

ABSTRACT

Oxidative stress-induced neurodegenerative diseases have become more prevalent lately due to the stressful environment and lifestyle. Growing empirical scientific evidences which support the use of plant-derived antioxidants in the control of neurodegenerative disorders has been validated in the present investigation. *Loranthus parasiticus* (L.) Merr, a chinese traditional folk medicine which has been used in treating brain diseases was selected for the present study. Therefore, *L. parasiticus* was hypothesized to exhibit antioxidative and neuroprotective properties in NG108-15 neuroprotection model. *Loranthus parasiticus* aqueous fraction (LPAF) which showed the highest antioxidative and neuroprotective activities against H₂O₂ among the tested extract and fractions was subjected to a bioassay-guided fractionation and isolation approach to identify the most potent neuroprotective compound. (+)-Catechin was found to be the most potent neuroprotective compound and its underlying mechanisms were evaluated subsequently. (+)-Catechin significantly reduced reactive oxygen species production, phosphatidylserine externalization, mitochondrial membrane potential depolarization, sub-G₁ apoptotic fraction induction, and increased the percentage of cell viability following H₂O₂-induced oxidative stress insult. Moreover, (+)-catechin increased the H₂O₂-induced reduction of SOD and GPx activities. (+)-Catechin also upregulated Bcl-2 and downregulated Bax, resulting in a decreased ratio of Bax/Bcl-2. Interestingly, oxidative stress-induced overexpression of chemokine CCL21 was significantly attenuated by (+)-catechin, indicating a novel role of (+)-catechin in neuroprotection context via the regulation of neuronal chemokine CCL21. Collectively, the present findings have proven our hypothesis and support the use of *L. parasiticus* in managing oxidative stress related neurodegenerative diseases.

ABSTRAK

Penyakit neurodegeneratif akibat daripada tekanan oksidatif persekitaran dan gaya hidup semakin mendapat perhatian pada masa kini. Semakin banyak bukti saintifik empirikal yang menyokong penggunaan faktor antioksida tumbuh-tumbuhan dalam pengawalan neurodegeneratif telah dibabitkan dalam pengesahan ini. *Loranthus parasiticus* (L.) Merr, perubatan tradisional Cina yang telah lama digunakan dalam rawatan penyakit-penyakit otak dipilih untuk kajian ini. Oleh itu, *L. parasiticus* dihipotesiskan mempamerkan sifat-sifat antioksida dan neuroprotektif dalam model perlindungan saraf yang menggunakan NG108-15. Fraksi akueus *L. parasiticus* (LPAF) yang menunjukkan aktiviti antioksida dan neuroprotektif tertinggi di antara ekstrak dan pecahan lain terhadap kesan H₂O₂ dipilih untuk fraksinasi dan isolasi berpandukan bioassai untuk mengenal pasti sebatian yang paling neuroprotektif. (+)-Catechin ditemui sebagai sebatian yang paling neuroprotektif dan mekanisme dasarnya telah dinilai kemudian. (+)-Catechin didapati mengurangkan pembentukan spesies oksigen reaktif, pengeluaran phosphatidylserine, potensi mendepolarisasi membran mitokondrion, menginduksi pecahan apoptotic sub-G₁, dan meningkatkan peratusan daya maju sel selepas induksi tekanan oksidatif oleh H₂O₂. Tambahan pula, (+)-catechin meningkatkan aktiviti SOD dan GPx selepas induksi H₂O₂. (+)-Catechin juga meningkatkan Bcl-2 dan menurunkan Bax, dan mengakibatkan penurunan nisbah Bax/Bcl-2. Keputusan yang menarik telah dijumpai di mana tekanan oksidatif yang meningkatkan pengawalaturan CCL21 chemokine telah dilemahkan oleh (+)-catechin dan ini telah menunjukkan peranan novel (+)-catechin dalam konteks perlindungan saraf melalui pengawalaturan CCL21 chemokine. Secara kolektif, pengesahan saintifik ini telah membuktikan hipotesis kami dan menyokong penggunaan *L. parasiticus* dalam pengurusan penyakit neurodegeneratif yang berkaitan dengan tekanan oksidatif.

ACKNOWLEDGEMENTS

First of all I would like to thank Almighty God for giving me strength, confidence, mercy, and supremacy to have my research project completed along this tough journey.

I have been indebted in the preparation of this thesis to my supervisor, Assoc. Prof. Dr. Habsah Abdul Kadir whose patience and benevolence, as well as her intelligent guidance and research experience, have been invaluable to me. Under her sharp supervision, I have obtained research grants and scholarship awards continually throughout the research project.

The help of the staff particularly Mr Asokan and Ms Ng Swee Yee has directly and indirectly accelerate my research progress during the work. I am very grateful to their responsibilities, kindness, and assistance.

My special appreciation goes to Dr Lee Hong Boon and Mr Lim Siang Hui at CARIF for the use of flow cytometry and Dr Ling Sui Kiong and Ms Tan Hooi Poay for their intellectual guidance in conducting the phytochemical research.

On a special note I would like to thank also to all my friends for their caring and support to make this thesis a success.

I would also like to express my gratitude to all my lab mates for their numerous ideas and practical discussion to accomplish the work.

Outside the lab, despite the geographical distance, my family members are always nearby. I thank my family members and relatives for their continually courage, love, and support over my researching life.

Thank you.

TABLE OF CONTENTS

ABSTRACT	ii
ACKNOWLEDGEMENTS	iv
TABLE OF CONTENTS	vi
LIST OF FIGURES	xii
LIST OF TABLES	xv
LIST OF ABBREVIATIONS	xvi

CHAPTER 1 INTRODUCTION

1.1 Background	1
1.2 Objectives	2
1.2.1 Overall objective	2
1.2.2 Specific objectives	2

CHAPTER 2 LITERATURE REVIEW

2.1 Neurodegeneration	4
2.1.1 Brain and nervous system	5
2.1.2 Reactive oxygen species	6
2.1.3 Oxidative stress	7
2.1.4 Mitochondria in relation to neurodegeneration	9
2.1.5 Apoptosis in neuronal cell death	12
2.1.6 Cell cycle event	15
2.2 Neuroprotection	19
2.2.1 Phytochemicals as neuroprotective agents	22
2.2.2 Catechins	23
2.2.3 The role of intracellular GSH in neuroprotection	25
2.2.4 The role of intracellular antioxidant enzymes in neuroprotection	31

2.2.5 Bcl-2 family members as regulators of apoptosis	33
2.2.5.1 Bcl-2	33
2.2.5.2 Bax	35
2.2.6 The role of neuronal chemokines in neuroprotection	36
2.2.7 Chemokine CCL21	38
2.3 Mistletoes	38
2.3.1 The plant, <i>L. parasiticus</i>	39
2.3.2 Ethnopharmacological uses of <i>L. parasiticus</i>	39
2.3.3 Reported phytochemical and biological activities of <i>L. parasiticus</i>.....	39
2.4 NG108-15 neuroprotection model	40

CHAPTER 3 MATERIALS AND METHODS

3.1 Materials	48
3.1.1 Solvents	48
3.1.2 Cell line	48
3.1.3 Culture medium	48
3.1.4 Reagents and chemicals	48
3.1.5 Biochemical assay kits	49
3.1.6 Oligonucleotides	49
3.1.7 Antibodies	50
3.1.8 Instruments/Equipments	50
3.2 Methods	52
3.2.1 Extraction and fractionation of <i>L. parasiticus</i>	52
3.2.1.1 Collection of <i>L. parasiticus</i>	52
3.2.1.2 Preparation of <i>L. parasiticus</i>	52
3.2.1.3 Solvent extraction and fractionation of <i>L. parasiticus</i>	52

3.2.2 Antioxidant studies	53
3.2.2.1 DPPH free radical scavenging activity assay	53
3.2.2.2 Reducing power activity assay	53
3.2.2.3 Lipid peroxidation inhibitory assay	54
3.2.2.4 Total phenolic content	54
3.2.3 Cell culture	55
3.2.3.1 Maintenance of cells	55
3.2.3.2 Cryopreservation of cells	55
3.2.3.3 Reviving of cells	56
3.2.3.4 Subculturing of cells	56
3.2.3.5 Counting of cells	56
3.2.4 Neuroprotective experimental design	57
3.2.4.1 Induction of oxidative damage	57
3.2.4.2 MTT cell viability assay	57
3.2.5 Bioassay-guided isolation of neuroprotective compounds from LPAF.....	58
3.2.5.1 Fractionation and purification of LPAF	58
3.2.5.2 Thin layer chromatography	59
3.2.5.3 Liquid chromatography mass spectroscopy	60
3.2.5.4 NMR spectroscopy	60
3.2.5.4.1 AC trimer	60
3.2.5.4.2 (+)-Catechin	60
3.2.6 Fluorescence microscopy detection of DAPI nuclear stain	61
3.2.7 Measurement of total intracellular GSH content	61
3.2.8 Determination of intracellular reactive oxygen species using DCF-DA stain	62

3.2.9 Assessment of externalization of phosphatidylserine using annexin V/PI labeling.....	62
3.2.10 Detection of mitochondrial membrane potential by JC-1 stain	62
3.2.11 Analysis of cell cycle events using PI stain	63
3.2.12 SOD enzyme activity assay	64
3.2.13 GPx enzyme activity assay	64
3.2.14 Analysis of Bax, Bcl-2, and CCL21 gene expression via Q-PCR	65
3.2.15 Flow cytometric analysis of Bax, Bcl-2, and CCL21 protein expression by immunofluorescence staining	70
3.2.16 Data analysis	70
 CHAPTER 4 RESULTS	
4.1 Evaluation of antioxidative activities of the extract and fractions of <i>L. parasiticus</i>	71
4.1.1 LPAF showed the strongest DPPH free radical scavenging activity.....	71
4.1.2 LPAF possessed the highest reducing power activity	74
4.1.3 LPAF exhibited the strongest inhibitory activity in lipid peroxidation	74
4.1.4 LPAF yielded the highest phenolic content	77
4.2 Assessment of neuroprotective effect of the extract and fractions of <i>L. parasiticus</i>	77
4.2.1 Effect of H₂O₂-induced oxidative damage in NG108-15 cells	77
4.2.2 LPAF increased the cell viability after H₂O₂ insult	79
4.3 Evaluation of neuroprotective potential of LPAF	79

4.3.1 LPAF inhibited apoptotic nuclear morphological changes	79
4.3.2 LPAF attenuated H₂O₂-induced depletion of GSH	82
4.3.3 LPAF decreased H₂O₂-induced reactive oxygen species formation	82
4.3.4 LPAF mitigated H₂O₂-induced externalization of phosphatidylserine	85
4.3.5 LPAF prevented H₂O₂-induced depolarization of mitochondrial membrane potential	85
4.3.6 LPAF abrogated H₂O₂-induced appearance of subG₁-cells	90
4.4 Isolation of AC trimer and (+)-catechin via bioactivity-guided approach	90
4.4.1 Phytochemical analysis of LPAF	90
4.4.2 (+)-Catechin exerted stronger neuroprotective activity compared to AC trimer by showing a higher precentage cell viability after H₂O₂ insult	97
4.5 Neuroprotective mechanism elucidation on (+)-catechin	97
4.5.1 Effect of (+)-catechin on apoptotic markers	97
4.5.1.1 (+)-Catechin reduced reactive oxygen species formation	97
4.5.1.2 (+)-Catechin alleviated externalization of phosphatidylserine	100
4.5.1.3 (+)-Catechin attenuated dissipation of mitochondrial membrane potential	103
4.5.1.4 (+)-Catechin blocked accumulation of subG₁-cells	106
4.5.2 Effect of (+)-catechin on intracellular antioxidant enzymes	106
4.5.2.1 (+)-Catechin improved SOD enzyme activity	106
4.5.2.2 (+)-Catechin increased GPx enzyme acitivity	110

4.5.3 Effect of (+)-catechin on gene expression analysis	110
4.5.3.1 (+)-Catechin decreased the expression of Bax, increased the expression of Bcl-2, and reduced the ratio of Bax/Bcl-2	110
4.5.3.2 (+)-Catechin attenuated the induction of chemokine CCL21.....	111
4.5.4 Effect of (+)-catechin on protein expression level	118
4.5.4.1 (+)-Catechin downregulated Bax expression and upregulated Bcl-2 expression	118
4.5.4.2 (+)-Catechin downregulated the expression of chemokine CCL21	118
CHAPTER 5 DISCUSSION	121
CHAPTER 6 CONCLUSION	138
BIBLIOGRAPHY	139
APPENDIX	175

LIST OF FIGURES

Figure 1. Free radical and reactive oxygen species formation as shown in equations (Kehrer, 2000)	8
Figure 2. Two major apoptotic signaling routes: extrinsic (death receptor-mediated) pathway and intrinsic (mitochondrial-mediated) pathway (Gomez-Sintes et al., 2011)	16
Figure 3. Phases of cell cycle	20
Figure 4. Chemical structure of (-)-epicatechin	26
Figure 5. Chemical structure of (-)-epicatechin gallate	26
Figure 6. Chemical structure of (-)-epigallocatechin	27
Figure 7. Chemical structure of (-)-epigallocatechin gallate	27
Figure 8. Chemical structure of (+)-catechin	28
Figure 9. Chemical structure of (+)-gallocatechin	28
Figure 10. Outline of main intracellular antioxidant enzymes namely SOD, GPx, and catalase involved in the prevention of reactive oxygen species (Reiter, 1995)	32
Figure 11. Image of <i>Loranthus parasiticus</i>	42
Figure 12. Chemical structure of coriamyrtin	43
Figure 13. Chemical structure of tutin	43
Figure 14. Chemical structure of coriatin	44
Figure 15. Chemical structure of corianin	44
Figure 16. Chemical structure of avicularin	45
Figure 17. Chemical structure of quercetin	45
Figure 18. Chemical structure of quercetin-3-arabinoside	46
Figure 19. Graphical abstract illustrating experimental design	47
Figure 20. Principle of SOD assay kit	66

Figure 21. Principle of GPx assay kit	67
Figure 22. Effects of LPEE, LPEAF, and LPAF on DPPH free radical scavenging activity	72
Figure 23. Effects of LPEE, LPEAF, and LPAF on reducing power activity	75
Figure 24. Effects of LPEE, LPEAF, and LPAF on lipid peroxidation inhibitory activity	76
Figure 25. Effects of H ₂ O ₂ -induced oxidative stress in NG108-15 cells by MTT cell viability assay	78
Figure 26. Neuroprotective effects of LPEE, LPEAF, and LPAF on the viability of NG108-15 cells	80
Figure 27. Morphological apoptotic nuclear analysis of LPAF treated NG108-15 cells by DAPI staining	81
Figure 28. Effect of LPAF against H ₂ O ₂ -induced GSH depletion in NG108-15 cells ..	83
Figure 29. Effect of LPAF on reactive oxygen species formation by H ₂ O ₂ in NG108-15 cells	84
Figure 30. Effect of LPAF against H ₂ O ₂ -induced externalization of phosphatidylserine in NG108-15 cells	87
Figure 31. Effect of LPAF on the change of mitochondrial membrane potential following H ₂ O ₂ insult in NG108-15 cells	89
Figure 32. Effect of LPAF on cell cycle event in NG108-15 cells	92
Figure 33. Flow chart of the bioassay-guided fractionation and isolation of neuroprotective compounds from LPAF	93
Figure 34. AC trimer isolated from <i>L. parasiticus</i>	94
Figure 35. (+)-Catechin isolated from <i>L. parasiticus</i>	94
Figure 36. Effect of (+)-catechin on H ₂ O ₂ -induced reactive oxygen species formation in NG108-15 cells	99

Figure 37. Effect of (+)-catechin on the phosphatidylserine externalization in NG108-15 cells	102
Figure 38. Effect of (+)-catechin on mitochondrial membrane potential in NG108-15 cells	105
Figure 39. Effect of (+)-catechin on cell cycle distribution in NG108-15 cells	108
Figure 40. Effect of (+)-catechin on SOD activity in NG108-15 cells	109
Figure 41. Effect of (+)-catechin on GPx activity in NG108-15 cells	112
Figure 42. Effect of (+)-catechin on the Bcl-2 and Bax gene expression levels in NG108-15 cells	116
Figure 43. Effect of (+)-catechin on chemokine CCL21 gene expression level in NG108-15 cells	117
Figure 44. Effect of (+)-catechin on Bcl-2 and Bax protein expression in NG108-15 cells	119
Figure 45. Effect of (+)-catechin on CCL21 protein expression in NG108-15 cells ..	120

LIST OF TABLES

Table 1. Primer sequence for Bax, Bcl-2, CCL21, and HMBS	51
Table 2. Cycling conditions for Q-PCR	68
Table 3. Reaction mixture for Q-PCR	69
Table 4. Antioxidative properties of <i>L. parasiticus</i> extract and fractions (LPEE, LPEAF, and LPAF) in DPPH free radical scavenging activity, lipid peroxidation inhibitory activity, and total phenolic content quantified by Folin-Ciocalteu reaction	73
Table 5. ^1H -NMR spectroscopic data of AC trimer	95
Table 6. ^{13}C -NMR spectroscopic data of AC trimer	95
Table 7. ^1H -NMR spectroscopic data of (+)-catechin	96
Table 8. ^{13}C -NMR spectroscopic data of (+)-catechin	96
Table 9. Neuroprotective activities of two isolated proanthocyanidins from LPAF namely AC trimer and (+)-catechin following H_2O_2 insult in NG108-15 cells	98
Table 10. Alterations in the activities of SOD and GPx by (+)-catechin and EGCG following H_2O_2 exposure in NG108-15 cells	113

LIST OF ABBREVIATIONS

ATCC	American type culture collection
AIF	Apoptosis-inducing factor
APAF	Apoptosis protease activating factor
BH	Bcl-2 homology
BHT	Butylated hydroxytoluene
BD	Becton Dickinson
CD	Cluster of differentiation
CDK	Cyclin-dependent kinase
DAPI	4',6-diamidino-2-phenylindole
DCH-DA	2,7 dichlorofluorescein diacetate
DMEM	Dulbecco's modified Eagle's medium
DMSO	Dimethyl sulfoxide
DPPH	2,2-diphenyl-1-picrylhydrazyl
DTNB	5',5'-dithio-bis(2-nitrobenzoic acid)
EDTA	Ethylenediaminetetraacetic acid
EGCG	(–)-epigallocatechin-3-gallate
FACS	Fluorescent activated cell sorting
FADD	Fas-associated protein with death domain
FeCl ₃	Ferric chloride
FeSO ₄	Ferrous sulfate
FITC	Fluorescein isothiocyanate
FBS	Fetal bovine serum
GAE/g _{DW}	Gallic acid equivalent per gram of dry weight
GPCR	G-protein coupled receptor

GPx	Glutathione peroxidase
GR	Glutathione reductase
GSH	Glutathione
GSSG	Oxidized glutathione
GST	Glutathione-S-transferase
HAT	Hypoxanthine-aminopterin-thymidine
HMBS	Hydroxymethylbilane synthase
HNE	4-hydroxy-2-nonenal
H ₂ O	Water
H ₂ O ₂	Hydrogen peroxide
H ₂ SO ₄	Sulfuric acid
IC ₅₀	50% inhibitory concentration
IgG	Immunoglobulin G
JC-1	5,5',6,6'-tetrachloro-1,1',3,3'-tetraethylbenzimidazolylcarbocyanine iodide
LC-MS	Liquid chromatography mass spectroscopy
LPAF	<i>Loranthus parasiticus</i> aqueous fraction
LPEAF	<i>Loranthus parasiticus</i> ethyl acetate fraction
LPEE	<i>Loranthus parasiticus</i> ethanol extract
MTT	3-(4,5-domethylthiazol-2-yl)-2,5-diphenyltetrazolium
NADPH	Nicotinamide adenine dinucleotide phosphate
Na ₂ HPO ₄	Disodium hydrogen phosphate
NaH ₂ PO ₄	Sodium dihydrogen phosphate
Na ₂ CO ₃	Sodium carbonate
NG108-15	Mouse neuroblastoma x rat glioma hybrid cell line
NMDA	N-methyl-D-aspartate

NMR	Nuclear magnetic resonance
NO	Nitric oxide
O_2^-	Superoxide anions
OH^\cdot	Hydroxyl radicals
ONOO ⁻	Peroxynitrite
PBS	Phosphate buffer saline
PE	Phycoerythrin
PI	Propidium iodide
Q-PCR	Real time-polymerase chain reaction
SOD	Superoxide dismutase
TBA	Thiobarbituric acid
TCA	Trichloroacetic acid
TLC	Thin layer chromatography
TNF	Tumor necrosis factor
WST-1	2-(4-Iodophenyl)- 3-(4-nitrophenyl)-5-(2,4-disulfophenyl)-2H-tetrazolium, monosodium salt
XO	Xanthine oxidase