

## Appendix 1

### Material

### Samples

Water and sediment were collected from Kuala Sepetang, Taipain, Malaysia. Water and sediment samples were collected in eight different stations from Kuala Sepetang, Kuala Sangga Besar and Kuala Selinsing river of Matang mangrove estuarine.

### Media

All the media used for this study were obtained Oxoid Ltd, England and CHROMagar Ltd, France.

### General media

### Nutrient agar

Yeast extraction	2 g
Peptone	5g
Sodium Chloride	5g
Agar	15g
Distilled water	1000ml

(PH=7.4 ± 0.2 )

The formula for preparation is 28g/l. The medium is needed to be autoclaved before using.

### **Luria Bertani Broth**

Yeast extraction	0.5g
Sodium Chloride	0.5g
Tryptone	1g
Distilled water	100ml

PH=7.5

The ingredients were mixed and boiled till 100C° and autoclaved.

### **Luria-Bertani Agar**

Yeast extraction	0.5g
Sodium chloride	0.5g
Tryptone	1g
Agar	1.5g
Distilled water	100ml

PH=7.5

The ingredients were mixed and boiled till 100C° and autoclaved.

## **Enrichment Media**

### **Buffered Peptone Water (BPW)**

Peptone	10g
Sodium chloride	5g
Di-Sodium phosphate	3.5g
Potassium dihydrogen phosphate	1.5g
Distilled water	1000ml

The medium was boiled and autoclaved before using

## **Selective and Differentiated Media**

### **CHROM™ orientation**

Agar	15g
Yeast Extract	17g
Chromogenic Mix	1g
Distilled water	1000ml

PH= 7±2

The proportion of 33g/l was prepared using distilled water. The medium was boiled and autoclaved before using.

### **CHROMagar™ ECC**

Agar	15g
Peptone and Yeast extract	8g
NaCl	5g
Chromogenic Mix	4g
Distilled water	1000ml

PH= 7.2±0.2

The proportion of 32.8g/l was prepared using distilled water. The medium was boiled and used without autoclaving

### **MacConky**

Peptone	20g
Lactose	10g
Bile salt	5g
Sodium chloride	5g
Neutral red	0.075g
Agar	12g

Distilled water 1000ml

PH=7.5±0.2

The proportion of 52.075% g/l was prepared using distilled water. The medium was boiled and autoclaved before using.

### **Eosin-Methylene Blue (EMB)**

Peptone 10g

Sucrose 10g

Lactose 5g

Dipotassium phosphate 2g

Eosine Y 0.4g

Methylene Blue 0.065g

Agar 14g

Distilled water 1000ml

PH=7.1

The proportion of 36.4 g/l was prepared using distilled water. The medium was boiled and autoclaved before using

### **Material Biochemical Test**

## **Gram staining**

Glass slide

## **Reagents**

Crystal Violet dye

Iodine

Ethanol 95%

Safranin

Water

## **Oxidase test**

N, N, N', N' –tetra methyl-p-phenylenediamine dihydrochloride (7.95%) powder

## **MR-VP**

Peptone 7g

Glucose 5g

Phosphate buffer 5g

Distilled water 1000ml

PH 6.9

The proportion of 17g/l was prepared using distilled water. The medium was boiled and autoclaved before using

### **Methyl Red reagent**

Methyl Red	0.1g
95% ethyl alcohol	300 ml
Distilled water	500ml

### **Kovacs reagent**

Isoamyl alcohol	150 ml
Concerntated Hydrochloric Acid	50 ml
$\rho$ -dimethylaminobenzaldehyde	10 g

### **Alpha Naphthol solution**

Purified a- naphthol	5 g
Ethyl alcohol	100 ml

### **Sulfide Indole Motility (SIM)**

Tryptone	20g
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Peptone	6.1g
Ferric Ammonium Sulphate	0.2g
Sodium Chloride	0.2g
Agar	3.5g
Distilled water	1000ml

PH:  $7.3 \pm 0.2$

The proportion of 30g/l was prepared using distilled water. The medium was boiled and autoclaved before using

### **Simmons Citrate**

Magnesium sulfate	0.2 g
Monoammonium phosphate	1 g
Dipotassium phosphate	1g
Sodium citrate	2g
Sodium chloride	5g
Bromthymol Blue	0.08 g
Agar	15 g
Distilled water	1000ml



PH=  $6.9 \pm 0.2$

The proportion of 24.28g/l was prepared using distilled water. The medium was boiled and autoclaved before using

**H<sub>2</sub>O<sub>2</sub> 3%**

Hydrogen peroxide 30g

Distilled water 1000ml

**KOH 3%**

Potassium hydroxide 30g

Distilled water 1000ml

**3.1.3.11. KOH 40%**

Potassium hydroxide 40 g

Distilled water 100 ml

**Other reagent**

**Saline Buffred water (0.85%)**

NaCl	0.85g
Distilled water	1000ml

**Alcohol 70% for 500 mL (from 95%)**

95% alcohol	395ml
Distilled water	132ml

**PCR Material**

**Oligonucleotide primer**

The oligonucleotide primers used in this study for PCR assay and the expected size for PCR product are listed in Table 1. *phoA* primer was used to detect *E.coli* housekeeping gene in monoplex PCR assay.

**Chemicals and Enzymes**

All the PCR enzymes and chemicals used in this were purchased from Promega Corporation, USA.

Colourless and Green GoTaq® Flexi Buffer	5x
Deoxynucleotide triphosphates (dNTPs)	10mM
Magnesium Chloride (MgCl <sub>2</sub> )	25mM

Taq polymerase (GoTaq® DNA polymerase)

5U/1µl

### **Material for Agarose GEL Electrophoresis**

#### **Agarose Gel (1.5%)**

Agarose gel

1.5 g

0.5x TBE buffer

100ml

#### **Gel Loading Dye, 100bp DNA ladder and 1kb DNA ladder**

6x Loading Dye, 100bp DNA ladder and 1kb DNA ladder were purchased from Promega Corporation, USA.

#### **10x Tris Borate EDTA Buffer (TBE)**

Boric acid

61.8 g

Tris

121.2 g

Na<sub>2</sub>EDTA.2H<sub>2</sub>O

0.745 g

Distilled water

1000ml

The solution PH was adjusted to 8.3. 50ml of 10 x TBE was then diluted into 950ml of distilled water for preparing 0.5x TBE.

### **Ethidium Bromide**

Ethidium Bromide	100 mg
Deionised water	10ml

The solution was stored in a dark bottle at room temperature and diluted to 0.5 $\mu$ g/ml with distilled water before use.

### **Other reagent**

#### **Phosphate buffered saline (PBS)**

PBS tablets	10
Distilled water	1000ml

The solution pH was adjusted to 8.3.

#### **Tris-EDTA (TE) buffer**

1M Tris	10 ml
0.5M EDTA	2 ml
Distilled water	1000 ml

The solution pH was adjusted to 8

## Mater mixture for PCR experiment

**Table 1.1.** The PCR conditions for monoplex PCR

Material	Stock concentration	Working concentration	Volume ( $\mu$ l)
Green buffer	5x	1x	5.00
MgCl <sub>2</sub>	25mM	1mM	1.00
dNTPs	10mM	140 $\mu$ M	0.35
PhoA F	10mM	0.1 $\mu$ M	0.25
PhoA R	10mM	0.1 $\mu$ M	0.25
Taq polymerase	5U/ $\mu$ l	0.5U	0.10
Template			5.00
ddH <sub>2</sub> O			11.925
Total volume ( $\mu$ l)			25.00

**Table 1.2.** PEP-PCR Mater mixture

Material	Stock concentration	Working concentration	Volume ( $\mu$ l)
Colorless buffer	5x	1x	5.00
MgCl <sub>2</sub>	25mM	2.5mM	2.5
dNTPs	10mM	200 $\mu$ M	0.5
REP-Primer	10mM	0.5 $\mu$ M	1.25
Taq polymerase	5U/ $\mu$ l	1U	0.20
Template			5.00
ddH <sub>2</sub> O			10.55
Total volume ( $\mu$ l)			25.00

**Table 1.2.** Mater mixture condition for 16S rDNA

Material	Stock concentration	Working concentration	Volume ( $\mu$ l)
Green buffer	5x	1x	5.00
MgCl <sub>2</sub>	25mM	1mM	1.00
dNTPs	10mM	140 $\mu$ M	0.35
PhoA F	10mM	0.08 $\mu$ M	0.2
PhoA R	10mM	0.08 $\mu$ M	0.2
Taq polymerase	5U/ $\mu$ l	0.5U	0.10
Template			3.00
ddH <sub>2</sub> O			15.15
Total volume ( $\mu$ l)			25.00

## Appendix 2

REP-PCR gel using REP-PCR primer

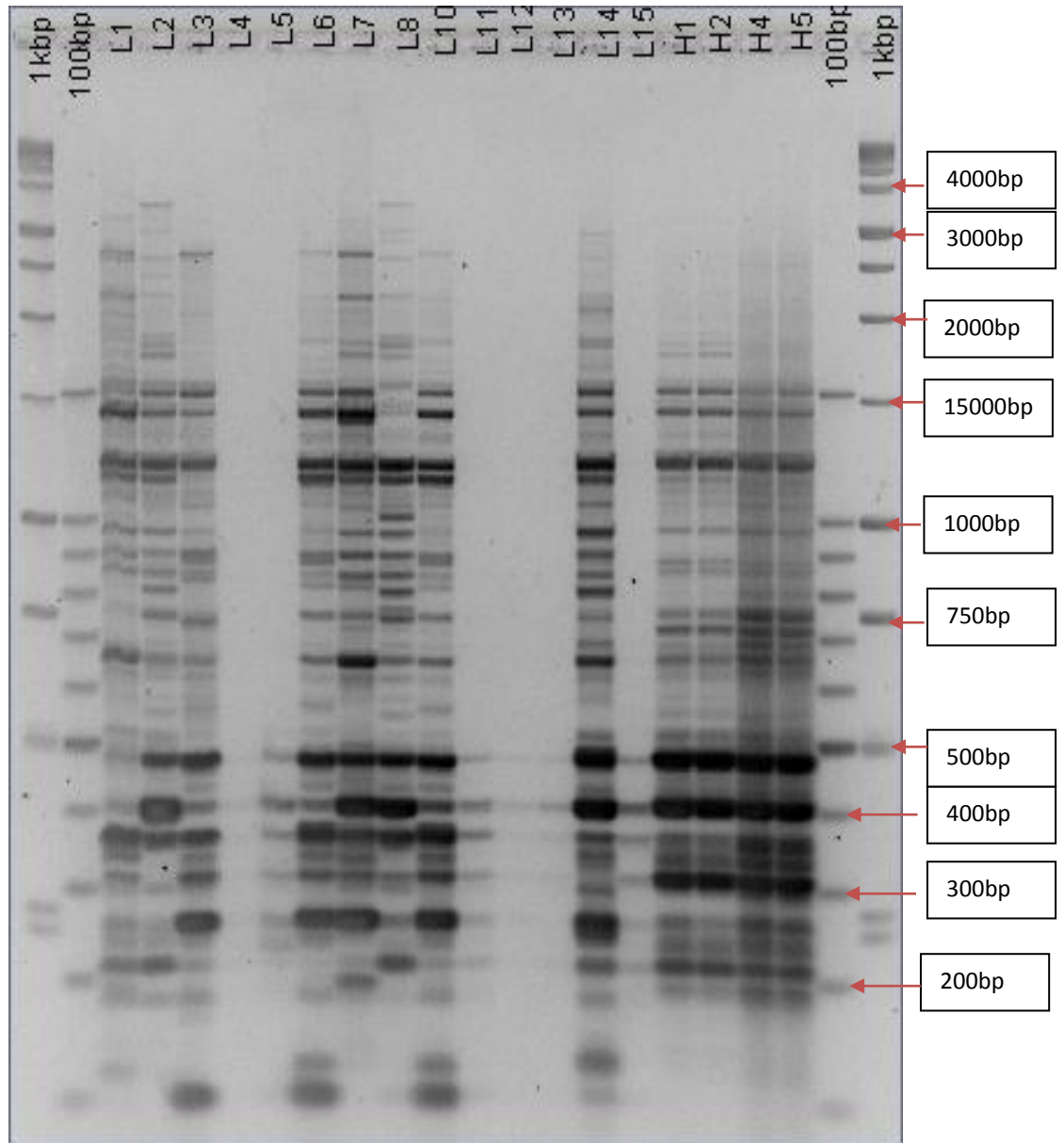


Fig1. Rep-PCR gel, L1, L2, L3, L4, L5, L6, L7, L8, L10, L11, L12, L13, and L15 strains were isolated from station H; H1, H2, H4 and H5 were isolated from station E.

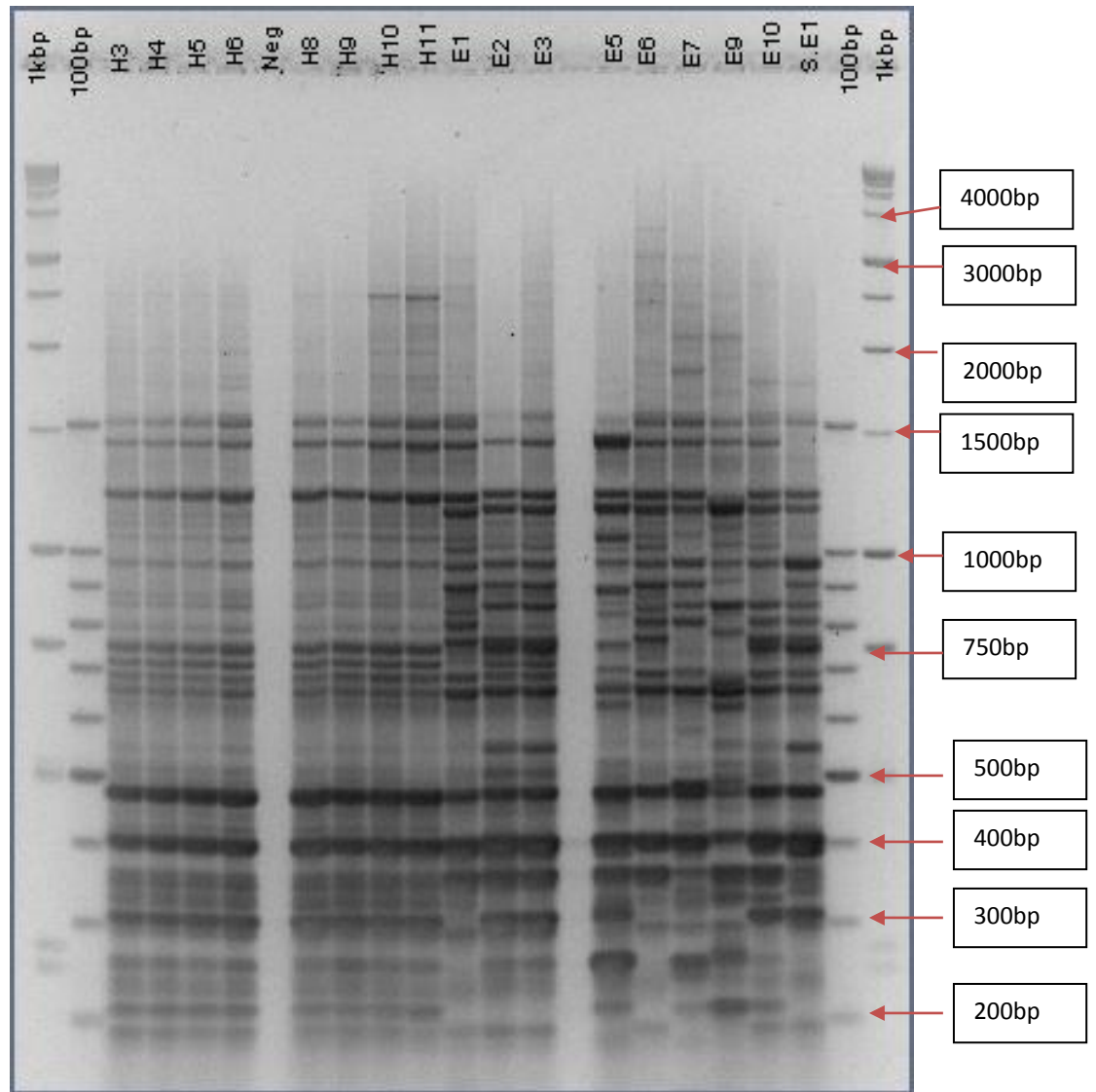


Fig 2. Rep-PCR gel, H3, H4, H5, H6, H8, H9, H10, and H11 were isolated from station E; E1, E2, E3, E5, E6, E7, E9, E,10 and S.E1 strains were isolated from station D.

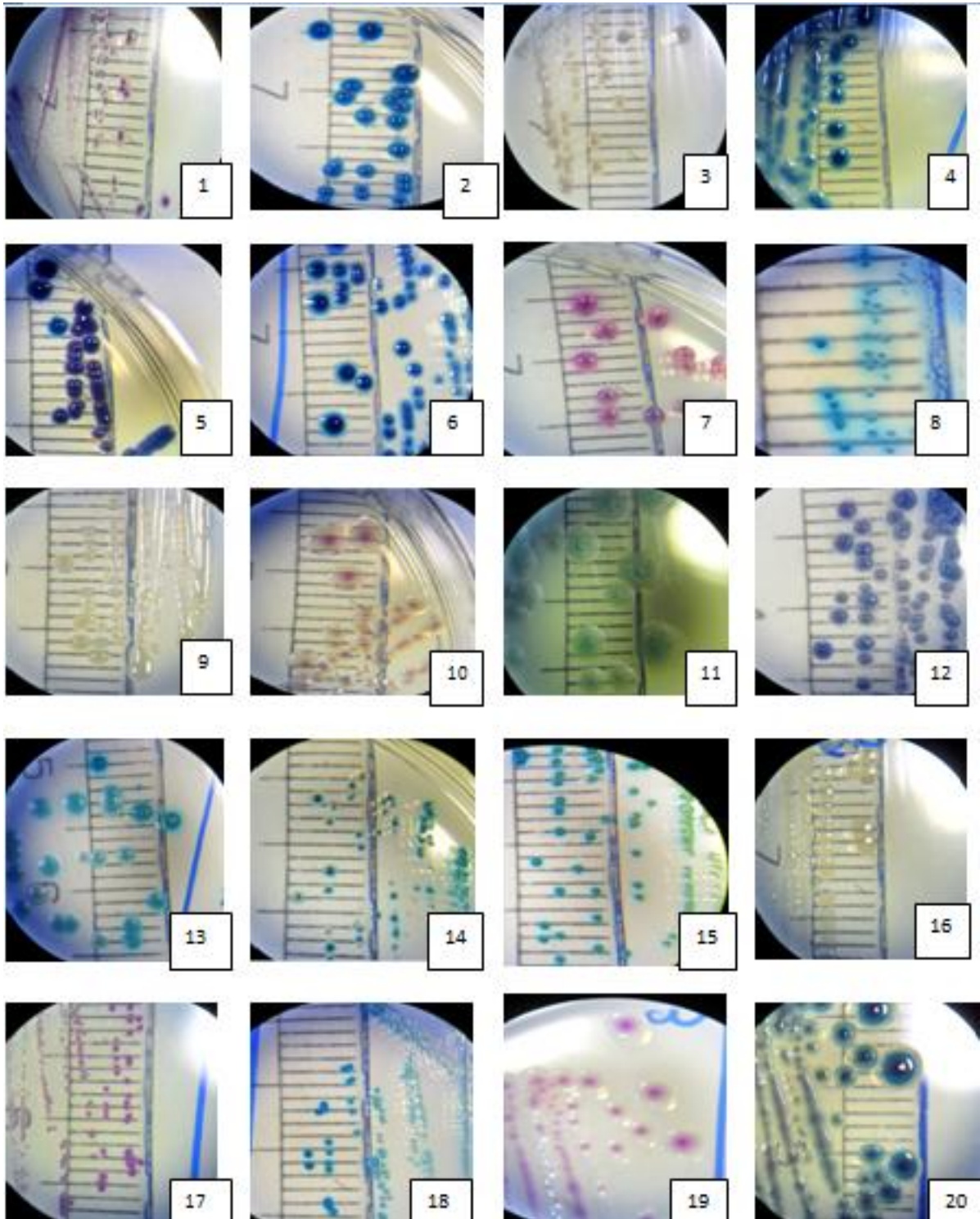


### Appendix 3

Some pictures that have been taken during sampling time in Kula Sepetang estuary



Appendix 4



1,*C. Koseri*; 2,*E. aerogenes*; 3,*H.alive*; 4,*K. Oxytoca*; 5, *S. marcescens*; 6,*R. Planticola*; 7,*E.coli*; 8,*Entrococcus casseliflavus*; 9,*Acinetobactor johansonii*; 10,*Aeromonas sp*; 11,12,13,*Bacillis sp*; 14,*Exigubacteria sp*; 15,*Paenibacillus papillae*; 16,*Pesudomonase sp*, 17,*Staphylococcus sciuris* and ,*S. hominis ss hominis* and *S.saprophyticus*; 18, *Stenotrophomonas rhizophila*; 19,*Vibrio sp*; 20, *Vibrio sp* at room temperature

# The color wheel



**View Details**

Project: ML5  
 Plate Number: 1  
 Plate Type: GEN III  
 Protocol: A  
 Strain Type:  
 Incubation Hours: 24  
 Sample ID: 366  
 Read Hour: 24h  
 Source: prawn gut  
 Temperature:  
 Agar Media:  
 Replicate:  
 Age of Isolate:  
 Passage:  
 Field 9:  
 Field 10:

Pos/Neg Graphic | Pos/Neg Numerical | ODs

	1	2	3	4	5	6	7	8	9	10	11	12
A	○	○	○	○	○	○	○	○	○	○	○	○
B	○	○	○	○	○	○	○	○	○	○	○	○
C	○	○	○	○	○	○	○	○	○	○	○	○
D	○	○	○	○	○	○	○	○	○	○	○	○
E	○	○	○	○	○	○	○	○	○	○	○	○
F	○	○	○	○	○	○	○	○	○	○	○	○
G	○	○	○	○	○	○	○	○	○	○	○	○
H	○	○	○	○	○	○	○	○	○	○	○	○

H12 Sodium Bromate

Species ID: *Hafnia alvei*

	PROB	SIM	DIST	Organism Type	Species
==>1	0.748	0.560	3.606	GN-Ent	<i>Hafnia alvei</i>
2	0.155	0.100	5.121	GN-Ent	<i>Yokenella regensburgei</i>
3	0.059	0.035	6.046	GN-Ent	<i>Enterobacter cancerogenus</i>
4	0.038	0.021	6.466	GN-Ent	<i>Salmonella enterica</i> (gp6) ss indica

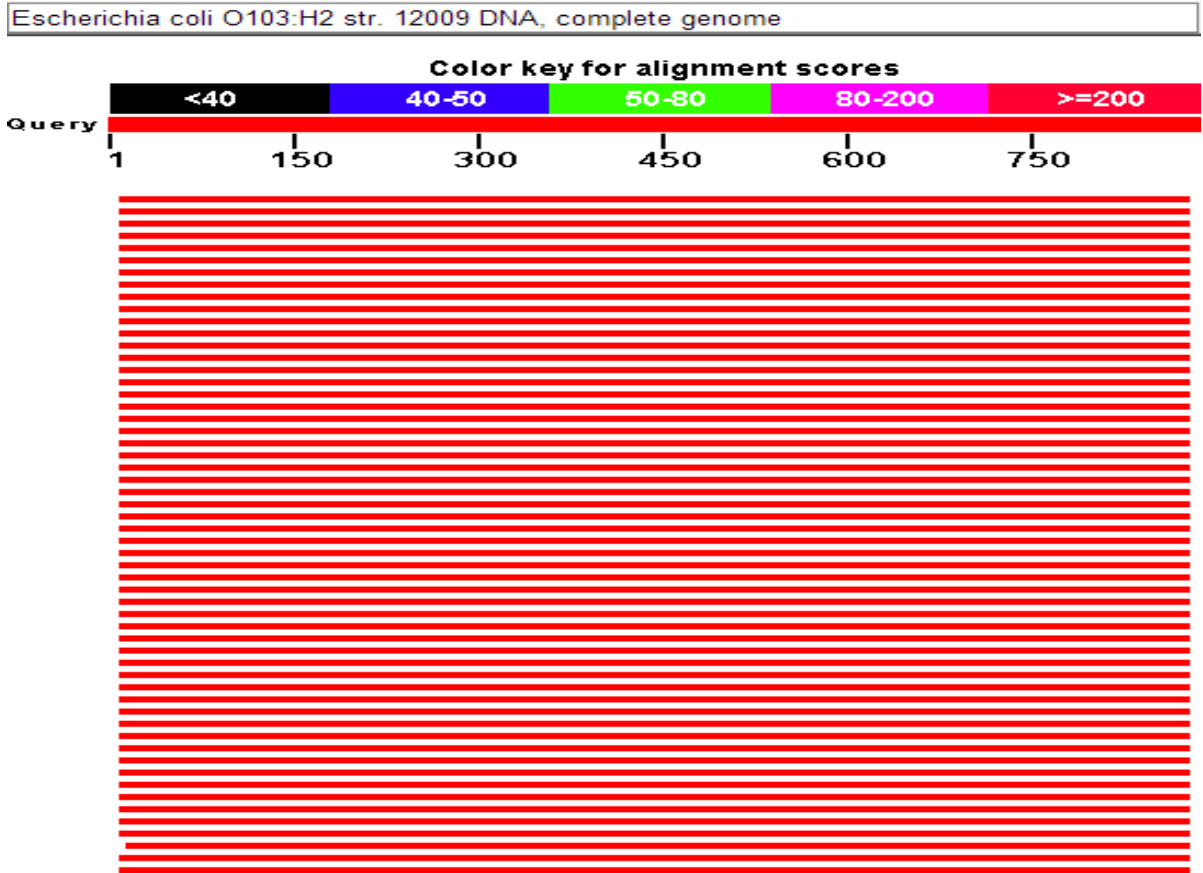
Compare Data To Other Species

View Selected Species  
 Compare To Other Species  
 Clear Other Species  
 Print Preview  
 Print  
 Close

Biochemical profile using Biolog Gen III to identify *Hafnia alvei*

## Appendix 5

*E. coli*; *phoA* BLAS sequencing result (*phoA* chromatogram result)



*E. coli*; *phoA* BLAS sequencing result

Sequences producing significant alignments:						
Sequences producing significant alignments:						
Description	Max score	Total score	Query cover	E value	Max ident	Accession
<i>Escherichia coli</i> O103:H2 str. 12009 DNA, complete genome	1568	1568	97%	0	99%	AP010958.1
<i>Escherichia coli</i> HS, complete genome	1568	1568	97%	0	99%	CP000802.1
<i>Escherichia coli</i> W, complete genome	1563	1563	97%	0	99%	CP002967.1
<i>Escherichia coli</i> KO11, complete genome	1563	1563	97%	0	99%	CP002516.1
<i>Escherichia coli</i> W, complete genome	1563	1563	97%	0	99%	CP002185.1



*Enterococci* 16S r RNA BLAST sequencing result

Sequences producing significant alignments:						
Sequences producing significant alignments:						
Description	Max score	Total score	Query cover	E value	Max ident	Accession
<i>Enterococcus casseliflavus</i> gene for 16S ribosomal RNA, partial sequence, strain: CN1	990	990	97%	0	99%	AB699730.1
<i>Enterococcus casseliflavus</i> gene for 16S rRNA, partial sequence, strain: C507	990	990	97%	0	99%	AB618809.1
<i>Enterococcus casseliflavus</i> gene for 16S rRNA, partial sequence, strain: RZC110	989	989	97%	0	99%	AB671565.1
<i>Enterococcus casseliflavus</i> EC20, complete genome	987	4925	98%	0	99%	CP004856.1
<i>Enterococcus casseliflavus</i> strain Z6006 16S ribosomal RNA gene, partial sequence	987	987	98%	0	99%	KC212047.1

*Klebsiella*; 16 S r RNA BLAST sequencing result

Sequences producing significant alignments:						
Sequences producing significant alignments:						
Description	Max score	Total score	Query cover	E value	Max indent	Accession
<i>Klebsiella</i> sp. SR1.6 16S ribosomal RNA gene, partial sequence	1000	1000	99%	0	99%	JQ912555.1
<i>Bacterium</i> 5-2(2013) 16S ribosomal RNA gene, partial sequence	1000	1000	99%	0	99%	KC753506.1
<i>Klebsiella</i> sp. SR-143 16S ribosomal RNA gene, partial sequence	1000	1000	99%	0	99%	KC455430.1
<i>Klebsiella</i> sp. ZH-08 16S ribosomal RNA gene, partial sequence	1000	1000	99%	0	99%	KC166142.1
<i>Klebsiella</i> sp. T-4 16S ribosomal RNA gene, partial sequence	1000	1000	99%	0	99%	KC109001.1

*Enterobacter* BLAST 16S r RNA sequencing result

Sequences producing significant alignments:						
Sequences producing significant alignments:						
Description	Max score	Total score	Query cover	E value	Max ident	Accession
Enterobacter aerogenes KCTC 2190 strain KCTC 2190 16S ribosomal RNA, complete sequence	990	990	97%	0	99%	AB699730.1
<i>Enterobacter aerogenes</i> strain RB21 16S ribosomal RNA gene, partial sequence	990	990	96%	0	99%	KC431799.1
<i>Enterobacter aerogenes</i> strain ATCC 13048 16S ribosomal RNA gene, partial sequence	990	990	96%	0	99%	KC429778.1
<i>Enterobacter aerogenes</i> EA1509E complete genome	990	7842	96%	0	99%	FO203355.1
<i>Enterobacter</i> sp. B1(2012) 16S ribosomal RNA gene, partial sequence	990	990	96%	0	99%	JQ886667.1
<i>Enterobacter aerogenes</i> strain KNUC5009 16S ribosomal RNA gene, partial sequence	990	990	96%	0	99%	JQ682634.1

Escherichia coli O103:H2 str. 12009 DNA, complete genome  
 Sequence ID: [dbj|AP010958.1](#) Length: 5449314 Number of Matches: 1

Range 1: 408260 to 409120 [GenBank](#) [Graphics](#) ▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1568 bits(849)	0.0	858/862(99%)	1/862(0%)	Plus/Plus

Features: [bacterial alkaline phosphatase PhoA](#)

Query	11	CGGGCTGCTCAGGGCGATATTACTGCACCCGGCGGTGCTCGCCGCTTAACGGGTGATCAT	70
Sbjct	408260	CGGGCTGCTCAGGGCGATATTACTGCACCCGGCGGTGCTCGCCGCTTAACGGGTGATCAT	408319
Query	71	ACCGCCGCTCTGCGTGATTCTCTTAGCGATAAACCTGCAAAAAATATTATTTTGTGATT	130
Sbjct	408320	ACCGCCGCTCTGCGTGATTCTCTTAGCGATAAACCTGCAAAAAATATTATTTTGTGATT	408379
Query	131	GGCGATGGGATGGGGGATTTCGGAAATTACTGCCGCACGTAATTATGCCGAAGGTGCGGGC	190
Sbjct	408380	GGCGATGGGATGGGGGATTTCGGAAATTACTGCCGCACGTAATTATGCCGAAGGTGCGGGC	408439
Query	191	GGCTTTTTTAAAGGTATCGATGCCTTACCGCTTACCGGGCAATACACTCACTATGCGCTG	250
Sbjct	408440	GGCTTTTTTAAAGGTATCGATGCCTTACCGCTTACCGGGCAATACACTCACTATGCGCTG	408499
Query	251	AATaaaaaaaaCTGGCAAACCGGACTACGTACCGACTCGGCTGCATCAGCAACCGCCTGG	310
Sbjct	408500	AATAAAAAAACTGGCAAACCGGACTACGTACCGACTCGGCTGCATCAGCAACCGCCTGG	408559
Query	311	TCAACCGGTGTCAAAAACCTATAACGGCGCGCTGGGCGTCGATATTCACGAAAAAGATCAC	370
Sbjct	408560	TCAACCGGTGTCAAAAACCTATAACGGCGCGCTGGGCGTCGATATTCACGAAAAAGATCAC	408619
Query	371	CCAACGATTCTGGAAATGGCAAAGCCGAGGTCTGGCGACCGGTAACGTTTCTACCGCA	430
Sbjct	408620	CCAACGATTCTGGAAATGGCAAAGCCGAGGTCTGGCGACCGGTAACGTTTCTACCGCA	408679
Query	431	GAGTTGCAGGATGCCACGCCCGCTGCGCTGGTGGCACATGTGACTTCGCGCAAATGCTAC	490
Sbjct	408680	GAGTTGCAGGATGCCACGCCCGCTGCGCTGGTGGCACATGTGACTTCGCGCAAATGCTAC	408739
Query	491	GGTCCGAGCGCGACCAGTGAAAAATGTCCGGGTAACGCTCTGGAAAAAGCGGAAAAAGGA	550
Sbjct	408740	GGTCCGAGCGCGACCAGTGAAAAATGTCCGGGTAACGCTCTGGAAAAAGCGGAAAAAGGA	408799
Query	551	TCGATTACCGAACAGCTGCTTAACGCTCGTGCCGACGTTACGCTTGGCGGCGGGCGCAAAA	610
Sbjct	408800	TCGATTACCGAACAGCTGCTTAACGCTCGTGCCGACGTTACGCTTGGCGGCGGGCGCAAAA	408859
Query	611	ACCTTTGCTGAAACGGCAACCGCTGGTGAATGGCAGGGAAAAACGCTGCGTGAACAGGCA	670
Sbjct	408860	ACCTTTGCTGAAACGGCAACCGCTGGTGAATGGCAGGGAAAAACGCTGCGTGAACAGGCA	408919
Query	671	CAGGCGGTGTTATCAGTTGGTGAACGATGCTGCCTCACTGAATTCGGTGACGGAAGCG	730
Sbjct	408920	CAGGCGGTGTTATCAGTTGGTGAACGATGCTGCCTCACTGAATTCGGTGACGGAAGCG	408979
Query	731	AATCAGCAAAAACCCCTGCTTGGCCTGTTTGTGACGGCAATATGCCAGTGCGCTGGCTA	790
Sbjct	408980	AATCAGCAAAAACCCCTGCTTGGCCTGTTTGTGACGGCAATATGCCAGTGCGCTGGCTA	409039
Query	791	GGGCCGAAAGCAACGTACCACGGCAATATCGATAAGCCCGCAGTCACCTGTACGCCAAAT	850
Sbjct	409040	GGGCCGAAAGCAACGTACCACGGCAATATCGATAAGCCCGCAGTCACCTGTACGCCAAAT	409099
Query	851	CCGCAACGTATGGACCAGTGTa 872	
Sbjct	409100	CCGCAACGTAATGAC-AGTGTA 409120	

**Alignment** of *Escherichia coli* O103:H2 str. 12009 DNA, complete genome

Enterococcus casseliflavus gene for 16S ribosomal RNA, partial sequence, strain: CN1  
 Sequence ID: [dbj|AB699730.1](#) Length: 878 Number of Matches: 1

Range 1: 329 to 872 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
990 bits(536)	0.0	543/546(99%)	2/546(0%)	Plus/Minus
Query 14	GCGGAGTGCTTATGCGTTT	GCTGCAGCACTGAAGGGCGGAAACCCTCCAACACTTAGCAC		73
Sbjct 872	GCGGAGTGCTTATGCGTTT	GCTGCAGCACTGAAGGGCGGAAACCCTCCAACACTTAGCAC		813
Query 74	TCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTCGCTCCCCACGCTTTTCGA			133
Sbjct 812	TCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTCGCTCCCCACGCTTTTCGA			753
Query 134	GCCTCAGCGTCAGTTACAGACCAGAGAGCCGCTTCGCCACTGGTGTCTCTCCATATATC			193
Sbjct 752	GCCTCAGCGTCAGTTACAGACCAGAGAGCCGCTTCGCCACTGGTGTCTCTCCATATATC			693
Query 194	TACGCATTTACCGCTACACATGGAATTCCTCTCTCTCTGCACTCAAGTCTCCCAG			253
Sbjct 692	TACGCATTTACCGCTACACATGGAATTCCTCTCTCTCTGCACTCAAGTCTCCCAG			633
Query 254	TTTCCAATGACCCTCCCCGGTTGAGCCGGGGGCTTTCACATCAGACTTAAGAAACCGCCT			313
Sbjct 632	TTTCCAATGACCCTCCCCGGTTGAGCCGGGGGCTTTCACATCAGACTTAAGAAACCGCCT			573
Query 314	GCGCTCGCTTTACGCCAATAAATCCGGACAACGCTTGCCACCTACGTATTACCGCGGCT			373
Sbjct 572	GCGCTCGCTTTACGCCAATAAATCCGGACAACGCTTGCCACCTACGTATTACCGCGGCT			513
Query 374	GCTGGCACGTAGTTAGCCGTGGCTTTCTGGTTAGATACCGTCAAGGGATGAACATTTTAC			433
Sbjct 512	GCTGGCACGTAGTTAGCCGTGGCTTTCTGGTTAGATACCGTCAAGGGATGAACATTTTAC			453
Query 434	TCTCATCCTTGTCTTCTCTAACAACAGAGTTTTACGATCCGAAAACCTTCTTCACTCAC			493
Sbjct 452	TCTCATCCTTGTCTTCTCTAACAACAGAGTTTTACGATCCGAAAACCTTCTTCACTCAC			393
Query 494	GCGGCGTTGCTCGGTCAGACTTTTCGTCCATTGCCGAAGATTCCCTACTGCTGcccccccG			553
Sbjct 392	GCGGCGTTGCTCGGTCAGACTTTTCGTCCATTGCCGAAGATTCCCTACTGCTGCCTCCC-G			334
Query 554	TAAGGA	559		
Sbjct 333	TA-GGA	329		

**Alignment** of *Enterococcus casseliflavus* gene for 16S ribosomal RNA, partial sequence, strain: CN1



Klebsiella sp. SR-143 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KC455430.1](#) Length: 1440 Number of Matches: 1

Range 1: 316 to 852 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
979 bits(530)	0.0	535/537(99%)	2/537(0%)	Plus/Minus
Query 6	ACT-CCCAGGCGGTTCGATTT-ACGCGTTAGCTCCGGAAGCCACGCCTCAAGGGCACAACC	63		
Sbjct 852	ACTCCCAGGCGGTTCGATTTAACGCGTTAGCTCCGGAAGCCACGCCTCAAGGGCACAACC	793		
Query 64	TCCAAATCGACATCGTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTGTCTCCCA	123		
Sbjct 792	TCCAAATCGACATCGTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTGTCTCCCA	733		
Query 124	CGCTTTCGCACCTGAGCGTCAGTCTTTGTCCAGGGGGCCGCTTCGCCACCGGTATTCCT	183		
Sbjct 732	CGCTTTCGCACCTGAGCGTCAGTCTTTGTCCAGGGGGCCGCTTCGCCACCGGTATTCCT	673		
Query 184	CCAGATCTCTACGCATTTACCGCTACACCTGGAATTCTACCCCTCTACAAGACTCTA	243		
Sbjct 672	CCAGATCTCTACGCATTTACCGCTACACCTGGAATTCTACCCCTCTACAAGACTCTA	613		
Query 244	GCCGTCCAGTTTCGAATGCAGTCCCAGGTTGAGCCCGGGGATTCACATCCGACTTGAC	303		
Sbjct 612	GCCGTCCAGTTTCGAATGCAGTCCCAGGTTGAGCCCGGGGATTCACATCCGACTTGAC	553		
Query 304	AGACCGCTGCGTGCCTTTACGCCAGTAATCCGATTAACGCTTGACCCCTCCGTATT	363		
Sbjct 552	AGACCGCTGCGTGCCTTTACGCCAGTAATCCGATTAACGCTTGACCCCTCCGTATT	493		
Query 364	ACCGCGGCTGCTGGCACGGAGTTAGCCGGTGCCTTCTTCTGCGGTAACGTCAATCGACAA	423		
Sbjct 492	ACCGCGGCTGCTGGCACGGAGTTAGCCGGTGCCTTCTTCTGCGGTAACGTCAATCGACAA	433		
Query 424	GGTTAATAACCTTATCGCCTTCCTCCCCGCTGAAAGTACTTTACAACCGAAGGCCTTCT	483		
Sbjct 432	GGTTAATAACCTTATCGCCTTCCTCCCCGCTGAAAGTACTTTACAACCGAAGGCCTTCT	373		
Query 484	TCATACACGCGCATGGCTGCATCAGGCTTGCGCCATTGTGCAATATCCCCACTG	540		
Sbjct 372	TCATACACGCGCATGGCTGCATCAGGCTTGCGCCATTGTGCAATATCCCCACTG	316		

**Alignment** of *Klebsiella* sp. SR1.6 16S ribosomal RNA gene, partial sequence

Enterobacter aerogenes KCTC 2190 strain KCTC 2190 16S ribosomal RNA, complete sequence  
 Sequence ID: [ref|NR\\_102493.1](#) Length: 1554 Number of Matches: 1

Range 1: 377 to 916 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
990 bits(536)	0.0	539/540(99%)	1/540(0%)	Plus/Plus
Query 9	TGGGCGC-AGCCTGATGCAGCCATGCCGCGTGTATGAAGAAGGCCTTCGGGTTGTAAAGT			67
Sbjct 377	TGGGCGCAAGCCTGATGCAGCCATGCCGCGTGTATGAAGAAGGCCTTCGGGTTGTAAAGT			436
Query 68	ACTTTCAGCGAGGAGGAAGGCGTTAAGGTTAATAACCTTGGCGATTGACGTACTCGCAG			127
Sbjct 437	ACTTTCAGCGAGGAGGAAGGCGTTAAGGTTAATAACCTTGGCGATTGACGTACTCGCAG			496
Query 128	AAGAAGCACCGGCTAACTCCGTGCCAGCAGCCGCGGTAATACGGAGGGTGCAAGCGTTAA			187
Sbjct 497	AAGAAGCACCGGCTAACTCCGTGCCAGCAGCCGCGGTAATACGGAGGGTGCAAGCGTTAA			556
Query 188	TCGGAATTACTGGGCGTAAAGCGCACGCAGGCGGTCTGTCAAGTCGGATGTGAAATCCCC			247
Sbjct 557	TCGGAATTACTGGGCGTAAAGCGCACGCAGGCGGTCTGTCAAGTCGGATGTGAAATCCCC			616
Query 248	GGGCTCAACCTGGGAACTGCATTGCGAACTGGCAGGCTAGAGTCTGTAGAGGGGGGTAG			307
Sbjct 617	GGGCTCAACCTGGGAACTGCATTGCGAACTGGCAGGCTAGAGTCTGTAGAGGGGGGTAG			676
Query 308	AATTCAGGTGTAGCGGTGAAATGCGTAGAGATCTGGAGGAATACCGGTGGCGAAGGCCG			367
Sbjct 677	AATTCAGGTGTAGCGGTGAAATGCGTAGAGATCTGGAGGAATACCGGTGGCGAAGGCCG			736
Query 368	CCCCCTGGACAAAGACTGACGCTCAGGTGCGAAAGCGTGGGGAGCAAACAGGATTAGATA			427
Sbjct 737	CCCCCTGGACAAAGACTGACGCTCAGGTGCGAAAGCGTGGGGAGCAAACAGGATTAGATA			796
Query 428	CCCTGGTAGTCCACGCCGTAACGATGTCGACTTGGAGGTTGTGCCCTTGAGGCGTGGCT			487
Sbjct 797	CCCTGGTAGTCCACGCCGTAACGATGTCGACTTGGAGGTTGTGCCCTTGAGGCGTGGCT			856
Query 488	TCCGGAGCTAACGCGTTAAGTCGACCGCCTGGGGAGTACGGCCGCAAGGTTAAAAC TCA			547
Sbjct 857	TCCGGAGCTAACGCGTTAAGTCGACCGCCTGGGGAGTACGGCCGCAAGGTTAAAAC TCA			916

**Alignment of *Enterobacter aerogenes* KCTC 2190 strain KCTC 2190 16S ribosomal RNA, complete sequence**

## Appendix 6

CHROMagar brochure

General Brochure

**Intense Colours**

*Salmonella*  
*Listeria*

**Cost Efficient**

*ESBL*  
*KPC*

**Fast Results**

*E.coli*  
*E.coli O157*

**Dehydrated Media**

*S.aureus*  
*MRSA*

*VRE*  
*Pseudomonas*

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For Colourful Microbial Detection

**CHROMagar**  
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