that any reproduction or use in any form or by any means whatsoever is prohibited without the written consent of UM having been first had and obtained;
 - (6) I am fully aware that if in the course of making this Work I have infringed any copyright whether intentionally or otherwise, I may be subject to legal action or any other action as may be determined by UM.

Candidate's Signature

Date

Subscribed and solemnly declared before,

Witness's Signature

Date

Name:

Designation:

Abstract

In recent years there has been renewed interest in natural medicines that are obtained from plant parts or plant extracts. *Asparagus officinalis* is a herbaceous perennial plant that belongs to Liliaceae family, a valued vegetable for its medicinal properties. The present study was carried out in order to establish an efficient *in vitro* propagation protocol for *Asparagus officinalis*. For this purpose, the nodal explants of *Asparagus officinalis* Cv. Mary Washington were cultured on MS medium containing 3% sucrose and different concentrations of NAA and BAP or IBA and Kn, mixed or separately with range of 0-1.5 mg/l for BAP/Kn and 0-0.5 mg/l for NAA/IBA in order to obtain callus, shoot and root formation. In this study, indirect organogenesis was tested under 2 different conditions (In light and in dark) for the regeneration of *Asparagus officinalis* *in vitro*. After 6 weeks of culture the results showed that 100% of callus formation in 17 of treatments under dark and 3 treatments under light condition. So between dark and light condition, dark condition was found to be more efficient than light condition in promoting callus formation. Also among the two groups of hormones (BAP + NAA, Kn + IBA concentrations), Kn + IBA was reported to be more efficient than BAP + NAA in promoting callus formation. Results also showed that the highest average number of shoots (4.25) of size 4 mm or more per explant, formed under dark condition using 1.5 mg/l BAP mixed with 0.05 mg/l NAA. The formed shoots under dark condition were less developed, with abnormal thick and yellow color compared with the shoots produced under light condition. In light condition the highest average numbers of shoots (3.63) of size 4 mm or more per explant were found on the MS medium supplemented with 0.8 mg/l BAP alone, not in combination with NAA. Rooting was best induced in shoots excised from shoot cultures which were proliferated on MS medium supplemented with an optimal concentration of 0.4 mg/l IBA (2 roots per explant).

In the second part of the study the antioxidant and antibacterial activities of ethanolic extracts of *in vivo* grown *Asparagus officinalis* cv. Mary Washington were investigated using superoxide dismutase, erythrocyte haemolysis and 2,2- diphenyl-1-picrylhydrazil free radical scavenging methods. The measured antioxidant and antimicrobial potential were then compared with the activities shown by the ethanolic extracts of *in vitro* grown *A. officinalis* as well as ethanolic extract of undifferentiated callus cells of *A. officinalis* produced on Murashige and Skoog medium containing 1.5 mg/l 6-benzylaminopurine combined with 0.5 mg/l naphthalene acetic acid. The highest antioxidant capacity was obtained from the *in vivo* grown plant extract followed by *in vitro* grown plant extract in all three examined assays. Although, no antibacterial activity was detected from both *in vivo* and *in vitro* grown plant extracts in the disc diffusion antimicrobial assay, ethanolic extract of *A. officinalis* offered antibacterial activity against *Bacillus cereus*.

Abstrak

Sejak kebelakangan ini terdapat minat atau kecenderungan baru terhadap ubat-ubatan semulajadi yang diperolehi dari bahagian-bahagian tumbuhan atau ekstrak tumbuhan. *Asparagus officinalis* adalah tumbuhan herba saka yang tergolong dalam keluarga Liliaceae, sayur-sayuran penting yang mempunyai nilai perubatan. Kajian ini telah dijalankan dalam usaha untuk menghasilkan regenerasi dan propagasi pesat tumbuhan *Asparagus officinalis*. Untuk tujuan ini eksplan nod *Asparagus officinalis* cv. Mary Washington telah dikultur di dalam media MS (Murashige & Skoog, 1962) yang mengandungi 3% sukrosa serta NAA dan BAP atau IBA dan kinetin, samada dalam kombinasi atau secara berasingan dengan kepekatan antara 0-1.5 mg/l BAP/Kn beserta 0-0.5 mg/l NAA/IBA untuk mendapatkan kalus, pucuk dan pembentukan akar. Dalam kajian ini, organogenesis secara tidak langsung diuji dibawah dua keadaan berbeza iaitu gelap dan cahaya untuk memperolehi regenerasi *Asparagus officinalis* *in vitro*. Selepas enam minggu dikultur, keputusan menunjukkan 100% pembentukan kalus dari 17 perlakuan dibawah keadaan gelap dan 3 perlakuan sahaja dibawah keadaan cahaya. Oleh itu, antara keadaan gelap dan cahaya, didapati keadaan gelap lebih efisien berbanding dibawah cahaya dalam pembentukan kalus bagi spesies ini. Diperhatikan juga, antara dua kumpulan hormon (BAP dan NAA; Kn dan IBA), Kn dan IBA adalah lebih berkesan daripada BAP dan NAA dalam mempromosi pembentukan kalus. Hasil keputusan juga menunjukkan bahawa bilangan purata pucuk yang paling tinggi (4.25) bersaiz 4 mm atau lebih bagi setiap eksplan terbentuk di bawah keadaan gelap dengan penambahan 1.5 mg/l BAP yang dicampur dengan 0.05 mg/l NAA kedalam media. Pucuk yang terbentuk di bawah keadaan gelap kurang sempurna atau tidak berkembang dengan baik, dengan warna yang tidak normal serta tebal dan kuning berbanding dengan pucuk yang dihasilkan di bawah keadaan cahaya.

Dalam keadaan cahaya, bilangan purata tertinggi bagi pucuk (3.63), bersaiz 4 mm atau lebih bagi setiap eksplan diperhatikan pada media MS yang ditambah dengan 0.8 mg/l BAP sahaja bukan dalam kombinasi dengan NAA. Pertumbuhan akar yang terbaik pada pucuk pula dihasilkan daripada kultur pucuk yang dikultur dalam media MS yang ditambah dengan IBA pada kepekatan optima iaitu 0.4 mg/l (2 akar per eksplan).

Dalam kajian seterusnya, antioksidan dan aktiviti antibakteria dari ekstrak etanolik dari tumbuhan *in vivo* *Asparagus officinalis* cv. Mary Washington telah dijalankan menggunakan superoxide dismutase, haemolisis eritrosit dan “2,2,- Diphenyl-1-picrylhydrazil free radical scavenging methods”. Antioksidan yang diukur dan potensi antimikrobial kemudiannya dibandingkan dengan aktiviti-aktiviti yang ditunjukkan oleh ekstrak etanolik dari tumbuhan yang dikultur secara *in vitro* dan juga dari tisu kalus yang dihasilkan di atas MS media yang mengandungi 1.5 mg/l 6-benzylaminopurine bersama 0.5 mg/l asid naftalena asetik. Kapasiti antioksidan tertinggi telah diperolehi dari ekstrak tumbuhan yang ditanam secara *in vivo*, diikuti dengan ekstrak *in vitro* dalam ketiga-tiga sampel yang diperiksa. Walaupun tiada aktiviti antibakteria yang dikesan dari kedua-dua ekstrak tumbuhan yang ditanam secara *in vivo* dan *in vitro* menggunakan “ disc diffusion antimicrobial assay”, tetapi ekstrak etanolik dari *Asparagus officinalis* menunjukkan aktiviti antibakteria terhadap bakteria *Bacillus cereus*.

Acknowledgments

I wish to express my sincere thanks to my supervisors,
Prof. Dr. Rosna Mat Taha and En. Wirakarnain Sani.

This thesis would not have been completed without his expert advice and
unfailing patience. I am also most grateful for his faith in this study
especially in difficult circumstances and those hard times of work.

It's my pleasure to offer my thanks to all my friends and laboratory mates
especially, Arash Rafat, Behrooz Banisalam, Reza Ebrahimi and Imdadul Haq
for maintaining a pleasant research atmosphere and making my stay in
Malaysia an enjoyable one.

I wish to thank Dr. Koshy Philip from
Microbiology Division, Faculty Science, University Malaya for his great
help and support.

I would like to thank University of Malaya for the use of laboratory facilities and for
providing research grant PS175/2009A.

Last but not least, it is difficult to word my gratitude towards my
family members for their encouragement and support
during this period. I am grateful to my parents for
supporting me in every possible ways.

Table of Contents

Title	page
Abstract	iii
Abstrak	v
Acknowledgment	vii
List of Figures.....	xi
List of Tables.....	xiv
List of symbols and abbreviations.....	xv

CHAPTER I

Introduction

General Introduction	1
1.1 Morphology of <i>Asparagus officinalis</i>	2
1.1.1 History of Asparagus	5
1.1.2 Asparagus in Malaysia	7
1.1.3 Importance of Asparagus	8
1.1.4 Properties of Asparagus	10
1.2 Plant biotechnology	10
1.2.1 The future of plant biotechnology.....	11
1.3 Importance of bioactivities discovery from plants.....	11
1.4 Problem statements.....	13
1.5 The objectives of study.....	13

CHAPTER II

Literature Review

2.1 Plant biotechnology and tissue culture	15
2.1.1 Benefits of tissue culture	16
2.2 Explant materials	18
2.3 Growth regulators	18
2.4 Media	20
2.5 <i>In vitro</i> propagation of medicinal plants.....	21
2.5.1 <i>In vitro</i> propagation of <i>Asparagus</i>	27
2.6 Importance of antioxidant.....	30
2.6.1 Antioxidant assay.....	31
2.6.2 Antioxidant activity of plants.....	33
2.7 Importance of antibacterial.....	34
2.7.1 Antibacterial assay.....	35
2.7.2 Antimicrobial activity of plants.....	35
2.8 Antioxidant and antibacterial activity of <i>Asparagus officinalis</i>	37

CHAPTER III

Materials and methods	
3.1 Study location	40
3.2 Micropropagation of <i>Asparagus officinalis</i>	40
3.2.1 Plant material and Explant sources	40
3.2.2 Explants sterilization.....	41
3.2.3 Media and culture vessels.....	41
3.2.4 Aseptic Manipulation and Inoculation.....	45
3.2.5 Treatments.....	46
3.2.6 Callus induction and shoot multiplication.....	46
3.2.7 Rooting of shoots.....	49
3.3 Antioxidant and antibacterial activities.....	50
3.3.1 Ethanolic extract preparation.....	50
3.3.2 Antioxidant activity.....	51
3.3.2.1 Superoxide dismutase (SOD) assay.....	51
3.3.2.2 Erythrocytes haemolysis prevention assay	51
3.3.2.3 Radical scavenging capacity assay (DPPH).....	52
3.3.3 Antibacterial activity assay.....	53
3.4 Statistical analysis.....	54

CHAPTER IV

Results

4.1 Regeneration	56
4.1.1 Effect of plant growth regulators on callus formation of <i>Asparagus officinalis</i>	56
4.1.2 Effect of plant growth regulators on shoot regeneration of <i>Asparagus officinalis</i>	68
4.1.3 Root development	75
4.2 Antioxidant activity	77
4.2.1 SOD assay	77
4.2.2 Erythrocytes haemolysis assay.....	78
4.2.3 DPPH* scavenging activity assay.....	79
4.3 Antibacterial activity assay.....	80

CHAPTER V

Discussion

5.1 Regeneration	83
5.2 Antioxidant and Antibacterial activities.....	88

CHAPTER VI

6. Conclusion remarks and recommendation	92
References	94

List of Figures

Figure		page
1.1	<i>Asparagus officinalis</i> in the field	4
1.2	<i>Asparagus officinalis</i> plant	4
1.3	<i>Asparagus officinalis</i> leave	5
1.4	Producer Price (USD/tone) of <i>Asparagus</i> in Malaysia 1991-2008	8
1.5	Spears of green <i>Asparagus</i> in the market	9
3.1	<i>Asparagus officinalis</i> cv. Mary Washington in the green house at Institute of Biological sciences University of Malaya	40
3.2	The nodal explant from 4 weeks old plant, cultured in MS medium	47
3.3	Flasks contain cultured explants in MS medium, placed in the growth room	48
3.4	Uniform shoot cuttings from the multiplication trial experiments cultured in MS medium for rooting	49
3.5	Uniform shoot cuttings from the multiplication trial experiments cultured in MS medium for rooting	50
4.1(I)	Callus formation in light condition, in MS medium supplemented with 0.8 mg/L BAP + 0.1 mg/L NAA, after 3 weeks of culture	57
4.1(II)	Callus formation in light condition, in MS medium supplemented with 0.8 mg/L BAP + 0.1 mg/L NAA, after 6 weeks of culture	57
4.2(I)	Callus formation in dark condition, in MS medium supplemented with 0.4 mg/L Kn + 0.2 mg/L IBA, after 3 weeks of culture	58

4.2(II)	Callus formation in dark condition, in MS medium supplemented with 0.4 mg/L Kn + 0.2 mg/L IBA, after 6 weeks of culture	58
4.3	Percentage of callus formation in MS media with different concentration of BAP + NAA under dark condition, after 6 weeks	61
4.4	Percentage of callus formation in MS media with different concentrations of Kn + IBA under light condition, after 6 weeks	63
4.5	Percentage of callus formation in MS media with different concentrations of Kn + IBA under dark condition, after 6 weeks	65
4.6	Percentage of callus formation in MS media with different concentrations of BAP + NAA under light condition, after 6 weeks	67
4.7	Shoot regeneration from cultured explant on MS medium supplemented with 1.5 mg/L BAP and 0.05 mg/L NAA under dark condition, after 6 weeks	69
4.8	Shoot regeneration from cultured explant on MS medium supplemented with 1.5 mg/L BAP and 0.05 mg/L NAA under dark condition after 6 weeks	69
4.9	Shoot regeneration from cultured explant on MS medium supplemented with 0.8 mg/L BAP under light condition after 6 weeks	70
4.10	Mean number of shoots per explant in MS media with different concentrations of BAP + NAA under light condition, after 6 weeks	71
4.11	Mean number of shoots per explant in MS media with different concentrations of BAP + NAA under dark condition, after 6 weeks	72
4.12	Mean number of shoots per explant in MS media with different concentrations of Kn + IBA under light condition, after 6 weeks	73

4.13	Mean number of shoots per explant in MS media with different concentrations of Kn + IBA under dark condition, after 6 weeks	74
4.14	Mean number of roots per explant in MS media containing different concentrations of IBA, after 6 weeks	75
4.15	Root development in MS medium supplemented with 0.4 mg/L IBA after 6 weeks of culture	76
4.16	Antioxidant activity of plant extracts (<i>in vivo</i> , <i>in vitro</i> and callus) of <i>Asparagus officinalis</i> measured using superoxide dismutase (SOD) assay presented as percentage of inhibition rate. Ascorbic acid (1 mg/L) was used as the positive control. The data were analyzed by one-way ANOVA and the inhibition rate means of samples were compared using Duncan's Multiple Comparison Test (DMCT). Mean of different samples labeled with different letters are significantly different ($p < 0.05$).	77
4.17	Antioxidant activity of examined plant extracts (<i>in vivo</i> plant, <i>in vitro</i> plant and callus) of <i>Asparagus officinalis</i> measured using rabbit erythrocytes haemolysis assay are stated as percentage value. Ascorbic acid (1 mg/L) was used as the positive control. The data were analyzed by one-way ANOVA and the haemolysis percentage means of samples were compared using Duncan's Multiple Comparison Test (DMCT). Mean of different samples labeled with different letters are significantly different ($p < 0.05$).	78
4.18	Antioxidant activity of examined plant extracts (<i>in vivo</i> plant, <i>in vitro</i> plant and callus) of <i>Asparagus officinalis</i> measured using DPPH* scavenging activity assay presented as percentage value. Ascorbic acid (1 mg/L) and BHT (1 mg/L) were applied as the positive controls. The data were analyzed by one-way ANOVA and the DPPH* scavenging activity percentage means of samples were compared using Duncan's Multiple Comparison Test (DMCT). Mean of different samples labeled with different letters are significantly different ($p < 0.05$).	79
4.19	comparison of antibacterial activity of <i>in vitro</i> , <i>in vivo</i> and callus extract of <i>Asparagus officinalis</i> in <i>Bacillus cerus</i> , using paper disk diffusion method	80

List of Tables

Table		page
1.1	Taxonomic classification of <i>Asparagus officinalis</i>	2
3.1	MS supplemented with Different Combinations of BAP and NAA during callus induction and shoot culture of <i>Asparagus officinalis</i>	43
3.2	MS supplemented with different combinations of Kn and IBA during callus induction and shoot culture of <i>Asparagus officinalis</i>	44
4.1	Inhibition effect of 100 mg/ml of <i>Asparagus officinalis</i> ethanolic extracts (<i>in vivo</i> plant, <i>in vitro</i> plant and callus) against the growth of four pathogenic bacteria	81

LIST OF ABBREVIATIONS

2,4 D	2,4-Dichlorophenoxyacetic acid
ABTS	2,2'-azinobis[3-ethylbenzothiazoline-6-sulphonate
ANOVA	Analysis of variance
B	Boron
BAP	6-Benzylaminopurine
BHA	Butylated hydroxyanisole
BHT	Butylated hydroxytoluene
°C	Degree celsius
Ca	Calcium
Cl	Chloroform
Cu	Copper
dm	decimeter
DPPH	2,2-diphenyl-1-picrylhydrazil
DNA	Deoxyribonucleotic acid
DMCT	Duncan's Multiple Comparison Test
ESR	Electron Spin Resonance
FRAP	Ferric reducing ability of plasma
Fe	Iron
g	gram
Hcl	Hydrochloride acid
IAA	Indole-3-Acetic Acid
IBA	Indole-3-Butric Acid
K	Potassium
Kn	kinetin
KJ	Kilojoule
Kcal	Kilocalorie
L	Litter
Mg	Magnesium
mg	milligram
MH	Mueller Hinton broth
Mn	Manganese

MS	Murashige and Skoog
N	Nitrogen
Na	Sodium
P	Phosphate
PBS	phosphate buffer solution
PGRs	Plant Growth Regulators
pCPA	p-chlorophenoxyacetic acid
ROS	Reactive Oxygen Species
SOD	Superoxide Dismutase
TEAC	Trolox equivalent antioxidant capacity
Zn	Zinc