

ABSTRACT

Medium-chain-length polyhydroxyalkanoate (mcl-PHA) is biodegradable polyester that gained serious attention recently, mainly because of its versatility to accomodate a wide range of applications especially in biomedical area. Like other biopolymers in PHA family, it is synthesized intracellularly by microorganisms under nutrient stress condition i.e abundant carbon sources than other essential nutrients like nitrogen, oxygen, sulphur, etc. It functions as carbon and energy storage to the cells during depletion of carbon sources. A well known group of bacteria able to specifically produce mcl-PHA is fluorescent pseudomonads belonging to rRNA homology group 1.

To make industrial production of mcl-PHA viable, significant efforts have been put into increasing the yield and productivity of the biopolymer inside the bacteria. Equally important is the efficient extraction method to get them. Currently, an extraction method that is both rapid and non-detrimental to the product i.e. mcl-PHA is not available. In this study, a method to achieve the said goal was investigated through the application of ultrasound-assisted process. In addition, a combination of acetone as solvent and heptane as marginal non-solvent was used as the extraction medium. The effects of volumetric energy dissipation, extraction medium ratio and irradiation time on the extraction process were investigated. Frequency of 37 kHz and heptane as marginal nonsolvent facilitated the process. Following optimization, high PHA extraction rate of $74 \times 10^{-3} \text{ g PHA g}^{-1} \text{ dried biomass min}^{-1}$ was observed at ultrasonic energy output, solvent–marginal nonsolvent ratio and irradiation time of $1151 \pm 3 \text{ J ml}^{-1}$, 50:50 and 5 min respectively.

The effects of exposure duration and ultrasonic power output on mcl-PHA solution in acetone were also studied. Molecular weight and thermal properties of ultrasound-irradiated mcl-PHA was characterized and compared to control (non-

irradiated mcl-PHA). It was found that at constant volumetric acoustic energy dissipation, prolonged exposure of up to 20 minutes caused slightly significant degradation of mcl-PHA. Under ultrasonic irradiation, degradation mechanism of mcl-PHA was proposed to involve random α -chain scission. It proceeds *via* cleavage of ester linkage at the main chain backbone to form alkenyl terminal.

In a related study, mcl-PHA produced by *Pseudomonas putida* Bet001 was used as a blending component with scl-PHA produced by *Delftia tsuruhatensis* Bet002. The thermal stability and film morphology of the blend between the brittle homopolymer polyhydroxybutyrate (scl-PHA) and flexible heteropolymer polyhydroxyoctanoate (mcl-PHA) were studied. The blends were prepared *via* film casting method and analyzed by differential scanning calorimetry (DSC), Field emission scanning electron microscopy (FESEM), X-ray diffraction (XRD) and thermogravimetric analysis (TGA). The blend compositions were varied from 0, 25, 50, and 75 % (w/w) of mcl-PHA. The blends showed immiscibility with a morphology that constitutes of crystallite and amorphous phases. Formation of crystallite was observed from the XRD results. Despite the immiscibility, the thermal stability was improved for all blends.

ABSTRAK

Polihidroksialcanoate berantai sederhana panjang (mcl-PHA) adalah poliester terbiodegradasi yang telah mendapat banyak perhatian kerana keseraboblehannya untuk digunakan di dalam pelbagai aplikasi terutama sekali dalam bidang bioperubatan. Seperti biopoliester lain dalam kumpulan PHA, ianya dapat disintesis oleh mikroorganisma di bawah keadaan ketidakstabilan nutrien iaitu semasa sumber karbon adalah lebih tinggi daripada nutrien penting lain seperti nitrogen, oksigen, sulfur, dan lain-lain. Fungsinya adalah sebagai penyimpanan tenaga untuk sel bakteria semasa susutan sumber karbon. Contoh bakteria yang dapat menghasilkan secara khususnya polimer ini adalah “fluorescent pseudomonads” dari kumpulan rRNA homologi 1.

Untuk meningkatkan produktiviti penghasilan PHA pada tahap industri, banyak kajian telah dilakukan seperti teknologi kultur baketeria. Walaubagaimanapun, kajian pengekstrakan PHA pada masa yang singkat tanpa merosakan polimer tersebut belum lagi dilakukan. Kajian ini sangat penting kerana ianya dapat mengurangkan kos penghasilan PHA. Sehubungan dengan itu, kajian tentang proses pengekstrakan untuk mencapai matlamat tersebut telah dijalankan dengan menggunakan teknologi gelombang ultrabunyi. Di samping itu, campuran aseton (pelarut) dan heptana (bukan pelarut) telah digunakan sebagai medium pengekstrakan. Kesan pelesapan tenaga, nisbah medium pengekstrakan dan masa penyinaran pada proses pengekstrakan telah disiasat. Kekerapan gelombang pada 37 kHz dan campuran heptana sebagai “marginal non-solvent” memudahkan proses tersebut. Optimumnya, kadar pengekstrakan PHA yang tinggi (74×10^{-3} g PHA g⁻¹ dried biomass min⁻¹) diperhatikan apabila tenaga output ultrasonik, nisbah pelarut-bukan pelarut dan masa penyinaran adalah 1151 ± 3 J ml⁻¹, 50:50 dan 5 minit, masing-masing

Kesan jangka masa pendedahan dan tenaga output ultrasonik pada larutan mcl-PHA di dalam aseton juga telah dikaji. Berat molekul dan sifat haba bagi PHA yang didedahkan dengan gelombang bunyi telah dicirikan dan dibanding dengan PHA kawalan (mcl-PHA yang tidak didedahkan). Telah didapati bahawa pendedahan yang berpanjangan sehingga 20 minit boleh menyebabkan degradasi mcl-PHA. Mekanisme pengdegradasian mcl-PHA telah dijangka melibatkan potongan rantai α secara rawak. Ia berlaku melalui proses pemotongan di ikatan ester pada rantai utama untuk menghasilkan hujung rantai yang berakenil .

Di samping itu, mcl-PHA yang dihasilkan oleh *Pseudomonas putida* Bet001 telah digunakan sebagai komponen pengadunan bersama dengan scl-PHA yang dihasilkan oleh *Delftia tsuruhatensis* Bet002. Kestabilan haba dan morfologi filem daripada campuran ini telah dikaji. Campuran ini disediakan dengan cara meruapkan pelarut dari larutan campuran polimer yang pekat dan dianalisis dengan menggunakan DSC, FESEM, XRD dan TGA. Komposisi campuran telah diubah dari 0, 25, 50 , dan 75 % mcl-PHA (w/w). Daripada keputusan FESEM , adunan filem menunjukkan sifat ketidakcampuran dan menunjukkan morfologi yang terdiri daripada gabungan “kristalit” dan fasa “amorphous”. Pembentukan separa kristal telah ditentukan dengan analisis XRD. Walaubagaimanapun, kestabilan haba bagi campuran polimer tersebut bertambah baik.

ACKNOWLEDGEMENT

Bismillahirrahmanirrahim,

In the name of the Almighty the Most Gracious and Most Merciful.

Alhamdulillah, I am very grateful to be one of the members in the Bioprocess and Enzyme Technology Lab, Institute of Biological Science, Faculty of Science. There, I have gained countless knowledge and priceless experience that benefit me in every aspects. I am also grateful to be blessed with fully supportive parents; En. Ishak Omar and Pn. Noor Hamidah Mahmud. The love given throughout the hardships in completing the research are trully priceless.

I would like to thank my main supervisor Associate Professor Dr. Mohamad Suffian Mohamad Annuar for the encouragments and consultations in conducting researches throughout my master study. Thanks to my co-supervisor Associate Professor Dr. Thorsten Heidelberg for helping me in analyzing the chemical aspects of the research. Because of them, I was able to finish my Master study within time as planned from the beginning.

I must also thank my labmates, the members of Bioprocess and Enzyme Technology Laboratory for giving the supports, mentally and physically, in completing this research. Thanks to Nadia, Kak Syairrah, Abg Naziz, Abg Alimin, Chong Boon, Kak Haneen, Kak Faeza, Rafais, Ana, Maryam, Mimi, Pey ling, Ahmad, Haziq, Ikhmal and all other lab members for the endless support given.

Last but no least, thanks to University of Malaya for assisting me financially through grant PV036-2012A to conduct the research and Malaysia government for giving me the MyMaster scholarship to pay the tuition fee.

May God bless you all

TABLE OF CONTENTS

CHAPTER ONE

1.0 INTRODUCTION	1
-------------------------	---

CHAPTER TWO

2.0 LITERATURE REVIEW	7
2.1 History of polyhydroxyalkanoates (PHA)	
2.1.1 (1888-2001)	7
2.1.2 (2001-2013)	8
2.1.3 (2014 onwards – future)	10
2.2 PHA produced by <i>Pseudomonas</i> species	11
2.3 Microbial PHA extraction	12
2.3.1 Solvent extraction	12
2.3.2 Additional marginal non-solvent (co-solvent) in PHA	15
extraction	
2.3.3 Various methods of PHA extraction	15
2.4 Ultrasound as a tool for biomass extraction	18
2.5 RSM as a statistical method to study relationship between parameters	19
2.6 Stability and degradation of PHA	19
2.7 Film morphology of neat PHA and its blend	20

CHAPTER THREE

3.0 MATERIALS AND METHODS	22
3.1 Materials	22
3.1.1 Microorganisms	22
3.1.2 Fatty acids	22
3.1.3 Media	23
3.1.4 Shaker-incubator setup	24
3.1.5 Sterilizer	24
3.1.6 Spectrophotometer	24
3.1.7 Microscope	25
3.1.8 Centrifugation	25
3.1.9 Water bath ultrasonicator	25
3.1.10 Rotary evaporator	25
3.1.11 Analytical instruments	
i. Gas chromatography (GC)	25
ii. ^1H -nuclear magnetic resonance (H-NMR)	26
iii. Fourier transform infrared spectroscopy (FTIR)	26
iv. Thermogravimetric analysis (TGA)	26
v. Differential scanning calorimetry (DSC)	26
vi. Gel permeation chromatography (GPC)	27
vii. Field emission scanning electron micrography (FESEM)	27
viii. X-ray diffraction (XRD)	27
3.1.12 Analytical chemicals	
i. Staining dyes	27
ii. Acidified methanol	28
iii. 3-hydroxyalkanoic methyl ester standards	28

3.2 Methods	29
3.2.1 Strain stock culture	29
3.2.2 Strain maintenance	29
3.2.3 Media preparation	30
3.2.4 Standard calibration curve of <i>P. putida</i> biomass	30
3.2.5 Growth profile of <i>P. putida</i>	31
3.2.6 Standard calibration of methyl 3-hydroxyalkanoate standards and their respective retention time	32
3.2.7 Mcl-polyhydroxyalkanoates (mcl-PHA) production	33
3.2.8 Effects of different fatty acids on mcl-PHA accumulation by <i>P. putida</i>	33
3.2.9 <i>P.putida</i> cells staining	33
3.2.10 Biomass harvesting and removal of fatty acid residue	34
3.2.11 Solvent extraction and purification of PHA	34
3.2.12 PHA methanolysis	35
3.2.13 PHA quantification and monomers identification	36
3.2.14 Ultrasound-assisted PHA extraction	36
3.2.15 Calculation of dissipation energy	38
3.2.16 Ultrasonic-mediated degradation	39
3.2.17 Determination of thermodynamic parameters for octanoic acid derived mcl-PHA	40
3.2.18 Study of film formation of mcl-PHA blends	41

CHAPTER FOUR

4.0 RESULTS AND DISCUSSIONS	42
4.1 <i>P. putida</i> and PHA granules	42
4.2 Direct cell and pure film methanolysis for PHA content determination	43
4.3 Effect of different fatty acids as substrate for mcl-PHA production	44
4.3.1 Total biomass production (final biomass weight)	44
4.3.2 PHA production	45
4.4 PHA extraction	49
4.4.1 Acetone vs chloroform for PHA extraction	49
4.4.2 Effect of time extraction on extraction yield of PHA	50
4.4.3 Effect of sonication frequency on extraction yield of PHA	52
4.4.4 Effect of marginal non-solvent on extraction yield of PHA	53
4.4.5 Optimization of the extraction parameters of mcl-PHA	54
4.4.6 Verification of the predictive model	58
4.4.7 Characterization of ultrasound extracted mcl-PHA	60
4.5 Stability and degradation of mcl-PHA	62
4.5.1 Thermo-kinetic analysis of octanoic acid derived mcl-PHA thermodegradation	62
4.5.2 Thermogravimetric analysis of control and ultrasound irradiated mcl-PHA	65
4.5.3 Differential scanning calorimetry analysis of control and ultrasound-irradiated mcl-PHA	67
4.5.4 Molecular weight analysis of control and ultrasound irradiated mcl-PHA	69

4.5.5 $^1\text{H-NMR}$ spectroscopy	70
4.5.6 FTIR spectroscopy	72
4.5.7 Mechanism of ultrasonic-mediated degradation of mcl-PHA	74
4.6 Thermal properties and film morphology of neat PHA and its blend	76
4.6.1 DSC analysis of scl-mcl PHA blends	76
4.6.2 Thermogravimetric analysis of scl-mcl PHA blends	80
4.6.3 Neat PHA and blend film morphology	80
CHAPTER FIVE	
5.0 CONCLUSIONS	85
BIBLIOGRAPHY	87
APPENDIX A	101
APPENDIX B	102
APPENDIX C	103
APPENDIX D	104
APPENDIX E	105
APPENDIX F	109

LIST OF FIGURES

CHAPTER 1

Figure 1.1	Film of mcl-PHA	5
-------------------	-----------------	---

CHAPTER 2

Figure 2.1	Solid-liquid interphase	14
Figure 2.2	Illustration of physical growth of cavitation bubble	18

CHAPTER 4

Figure 4.1	<i>P. putida</i> cells stained with safranine solution	42
Figure 4.2	PHA granules (stained black) in <i>P. putida</i> cells	42
Figure 4.3	Direct cell methanolysis and extracted PHA methanolysis	43
Figure 4.4	Final dried biomass weight	44
Figure 4.5	Total PHA production in 100 ml of E2 medium culture	45
Figure 4.6	PHA content (%) in total dried biomass	48
Figure 4.7	PHA content (%) as a function of biomass increment	48
Figure 4.8	Conventional PHA extraction (solvent reflux)	49
Figure 4.9	Ultrasound-assisted PHA extraction: (a) 80 kHz and (b) 37 kHz	51
Figure 4.10	Comparison of PHA extraction at two different frequencies	52
Figure 4.11	Comparison of heptane and <i>tert</i> -butanol as marginal non-solvent	53
Figure 4.12	Contour and surface plots showing the effect of ultrasonic Power output (X_1), acetone percentage (X_2), and extraction time (X_3) on the percentage of PHA extraction yield	59

Figure 4.13 ^1H -NMR spectra of (a) control and (b) ultrasound extracted 61

PHA at optimal conditions. (control: non-ultrasound treated
PHA in acetone/heptane 50:50)

Figure 4.14 (a) Thermogram of octanoic acid derived mcl-PHA at different 63

heating rates; (b) Kissinger plot for octanoic acid derived
mcl-PHA ($R^2 = 0.978$).

Figure 4.15 Linear relationships of T_{onset} and T_{final} with heating rate, q 65

Figure 4.16 Relative thermal stability of control PHA and ultrasound 67
irradiated mcl-PHA at different (a) exposure time and
(b) ultrasonic power output

Figure 4.17 DSC thermogram of control PHA and ultrasound irradiated 68

mcl-PHA at different (a) exposure time and (b) ultrasonic power
output

Figure 4.18 ^1H -NNMR spectrum of octanoic acid derived mcl-PHA 71

Figure 4.19 FTIR spectra of control PHA and ultrasound irradiated 73
mcl-PHA for different (a) exposure time and (b) ultrasonic
power output

Figure 4.20 A plausible terminal hydroxyl group dehydration and main 75
chain cleavage mechanism in mcl-PHA degradation during
ultrasound irradiation.

Figure 4.21 (a) DSC analysis - enthalpy peak profile of neat PHAs and 79

their blends upon heating at $10 \text{ } ^\circ\text{C min}^{-1}$; (b) Thermogravimetric
curves of scl/mcl PHA blends and neat polymers

Figure 4.22 (a) Neat PHAs and blend films; FESEM image of (b) neat PHB 83

film (10000 X), (c) neat mcl-PHA film (10000 X), (d) 25 % wt
mcl-PHA blend film (5000 X), (e) 50 % wt mcl-PHA blend film
(5000 X) and (f) 75 % wt mcl-PHA blend film (10000 X)

Figure 4.23 X-ray diffraction (XRD) of scl/mcl PHA blends and neat 84

polymers

LIST OF TABLES

CHAPTER 2

Table 2.1	World wide PHA production and researching companies	7
Table 2.2	PHA in various fields	9
Table 2.3	Various PHA recovery methods	16

CHAPTER 3

Table 3.1	List of saturated fatty acids as carbon substrate (MERCK)	22
Table 3.2	Rich medium	23
Table 3.3	E2 medium	23
Table 3.4	MT solution composition	24
Table 3.5	PHA monomer standards	28
Table 3.6	Composition of glycerol solution	29
Table 3.7	The experimental points in the Box-Behnken design	37
Table 3.8	Relative ultrasonic power	38
Table 3.9	Sonication energy dissipation per unit volume as a function of irradiation time	39

CHAPTER 4

Table 4.1	3-hydroxy monomers composition in mcl-PHA (%) as function of fatty acids substrates	47
Table 4.2	Anova table (solvent reflux of PHA extraction)	50
Table 4.3	Randomized runs of the design combinations and their responses	55
Table 4.4	Comparison between predicted and actual responses	58

Table 4.5	Molecular weight and thermal properties of extracted PHAs (control: non-ultrasound treated PHA in acetone/heptane 50:50)	60
Table 4.6	Thermodynamic parameters for thermodegradation of octanoic acid derived mcl-PHA	64
Table 4.7	Number-average molecular weight (M_n), weight-average molecular weight (M_w) and molecular weight distribution (PDI) of control and ultrasound-irradiated mcl-PHA.	70
Table 4.8	Proton ratio of control and ultrasound-irradiated mcl-PHA	72
Table 4.9	Thermal properties of scl/mcl PHA blends and neat polymers	76

LIST OF SYMBOLS AND ABBREVIATIONS

mcl-PHA	medium chain length polyhydroxyalkanoate
scl-PHA	short chain length polyhydroxyalkanoate
PHB	Polyhydroxybutyrate
PHBV	Polyhydroxybutyrate- <i>co</i> -valerate
PHBHHx	Polyhydroxybutyrate- <i>co</i> -hexanoate
PHO	Polyhydroxyoctanoate
PHHp	Polyhydroxyheptanoate
PHHx	Polyhydroxyhexanoate
PLA	Polylactic acid
DSC	Differential scanning calorimetry
TGA	Thermogravimetric analysis
FESEM	Field emission scanning electron micrography
¹ H-NMR	Proton nuclear magnetic resonance
FTIR	Fourier transform infrared spectroscopy
XRD	X-ray diffraction
GPC	Gel permeation chromatography
GC	Gas chromatography
PTFE	Polytetrafluoroethylene
TMS	Tetramethylsilane
PS	Polystyrene
ANOVA	Analysis of variance
DF	Degree of freedom
SS	Sum of squares
MS	Mean square

<i>F</i>	<i>F</i> -value
<i>P</i>	<i>P</i> -value
<i>P</i>	Power
<i>cp</i>	Specific heat capacity of water
ΔT	Temperature difference
Δt	Time difference
<i>E</i>	Sonication-dissipated energy
<i>V</i>	Volume
<i>M_n</i>	Number-averaged molecular weight
<i>M_w</i>	Weight-averaged molecular weight
<i>PDI</i>	Polydispersity index
<i>q</i>	heating rate (K min ⁻¹)
<i>k</i>	Boltzman constant (1.3807x10 ⁻²³ J K ⁻¹)
<i>h</i>	Planck constant (6.626x10 ⁻³⁴ J s)
<i>E_d</i>	Degradation activation energy (J mol ⁻¹)
<i>R</i>	General gas constant (8.3143 J K ⁻¹ mol ⁻¹)
<i>A</i>	Pre-exponential factor/collision factor (s ⁻¹)
ΔS	Entropy of activation (J K ⁻¹ mol ⁻¹)
<i>T_g</i>	Glass transition temperature
<i>T_c</i>	Crystallization temperature
<i>T_m</i>	Melting temperature
<i>T_p</i>	Peak temperature / most rapid degradation temperature
<i>T_{onset}</i>	Starting temperature of degradation
<i>T_{final}</i>	End temperature of degradation
kPa	kilo Pascal
kDa	kilo Dalton
K	Kelvin

J	Joule
W	watt
min	minute
s	second
ml	mililitre
g	gram
mg	miligram
ppm	part per million
rpm	round per minute
(w/v)	weight per volume
(w/w)	weight per weight
(v/v)	volume per volume

LIST OF APPENDICES

Appendix A	¹ H-NMR spectra of ultrasound irradiated mcl-PHA at different exposure time, as indicated.	107
Appendix B	¹ H-NMR spectra of ultrasound irradiated mcl-PHA at different ultrasonic power output, as indicated.	108
Appendix C	Standard calibration of optical density at 600 nm to dried biomass.	103
Appendix D	Growth profile of <i>Pseudomonas putida</i> .	104
Appendix E	Standard curve of methyl 3-hydroxyalkanoates.	105
Appendix F	Standard calibration of retention time to its corresponding monomer standard.	109